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**Novel Peptidyl Carbamate Inhibitors of the Enzyme Elastase**

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Compounds selected from the group consisting of a compound of the formula

\[ \text{Y} - \text{NH} \]
\[ \text{MeO} - \text{CH} - \text{CH} = \text{CH} - \text{N} - \text{C} = \text{O} - \text{N} - \text{Ph} \]

and compound of the formula

\[ \text{Y} - \text{NH} \]
\[ \text{MeO} - \text{CH} - \text{CH} = \text{CH} - \text{N} - \text{C} = \text{O} - \text{N} - \text{Ph} \]

wherein
- x is 1 or 2,
- Y is carbobenzoxy or benzoyl, and
- XR is

have use as elastase enzyme inhibitors. Particularly potent are the L-proline diastereomers.

Elastase enzyme inhibitory compositions comprise a carrier and an elastase enzyme inhibiting amount of one of the compounds of the invention.

A method of selectively inhibiting the enzyme elastase in an animal or a human in need of such treatment comprises administering to the animal or human an enzyme elastase inhibiting amount of one of the compounds of the invention or a composition thereof.

19 Claims, No Drawings
OTHER PUBLICATIONS


NOVEL PEPTIDYL CARBAMATE INHIBITORS OF THE ENZYME ELASTASE

TECHNICAL FIELD

This invention relates to novel peptidyl carbamate inhibitors of the enzyme elastase. This invention also relates to novel synthetic routes to synthesize the peptidyl carbamates of the invention and to methods of inhibiting the enzyme elastase with the compounds of the invention.

BACKGROUND ART

Proteinases from polymorphonuclear leukocytes and macrophages, especially elastases (human leukocyte elastase and cathepsin G), appear to be responsible for the chronic tissue destruction associated with inflammation, arthritis and emphysema. During infection or inflammation the normal lung is protected from proteolytic digestion by the protease inhibitor α1-antitrypsin. The protective mechanism appears to be non-operative in individuals with an α1-antitrypsin elastase inhibitors capable of replacing α1-antitrypsin therefore appear to be useful in the treatment of pulmonary emphysema and related diseases.

Several types of elastase inhibitors have been reported in the literature. These include peptide chloromethyl ketones as described in “Inhibition of Human Leukocyte Elastase by Peptide Chloromethyl Ketones”, P. M. Tuh and J. C. Powers, FEBS Letters, 50, 359-61 (1975); “Specificity of Porcine Pancreatic Elastase, Human Leukocyte Elastase and Cathepsin G Inhibition with Peptide Chloromethyl Ketones”, J. C. Powers, B. F. Gupton, A. D. Harley, N. Nishino and R. J. Whitley, Biochem. Biophys. Acta. 145, 156-66 (1977); azapeptides “Protease Inhibitors. I. Inhibitors of Elastase”, C. P. Dorn, M. Zimmerman, S. S. Yang, E. C. Yurewicz, B. M. Ashe, R. Frankshun and H. Jones, J. Med. Chem., 20: 1464-68 (1977); “Reaction of Serine Proteases with Aza-amino Acid and Aza-peptide Derivatives”, J. C. Powers and B. F. Gupton, Meth. Enzymol., 46: 208-16 (1977); sulfonyl fluorides “Specificity and Reactivity of Human Leukocyte Elastase, Porcine Pancreatic Elastase, Human Granulocyte Cathepsin G, and Bovine Pancreatic Elastase” (1977). Although some peptide chloromethyl ketones have been shown to be effective in preventing elastase in-reduced emphysema in animal models there is considerable question whether such reactive agents could be used for treating emphysema in humans. (“Prevention of Elastase Induced Experimental Emphysema by Oral Administration of a Synthetic Elastase Inhibitor,” A. Janoff and R. Dearing, Am. J. Respir. Dis., 121: 1025-3 (1980)). This is not surprising since the alkylation moieties in these inhibitors might render them toxic when used on a continuous basis. To be suitable for human use, an enzyme inhibitor has to show a high degree of selectivity and must have minimal toxic side effects. As a result, most drugs are molecules that reversibly bind to specific enzymes or receptor sites. Examples are the carbamate esters physostigmine and neostigmine which have been clinically used as inhibitors of acetyl choline esterases, A. G. Gilman, L. S. Goodman and A. Gilman, “The pharmacological Basis of Therapeutics”, p. 101, MacMillan Publishing Co. (1980).

A series of peptide elastase inhibitors were disclosed in U.S. Pat. No. 4,643,991 to Digenis et al. Another group of polymer-bound elastase inhibitors was disclosed in U.S. application Ser. No. 242,294 by Digenis et al filed on Sept. 9, 1988. There still remains a need in the art for compounds which are superior specific, active-site directed inhibitors of the enzyme elastase without the concomittant detrimental features of other prior art compounds.

SUMMARY OF THE INVENTION

This invention relates to a compound of the formula selected from the group consisting of a compound of the formula

\[ \text{MeO} - \text{O} - \text{N} - \text{H} - \text{C} (\text{CH}_2)_2 - \text{N} - \text{Y} - \text{XR} \]

and a compound of the formula

\[ \text{MeO} - \text{O} - \text{N} - \text{H} - \text{C} (\text{CH}_2)_2 - \text{N} - \text{Y} - \text{XR} \]

wherein

- \( x \) is 1 or 2,
- \( Y \) is carbobenzyo or benzoyl, and
- \( XR = \)
This invention also provides an enzyme elastase inhibitory composition comprising an enzyme elastase inhibitory amount of the compound of the invention, and a carrier.

Also part of the invention is a method of inhibiting the activity of the enzyme elastase comprising adding to an enzyme solution an enzyme elastase inhibitory amount of the compound of the invention.

This invention also relates to a method of selectively inhibiting the activity of the enzyme elastase in the presence of an enzyme selected from the group consisting of trypsin and chymotrypsin comprising adding to an elastase solution an enzyme elastase inhibitory amount of the compound of the invention.

Still part of this invention is a method of selectively inhibiting the enzyme elastase in an animal or human in need of such treatment comprising administering to said animal or human an enzyme elastase inhibiting amount of the compound of the invention.

Other objects, advantages and features of the present invention will become apparent to those skilled in the art from the following discussion.

**BEST MODE FOR CARRYING OUT THE INVENTION**

This invention arose from the desire of providing novel, more potent and selective peptidyl carbamate inhibitors of the enzyme elastase. Thus, the present genus of peptidyl carbamate inhibitors incorporates derivatized lysine and ornithine residues into the P₃ or P₄ positions of the dipeptides, i.e., the compounds of formulas I and II. The present compounds resemble the desmosine cross-linking units in mature elastin, which is the natural substrate of the human leukocyte elastase enzyme.

Thus, this invention provides certain novel substituted peptidyl carbamate compounds, pharmaceutical compositions containing these compounds, and methods for using these pharmaceutical compositions in the selective inhibition of the enzyme elastase without affecting similar serine dependent proteases, e.g., trypsin and chymotrypsin.

It is known from the art that proteases from polymorphonuclear leukocytes and macrophages, especially elastases (human leukocyte HL elastase and cathepsin G) appear to be responsible for the chronic tissue destruction associated with inflammation, arthritis and emphysema. During infection or inflammation, the normal lung is protected from proteolytic digestion by the protease inhibitor, α₁-antitrypsin. This protective mechanism appears to be non-operative in individuals with an α₁-antitrypsin deficiency due to genetic or other causes. Synthetic elastase inhibitors capable of replacing α₁-antitrypsin are therefore useful in the treatment of pulmonary emphysema and related diseases.

According to the present invention, a class of compounds containing the carbamate functionality and oligopeptides have been found to be superior active site directed inhibitors of the enzyme elastase in animals and humans. This class of compounds, therefore, provides an opportunity to incorporate chemical moieties of increased affinity towards the enzyme, and greater capability for the transfer of the acylating moiety to the active site of the enzyme. The nature of the acylating moiety may be varied to optimize the duration of the enzymatic inactivation.

The mechanism of the invention appears to take advantage of the fact that these carbamate esters will react with proteases and esterases at the carbonyl carbon by losing the alkoxy portion and transferring the carbamyl moiety to the active site of the enzyme. Acylation will then lead to recovery of enzymatic activity.

The present invention provides a series of carbamate compounds which are active in accordance with the above proposals as elastase enzyme inhibitors. These novel compounds are carbamates substituted by oligopeptides which are selected from the group consisting of a compound of the formula

\[
\text{MeO} - O - H - N - \text{CH₃}
\]

and a compound of the formula

\[
\text{MeO} - O - H - N - \text{CH₃}
\]

wherein

- x is 1 or 2;
- Y is carbobenzyloxy or benzyloxy; and
- XR =

\[
\begin{align*}
\text{NO}_2 \text{or} & \text{NO}_2 \\
\text{Ph} & \text{Ph}
\end{align*}
\]

In a more preferred and detailed embodiment the compound of the invention is selected from the group consisting of

1. p-Nitrophenyl N-[(Methoxysuccinyl)-L-alanyl-L-alanyl-L-prolylmethyl]-N-isopropylcarbamate,
2. Methyl succinimide succinate
3. t-Butyl Methoxysuccinyl-L-alanine ester,
4. Methoxysuccinyl-L-alanine,
5. Na-Methoxysuccinyl-L-alanyl-Nᵦ-benzoyl-L-lysine,
5

6. N₆-Methoxy succinyl-L-alanyl-N₆-carbobenzoxy-L-lysine phenacyl ester,
7. N₆-Methoxy succinyl-L-alanyl-N₆-carbobenzoxy-L-lysine,
8. N₆-Methoxy succinyl-L-alanyl-N₆-carbobenzoxy-L-ornithine phenacyl ester,
10. N₆-Methoxy succinyl-N₆-carbobenzoxy-L-ornithine,
11. N₆-Methoxy succinyl-N₆-carbobenzoxy-L-ornithyl-L-alanine t-butyl ester,
12. N₆-Methoxy succinyl-N₆-carbobenzoxy-L-ornithyl-L-alanine,
13. N₆-Methoxy succinyl-N₆-carbobenzoxy-L-lysine,
15. N₆-Methoxy succinyl-N₆-carbobenzoxy-L-lysyl-L-alanine,
16. N₆-Methoxy succinyl-N₆-benzoyl-L-lysine,
17. N₆-Methoxy succinyl-N₆-benzoyl-L-lysyl-L-alanine t-butyl ester,
18. N₆-Methoxy succinyl-N₆-benzoyl-L-lysyl-L-alanine,
19. N-Boc-L-prolyl chloromethyl ketone,
20. N-[N-Boc-L-prolyl[methylisopropylamino,  
21. N-{[N-Boc-L-prolyl]methyl]-N-isopropylcarbamate,  
22. p-Nitrophenyl N-(L-prolyl)methyl-N-isopropyl-carbamate hydrochloride,
23. N₆-Methoxy succinyl-L-alanyl-N₆-carbobenzoxy-L-lysyl-D-proline phenacyl ester,
24. N₆-Methoxy succinyl-L-alanyl-N₆-carbobenzoxy-L-lysyl-D-proline phenacyl ester,
25. N₆-Methoxy succinyl-L-alanyl-N₆-carbobenzoxy-L-lysyl-D-proline,
27. N₆-Methoxy succinyl-L-alanyl-N₆-carbobenzoxy-L-lysyl-D-prolyl chloromethyl ketone,
29. N₆-Methoxy succinyl-L-alanyl-N₆-carbobenzyox-L-lysyl-D-propylchloromethyl ketone,
31. N₆-t-Boc-N₆-benzoyl-L-lysine,
32. N₆-t-Boc-N₆-carboxbenzoxy-L-lysine phenacyl ester,
33. N₆-Carboxbenzoxy-L-lysine phenacyl ester hydrochloride,
34. N₆-t-Boc-N₆-carboxbenzoxy-L-ornithine,
35. N₆-t-Boc-N₆-carboxbenzoxy-L-ornithine phenacyl ester,
36. N₆-Carboxbenzoxy-L-ornithine phenacyl ester hydrochloride,
37. N₆-t-Boc-D-proline,  
38. N₆-t-Boc-L-proline,  
39. N₆-t-Boc-D-proline phenacyl ester,
40. N₆-t-Boc-L-proline phenacyl ester,
41. D-Proline phenacyl ester hydrochloride,
42. L-Proline phenacyl ester hydrochloride,
43. N₆-Benzoyl-L-lysine,
44a. p-Nitrophenyl N-{[Methoxy succinyl-(N₆-carbobenzoxy)-L-lysyl-L-alanyl-L-prolyl]methyl]-N-isopropylcarbamate,  
44b. p-Nitrophenyl N-[Methoxy succinyl-(N₆-carbobenzoxy)-L-lysyl-L-alanyl-D-prolyl]methyl]-N-isopropylcarbamate,  
45a. p-Nitrophenyl-N-[Methoxy succinyl-(N₆-benzoyl)-L-lysyl-L-alanyl-L-prolyl]methyl]-N-isopropylcarbamate,  
45b. p-Nitrophenyl-N-[Methoxy succinyl-(N₆-benzoyl)-L-lysyl-L-alanyl-D-prolyl]methyl]-N-isopropylcarbamate,  
46a. p-Nitrophenyl-N-[Methoxy succinyl-(N₆-carbobenzoxy)-L-ornithyry-L-alanyl-L-prolyl]methyl]-N-isopropylcarbamate,  
46b. p-Nitrophenyl-(N₆-carbonyl)-L-ornithyl-L-alanyl-D-prolyl]methyl]-N-isopropylcarbamate,  
47a. p-Nitrophenyl-N-[Methoxy succinyl-(N₆-benzoyl)-L-ornithyl-L-alanyl-L-prolyl]methyl]-N-isopropylcarbamate,  
47b. p-Nitrophenyl-N-[Methoxy succinyl-(N₆-benzoyl)-L-ornithyl-L-alanyl-D-prolyl]methyl]-N-isopropylcarbamate,  
48a. p-Nitrophenyl-N-[Methoxy succinyl-L-alanyl-(N₆-carbobenzoxy)-L-lysyl-L-prolyl]methyl]-N-isopropylcarbamate,  
48b. p-Nitrophenyl N-[Methoxy succinyl-L-alanyl-(N₆-carbonyl)-L-lysyl-L-prolyl]methyl]-N-isopropylcarbamate,  
49a. p-Nitrophenyl N-[Methoxy succinyl-L-alanyl-(N₆-benzoyl)-L-lysyl-L-prolyl]methyl]-N-isopropylcarbamate,  
49b. p-Nitrophenyl N-[Methoxy succinyl-L-alanyl-(N₆-benzoyl)-L-lysyl-L-prolyl]methyl]-N-isopropylcarbamate,  
50a. p-Nitrophenyl N-[Methoxy succinyl-L-alanyl-(N₆-carbonyl)-L-ornithyl-L-prolyl]methyl]-N-isopropylcarbamate,  
50b. p-Nitrophenyl N-[Methoxy succinyl-L-alanyl-(N₆-carbonyl)-L-ornithyl-D-prolyl]methyl]-N-isopropylcarbamate,  
51a. p-Nitrophenyl N-[Methoxy succinyl-L-alanyl-(N₆-benzoyl)-L-ornithyl-L-prolyl]methyl]-N-isopropylcarbamate,  
51b. p-Nitrophenyl N-[Methoxy succinyl-L-alanyl-(N₆-benzoyl)-L-ornithyl-D-prolyl]methyl]-N-isopropylcarbamate,  
52. S-(1-phenyl-5-tetrazol) chloroformate,
53. S-(1-phenyl-5-tetrazol)-N-[N-Boc-L-prolyl]methyl]-N-isopropyl-thiobacarlate,  
54. S-(1-phenyl-5-tetrazolyl-N-propyl]methyl]-N-isopropyl-thio carbamate hydrochloride,
55. S-(1-phenyl-5-tetrazolyl)-N-[methoxy succinyl-alanyl-(N₆-Carboxbenzoxy) lysyl prolyl methyl]-N-isopropyl-thio carbamate, and
56. S-(1-phenyl-5-tetrazolyl)-N-[methoxy succinyl-(N₆-carbonyl) ornithylalanlyl(D-prolyl methyl)]-N-isopropylthio carbamate, and
57. In another particularly preferred embodiment of this invention the elastase enzyme inhibitors of this invention are selected from the group consisting of
1. p-Nitrophenyl N-[Methoxy succinyl]-L-alanyl-L-alanyl-N₆-benzoyl-L-lysine phenacyl ester,
2. Methyl succinimide succinate,
9. \(N_2\)-Methoxy succinyl-L-alanyl-N\(5\)-carbonyl-L-ornithine, 
10. \(N_3\)-Methoxy succinyl-N\(5\)-carbonyl-L-ornithine, 
11. \(N_3\)-Methoxy succinyl-L-alanyl-N\(5\)-carbonyl-L-ornithine, 
12. \(N_3\)-Methoxy succinyl-N\(5\)-carbonyl-L-ornithyl-L-alanine t-butyler, 
13. \(N_3\)-Methoxy succinyl-N\(5\)-carbonyl-L-lysine, 
14. \(N_3\)-Methoxy succinyl-L-carboxy-L-lysyl-L-alanine t-butyler, 
15. \(N_3\)-Methoxy succinyl-N\(5\)-carbonyl-L-lysyl-L-alanine, 
16. \(N_3\)-Methoxy succinyl-N\(5\)-benzoyl-L-lysine, 
17. \(N_3\)-Methoxy succinyl-N\(5\)-benzoyl-L-lysyl-L-alanine t-butyler, 
18. \(N_3\)-Methoxy succinyl-N\(5\)-benzoyl-L-lysyl-L-alanine, 
19. N-BOC-L-prolyl chloromethyl ketone, 
20. N-[N-Boc-L-prolyl]methylisopropylamine, 
21. N-[N-Boc-L-prolyl]methyl-N-isopropyl carbamate, 
22. p-Nitrophenyl N-(L-prolyl)ethyl-N-isopropyl carbamate hydrochloride, 
23. \(N_3\)-Methoxy succinyl-L-alanyl-N\(5\)-carbonyl-L-lysyl-D-proline phenacyl ester, 
24. \(N_3\)-Methoxy succinyl-L-alanyl-N\(5\)-carbonyl-L-lysyl-L-proline phenacyl ester, 
25. \(N_3\)-Methoxy succinyl-L-alkyl-N\(5\)-carbonyl-L-lysyl-D-proline, 
26. \(N_3\)-Methoxy succinyl-L-alanyl-N\(5\)-carbonyl-L-lysyl-D-proline phenacyl ester, 
27. \(N_3\)-Methoxy succinyl-L-alkyl-N\(5\)-carbonyl-L-lysyl-D-prolyl chloromethyl ketone.

In yet another preferred embodiment the enzyme elastase inhibitors of this invention are selected from the group consisting of 
28. \(N_3\)-Methoxy succinyl-L-alanyl-N\(5\)-carbonyl-L-lysyl-L-prolyl chloromethyl ketone, 
29. N-[Methoxy succinyl-L-alanyl-(N\(5\)-carbonyl-L-lysyl-D-prolyl)methyl], N-isopropylamino, 
30. N-[Methoxy succinyl-L-alanyl-(N\(5\)-carbonyl-L-lysyl-D-prolyl)methyl]-N-isopropyl carbamate, 
31. N-t-Boc-N\(5\)-carbonyl-L-lysine, 
32. N-Boc-N\(5\)-carbonyl-L-lysine phenacyl ester, 
33. N\(5\)-Carboxybenzoyl-L-lysine phenacyl ester hydrochloride, 
34. N-t-Boc-N\(5\)-carbonyl-L-ornithine, 
35. N-t-Boc-N\(5\)-carbonyl-L-ornithine phenacyl ester, 
36. N\(5\)-Carboxybenzoyl-L-ornithine phenacyl ester hydrochloride, 
37. N-t-Boc-D-proline, 
38. N-t-Boc-D-proline, 
39. N-t-Boc-D-proline phenacyl ester, 
40. N-t-Boc-D-proline phenacyl ester hydrochloride, 
41. D-Proline phenacyl ester hydrochloride, 
42. L-Proline phenacyl ester hydrochloride, 
43. N\(5\)-Benzyloxycarbonyl-L-lysine, 
44a. p-Nitrophenyl N-[Methoxy succinyl-(N\(5\)-carbonyl-L-lysyl-L-alanyl)D-prolyl methyl]-N-isopropyl carbamate, 
44b. p-Nitrophenyl N-[Methoxy succinyl-(N\(5\)-carbonyl-L-lysyl-L-alanyl-D-prolyl methyl)]N-isopropyl carbamate, 
45a. p-Nitrophenyl N-[Methoxy succinyl-(N\(5\)-benzoyl-L-lysyl-L-alanyl-D-prolyl methyl)]N-isopropyl carbamate,
Two synthetic routes are provided herein to produce the p-nitrophenyl peptidyl carbamates of the invention. One of the synthetic routes is designed to incorporate D or L proline into the P₂ position of the molecule in a stereospecific manner. Derivatives incorporating D-proline, however, possess reduced elastase enzyme inhibitory properties when compared to their diastereomers which incorporate L-proline at the P₂ position.

The peptidyl carbamate inhibitors of the invention selectively inhibit the enzyme elastase, e.g., human leukocyte elastase (HLE) and porcine pancreatic elastase (PPE), with inhibitor dissociation constants ranging from $3 \times 10^{-9}$ M to $2 \times 10^{-8}$ M.

All the peptidyl carbamate inhibitors of the invention have been found to selectively inhibit the enzyme elastase without inhibiting other enzymes such as trypsin or chymotrypsin, among other enzymes.

The peptidyl carbamate inhibitors of the invention are desmosine-like derivatives incorporating L-lysine or L-ornithine residues at the P₃ or P₄ regions of their structures. These features simulate the protruding chains of desmosine cross-linking units in mature elastin.

The present compounds are synthesized in general by methods which are improvements over the method described in U.S. Pat. No. 4,643,991 to Digenis et al, the entire content of which is incorporated herein by reference. The synthetic approach of this invention involves the coupling of the P₆-P₃ moieties of the inhibitor molecules with the P₂-P₁ moieties as depicted in Scheme 1 herebelow.

Scheme 1

General synthesis of desmosine-like peptidyl carbamates. Coupling of P₆-P₃ moiety with the P₂-P₁ moiety

MeOSUC-NHCH(R)C-NHCH(R')CO₂H + HCl,HN → (O)CH₂N-C(O)₂(CH₃)₂

(1) isobutyl chloroformate

-15°C, THF

MeO SUC NHCH(R)C NHCH(R')C

P₆ P₅ P₄ P₃

N C(O) CH₂N-C(O)

CH₂CH₂ CO₂

P₂ P₁ P₁'

This coupling reaction is conducted in a solvent, preferably a polar solvent at low temperature. The conditions for this process are standard in the art and will be known to an artisan.

The synthesis of methoxysuccinyl-alanine aminoacid dipeptides of the invention is depicted in Scheme 2 herebelow.

Scheme 2

Synthesis of the MeOSUC-Ala-Amino acid dipeptides

CH₃OCCH₃CH₃OC-Cl + HON → Et₂N, L-alanine 1-buty] ester

Further reactions involving Et₂N, MeOSUC-ON → THF
The synthesis of methoxysuccinyl-alanine aminoacid dipeptides in which the aminoacid residue can be ornithine (residue 9) or lysine (residue 7) with carbobenzyoxyl (Cbz) or benzoyl (Bz) groups at their terminal amine function (Nε) is shown in Scheme 2. These reactions are conducted under standard conditions which are known to an artisan.

The synthesis of methoxysuccinyl-aminoacid-alanine dipeptides is depicted in Scheme 3 herebelow.

As previously indicated the aminoacid residue could be ornithine or lysine with carbobenzyoxyl or benzoyl at their terminal amino functions (Nε). Examples of these compounds are represented by the structures of Compounds 12, 15 and 18 which are products obtained in accordance with the procedure described in Scheme 3. The coupling of other inhibitor compounds similar to compounds 7 and 9 shown in Scheme 2 with intermediate compound 22 appearing in Scheme 4 results in the synthesis of the compounds 48–51 shown in Table 2 below.
Table 2 provides Ki values for 2 diastereomers of each compound (a and b). In all cases the diastereomers incorporate L proline, where L or D refers to the configuration at the propyla-carbon, exhibit greater inhibitory activity against the enzyme elastase than their corresponding diastereomers derived from D-proline.

The coupling of the intermediate compounds such as compounds 12, 15 and 18 shown in Scheme 3 with 35 compound 22 shown in Scheme 4 herebelow results in the synthesis of compounds 44-47 shown in Table 1 herebelow.

Scheme 4 is shown herebelow and provides the synthesis of Py-P1 moieties as their stable hydrochloride salts (compound 22).
Scheme 4

Scheme 5

Sterespecific synthesis of desmosine-like peptidyl carbamates.

As in the previous cases the conditions for conducting the various steps in this method are standard in the art and would be known to an artisan.

The peptidyl carbamate inhibitor of Formula 46 may be synthesized from all L aminoacids (L-L isomer).
When tested against human leukocyte elastase enzyme functionality at $P_1$ of Formulas I and II by $N$-phenylthiophenotetrazole. This results in compounds PC5 and PC6 which were shown to have $K_i$ values of $2.0 \times 10^{-8}$M to $3 \times 10^{-8}$M respectively.

The synthesis of these compounds are depicted in Scheme 6 below.

Table 3 below shows the effect of the stereochemistry at $P_1$ on the elastase enzyme inhibitory activity of compound 46 of this invention.

The coupling of the intermediate 7 with the compound 54 results in the elastase enzyme inhibitory compound PC5. The coupling of compound 12 with compound 54 results in the elastase enzyme inhibitory compound PC6. The synthesis of these compounds are depicted in Scheme 6 below.

### Table 3

<table>
<thead>
<tr>
<th>Compound</th>
<th>$K_i$ value</th>
<th>$k_{obs}^{act}$</th>
<th>$k_{obs}/[I]^a$</th>
</tr>
</thead>
<tbody>
<tr>
<td>L-L-L</td>
<td>0.65</td>
<td>0.376</td>
<td>8237</td>
</tr>
<tr>
<td>L-L-D-proline</td>
<td>7.95</td>
<td>0.021</td>
<td>461</td>
</tr>
<tr>
<td>L-L-DL-D</td>
<td>0.42</td>
<td>0.437</td>
<td>9587</td>
</tr>
</tbody>
</table>

$^{a}K_i$ value was determined by steady state kinetics.

$^{b}[I] = 7.6 \times 10^{-7}$M, $[E] = 7.6 \times 10^{-8}$M.

$^{c}$First and second order rate constants were determined by pre-steady state inhibition kinetics.

$^{d}$50:50 mixture of two inhibitors, each contributing $3.8 \times 10^{-8}$M.

Extremely highly potent inhibitors of the enzyme elastase are obtained by replacing the p-nitrophenyl proline into the structure of compound 46.

**Scheme 6**

| Synthesis of PC5 and PC6 | Preparation of thiocarbamate portion |
Scheme 6

Synthesis of PC5 and PC6

-continued

preparation of PC5

Preparation of PC6

*CBZ: \(-\text{CH}_2\text{OH}-\)}
The conditions for conducting the various steps encompassed by these methods are known in the art and to an artisan in the field.

As pointed out above, the compounds of the invention may be employed as specific active site directed inhibitors of the enzyme, elastase. For this purpose, the compounds are preferably combined with a pharmaceutically acceptable carrier for administration by injection or in the oral form. Conventional adjuvant and carriers may be employed in combination with about 0.001 to 2.0 weight percent of the active compound. The compounds may be administered to animals or humans at about 10 mg/kg, preferably an average amount of about 6 mg/kg.

The following examples illustrate preferred embodiments of the invention but the invention is not considered to be limited thereto. In the examples and throughout this specification, parts are by weight unless otherwise indicated.

In synthesis of the compounds of the invention, melting points were determined on a Thomas-Hoover Universal Melter apparatus and are uncorrected. 'H NMR spectra were obtained using a Varian EM-360 (60 MHz H2O or EM-390) (90 MHz) spectrometer. Infrared (IR) spectra were recorded on a Perkin-Elmer 567 spectrophotometer. Microanalyses were performed by Atlantic Microlab, Inc., Atlanta, Ga. or by Micro Analysis, Inc., Wilmington, Del.

Reactions were routinely followed by thin layer chromatography (TLC) using Whatman MK6F silica gel plates. Spots were detected by UV (254 nm), iodine or HBr-Ninhydrin spraying. Column chromatography was carried out using Silica Gel 60 from E. Merck, Darmstadt, Germany. All compounds were identified by spectral data and elemental analysis.

Having now generally described this invention, the same will be better understood by reference to certain specific examples, which are included herein for purposes of illustration only and are not intended to be limiting of the invention or any embodiment thereof, unless so specified.

**EXPERIMENTAL**

**EX. 1**  
Methyl succinimide succinate (2)

![Diagram of Methyl Succinimide Succinate](image)

This compound was synthesized by a modified procedure of Digenis et al. A solution of 3-carbethoxypropionyl chloride (5 g, 33.2 mmoles) and N-hydroxysuccinimide (3.8 g, 33.2 mmoles) in ethyl acetate (60 ml) was cooled to 5°C. While stirring, triethylamine (4.3 g, 33.2 mmoles) was slowly added to the cooled solution over a 15 min interval. The mixture was allowed to equilibrate to room temperature (22°C) and react for an additional 3 h. The formed triethylamine salt was filtered, washed with ethyl acetate and the filtrate evaporated under vacuum. The powder was recrystallized from ethyl acetate/hexane to give 5.5 g (24.2 mmoles) (72% yield) of white needles mp 84°–86°C. 'H-NMR (CDCl3) δ 2.44(2H,app.t, J = 8Hz); 2.64(2H,app.t, J = 8Hz); 2.86

**EX. 2**  
t-ButyI Methoxysuccinylalanine ester (3)

![Diagram of t-ButyI Methoxysuccinylalanine ester](image)

To an ice cooled suspension of the activated ester 2 (3.6 g, 16 mmoles) and L-alanine t-butyI ester (2.3 g, 16 mmoles) in THF (50 ml), triethylamine (2.0 g, 16 mmoles) in THF (1 ml) was added dropwise. The progress of the reaction was monitored by TLC (10% methanol in chloroform) and stirred at 5°C for 3.5 h. The precipitate was filtered under vacuum and washed with ethyl acetate. The residual oil, containing the product, was chromatographed using 40 g of silica gel column (2 x 50 cm). Impurities were eliminated by first passing 50 ml of methylene chloride and subsequently compound 3 was eluted with 2% methanol in methylene chloride. Upon evaporation of the eluent solvent under reduced pressure the product was obtained. The latter was then crystallized from ethyl acetate/petroleum ether to give 2.9 g (11.3 mmoles) (71% yield) of a crystalline powder mp 91°–92°C. 'H-NMR (CDCl3) δ 1.36(3H,app.t, J = 8Hz); 1.40(9H,s); 2.44(2H,app.t, J = 8Hz); 2.64(2H,app.t, J = 8Hz); 2.86(3H,s); 4.20–4.60(1H,m); 6.85(1H,m, rotamer of amide —NH)ppm. IR (CDCl3) 3400, 1815, 1785, 1740 cm⁻¹.

**EX. 3**  
Methoxysuccinylalanine (4)

![Diagram of Methoxysuccinylalanine](image)

The t-butyI ester 3 was hydrolyzed by either of the following methods in good yield.

**Procedure A:**

Formic acid (98%, 1.5 ml) was added to an ice cooled solution of 3 (1.0 g, 3.9 mmoles) dissolved in ethyl acetate (10 ml). Hydrogen chloride gas was slowly bubbled through the cooled solution in two short (30 s) intervals 10 min apart. The solution was allowed to warm to room temperature and stirred for 2 h. The volatile liquids were evaporated in vacuo. The residue, containing the product, was chromatographed on 15 g of silica gel column (1 x 25 cm). The product 4 was eluted with 4% methanol in methylene chloride. Upon evaporation of the eluent solvent under reduced pressure 0.77 g (3.8 mmoles) (98% yield) of a transparent oil was obtained.

**Procedure B:**

A solution of 3 (1.0 g, 3.9 mmoles) in glacial acetic acid (5 ml) at room temperature was slowly diluted with 30% HBr in acetic acid (5 ml) and stirred for 30 min. The reaction was stopped by the addition of ice-water (15 ml) and the fine suspension was extracted with methylene chloride (5 x 25 ml). The organic layer was washed with brine, dried (5 g of MgSO4), and the solvent was azeotropically removed under vacuum.
using n-heptane. The product was chromatographically purified as described in Procedure A, to give 0.77 g (3.8 mmoles) (98% yield) of a transparent oil. 1H-NMR (CDCl3) δ 1.36(3Hα,d, J = 8Hz); 2.56(4Hβ,app. t, J = 8Hz); 3.66(3Hβ,s); 4.20-4.60(1Hα,m); 6.56(1Hα,m, rotamer of amide —NH); 9.50(1Hβ,app. ppm IR (CHCl3)

—NH); 9.56(1Hα,app. ppm IR (CHCl3)) 3330, 1730, 1645, 1600, 1540 cm⁻¹.

EX. 5

Nα-Methoxysuccinylalanyl-Nε-carbobenzyloxylysine phenacyl ester (6)

3280, 1735, 1690, 1630, 1540 cm⁻¹.

EX. 4

Nα-Methoxysuccinylalanyl-Nε-benzoyllysine (5)

30 Methoxysuccinylalanine (4) (0.37 g, 1.8 mmoles) and N-hydroxysuccinimide (0.2 g, 1.8 mmoles) were mixed in THF (3 mL) and cooled to 5° C. A concentrated solution of N,N′-dicyclohexylcarbodiimide (0.4 g, 1.8 mmoles) in THF was added dropwise to the cooled solution. The suspension was stirred for 14 h at 5° C, and the precipitated urea formed was filtered under vacuum. The filtrate was cooled to 5° C and used in the next reaction without further purification.

To a mixture of Nε-carbobenzyloxylysine phenacyl ester hydrochloride (35) (0.7 g, 1.7 mmoles) and the above N-hydroxysuccinimide ester in cooled THF (7 mL), triethylamine (0.17 g, 1.7 mmoles) in THF (0.5 mL) was added dropwise. A solution was observed for a short period of time before a precipitate was formed. The progress of the reaction was monitored by TLC (10% methanol in chloroform). Upon completion of the reaction (4 h), the formed triethylamine salt was filtered, washed with ethyl acetate and the filtrate evaporated under vacuum. The residue, containing the product, was chromatographed on 15 g of silica gel column (1x25 cm). Impurities were eliminated by first passing 50 mL of methylene chloride and subsequently the compound was eluted with 2% methanol in methylene chloride. Upon evaporation of the eluent solvent under reduced pressure a hygroscopic product was obtained. The latter was then recrystallized from ethyl acetate/diethyl ether to give 0.8 g (1.3 mmoles) (84% yield) of a crystalline powder, mp 114°-116° C. 1H-NMR (CDCl3) δ 1.36(3Hα,d, J = 8Hz); 1.50-2.20(6Hβ,m); 2.44(2Hα,app. t, J = 8Hz); 2.64(2Hβ,app. t, J = 8Hz); 3.23(2Hα,m); 3.66(3Hβ,s); 4.00-4.70(2Hα,m); 5.80(1Hα,m, rotamer of amide —NH); 7.22(1Hβ,m, rotamer of amide 65 —NH); 7.34(2Hβ,app. dd, Jβ = 8Hz, Jα = 2Hz); 7.43(1Hβ,app. dd, Jβ = 8Hz, Jζ = 2Hz); 7.86(2Hα,app. dd, Jα = 8Hz, Jζ = 2Hz); 8.10(1Hα,m, rotamer of amide —NH); 7.40(5Hα,s); 7.63(3Hβ,m,app. dd, Jα = 8Hz, Jαα = 2Hz); 8.06(2Hα,app. dd, Jα = 8Hz, Jαα = 2Hz)ppm IR (CHCl3)) 3330, 1755, 1730, 1645, 1600, 1540 cm⁻¹.
EX. 6

Na-Methoxysuccinylalanyl-N<sub>ε</sub>-carbobenzoxylysine (7)

Small portions of zinc metal (total 2 g) were added over a 1 h period to a solution of phenacyl ester 6 (0.6 g, 1.1 mmoles) in glacial acetic acid (10 mL). The reaction was completed within one additional hour of stirring at room temperature. The suspension was diluted with 20% methanol in chloroform (50 mL), filtered under vacuum and the precipitate washed with 50 mL of 20% methanol in chloroform. The filtrate, containing the product, was evaporated under reduced pressure. The residue was partially dissolved in 5% methanol in chloroform and filtered to remove the zinc oxide. The second filtrate was evaporated under reduced vacuum to an impure oil. The latter was dissolved in methylene chloride and chromatographed on 10 g of silica gel column (1 × 25 cm). Impurities were eliminated by first passing 100 mL of 2% methanol in methylene chloride and subsequently compound 7 was eluted with 5% methanol in methylene chloride. Upon evaporation of the eluent solvent under reduced pressure an oil with a tendency to foam under vacuum was obtained. The product was crystallized from ethyl acetate/hexane to give 0.37 g (0.74 mmole) (74% yield) of a crystalline white powder, mp 116°-118° C. 1H-NMR (CDCl<sub>3</sub>) δ 1.36(3H<sub>δ</sub>,d, J = 8Hz); 1.40-2.20(6H<sub>α,m</sub>); 2.56(4H<sub>c,c'</sub>,app. t, J = 8Hz); 3.23(2H<sub>δ,d</sub>); 3.66(3H<sub>α</sub>,s); 4.30-4.90(2H<sub>α</sub>,m); 5.10(2H<sub>δ</sub>); 5.60(1H<sub>α</sub>,m, rotamer of amide —NH); 6.80-7.20(2H<sub>δ</sub>,rotamer of amide —NH); 7.34(5H<sub>α</sub>); 9.60(1H<sub>δ</sub>)ppm. IR (CHCl<sub>3</sub>) 3330, 1730, 1710, 1590, 1540 cm<sup>-1</sup>.

EX. 7

Na-Methoxysuccinylalanyl-N<sub>ε</sub>-carbobenzoxyornithine phenacyl ester (8)

The title compound was prepared from N<sub>ε</sub>-carbobenzoxy-L-ornithine phenacyl ester hydrochloride following an analogous procedure to that described for 6. The product was crystallized from ethyl acetate/diethyl ether to give 0.46 g (0.8 mmole) (46% yield) of a crystalline powder mp 112°-113° C. 1H-NMR (CDCl<sub>3</sub>) δ 1.36(3H<sub>δ</sub>,d, J = 8Hz); 1.40-2.20(4H<sub>α,m</sub>); 2.44(2H<sub>c</sub>,app. t, J = 8Hz); 2.64(2H<sub>δ</sub>,app. t, J = 8Hz); 3.18(2H<sub>c</sub>); 3.66(3H<sub>α</sub>); 4.36-5.00(2H<sub>α</sub>,m); 5.10(2H<sub>δ</sub>); 5.46(2H<sub>α</sub>); 5.80(1H<sub>α</sub>,m, rotamer of amide —NH); 6.56(1H<sub>α</sub>,m, rotamer of amide —NH); 7.10(1H<sub>α</sub>,m, rotamer of amide —NH); 7.34(5H<sub>α</sub>); 7.63(3H<sub>α</sub>,m,app. dd, J<sub>α</sub> = 8Hz, J<sub>α,α</sub> = 2Hz); 8.06(2H<sub>c</sub>,app. dd, J<sub>α</sub> = 8Hz), J<sub>α,α</sub> = 2Hz)ppm. IR (CHCl<sub>3</sub>) 3330, 1755, 1730, 1645, 1600, 1540 cm<sup>-1</sup>.

EX. 8

Na-Methoxysuccinylalanyl-N<sub>ε</sub>-carbobenzoxyornithine (9)
This compound was prepared from 8 according to an analogous procedure to that described for 7. Upon evaporation of the eluent solvent under reduced pressure, 0.3 g (0.75 mmole) (68% yield) of a transparent oil was obtained. $^{1}$H-NMR (CDCl$_3$) δ 1.36(3H$_d$,dd, J =8Hz); 1.40-1.90(4H$_b$,m); 2.56(4H$_c$,app. t, J =8Hz); 3.23(2H$_d$,m); 3.66(3H$_e$,s); 4.40-4.80(2H$_f$,m); 5.10(2H$_g$,s); 6.80-7.10(2H$_h$,m, rotamer of amide —NH); 7.34(5H$_i$,s); 10.10(1H$_j$,s)ppm. IR (CHCl$_3$) 3330, 1730, 1710, 1590, 1540 cm$^{-1}$. EX. 9 Na-Methoxysuccinyl-N$_5$-Carbobenzoxyornithine (10)

A suspension of 2 (0.86 g, 3.8 mmole) and N$_5$-carbobenzoxy-L-ornithine (1 g, 3.8 mmole) in DMF were stirred during and after the addition of triethylamine (0.38 g, 3.8 mmole) at room temperature. The progress of the reaction was monitored by TLC (10% acetic acid in ethyl acetate), upon completion of the reaction (24 h), the formed triethylamine salt was filtered and the filtrate coevaporated with toluene under vacuum. The residue, containing the product, was dissolved in a 50:50 mixture of ethyl acetate/0.001M aqueous HCl (30 mL). The organic layer was washed with water (30 mL) and brine (30 mL); dried (5 g of MgSO$_4$) and evaporated under reduced pressure. The residue was chromatographed on 30 g of silica gel column (2×50 cm). The desired product was eluted with 4% methanol in chloroform. Upon evaporation of the eluent solvent under reduced pressure, 1.4 g (3.6 mmole) (96.7% yield) of a oil was obtained. The phenacyl ester derivative was recrystallized from ethyl acetate/hexane to give a crystalline powder mp 112$^\circ$-113$^\circ$ C. $^{1}$H-NMR (CDCl$_3$) δ 1.50-2.00(4H$_d$,m); 2.44(2H$_e$,app. t, J =8Hz); 2.64(2H$_f$,app. t, J =8Hz); 3.23(2H$_g$,m); 3.66(3H$_h$,s); 4.30-4.80(1H$_i$,m); 5.10(2H$_j$,s); 7.20(1H$_k$,m, rotamer of amide —NH); 7.40(5H$_l$,s); 8.13(1H$_m$,s, rotamer of amide —NH); 9.46(1H$_n$,s)ppm. IR (CHCl$_3$) 3320, 1720, 1700, 1660, 1540 cm$^{-1}$. EX. 10 Na-Methoxysuccinyl-N$_5$-carbobenzoxyornithylalanine t-butyl ester (11)

This compound was prepared from 11 according to an analogous procedure to that described for 14. The product (11) was crystallized from ethyl acetate/hexane to give 1.7 g (3.3 mmole) (87% yield) of a white crystalline powder mp 133$^\circ$-135$^\circ$ C. $^{1}$H-NMR (CDCl$_3$) δ 1.36(3H$_d$,dd, J =8Hz); 1.46(9H$_b$,s); 1.50-2.20(4H$_c$,m); 2.44(2H$_d$,app. t, J =8Hz); 2.64(2H$_e$,app. t, J =8Hz); 3.23(2H$_f$,m); 3.66(3H$_g$,s); 4.00-4.52(2H$_h$,m); 5.10(2H$_i$,s); 6.80-7.20(3H$_j$,m, rotamer of amide —NH); 7.40(5H$_k$,s)ppm. IR (CHCl$_3$) 3330, 1735, 1725, 1590, 1540 cm$^{-1}$. EX. 11 Na-Methoxysuccinyl-N$_5$-carbobenzoxyornithylalanine (12)
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1.36(3H, d, J = 8Hz); 1.50-2.20(4H, m); 2.44(2H, app. t, J = 8Hz); 2.64(2H, app. t, J = 8Hz); 3.23(2H, m); 3.66(3H, s); 4.00-4.52(2H, m); 5.10(2H, s); 6.56(1H, m, rotamer of amide —NH); 6.80-7.20(2H, m, rotamer of amide —NH); 7.34(5H, s); 9.36(1H, s) ppm. IR(CHCl₃) 3330, 1730, 1645, 1590, 1540 cm⁻¹.

EX. 12
N₆-Methoxysuccinyl-N₆-carbobenzoxylysine (13).

15 The carboxylic acid function of 13 (3.8 mmole) was activated with N-hydroxysuccinimide (0.4 g, 3.8 mmole) using N,N'-dicyclohexylcarbodiimide (0.8 g, 3.8 mmole) in THF at 0°C. The cold reaction was stirred for one hour and stored in the refrigerator overnight (14 h). The precipitated urea was filtered under vacuum and washed with a small amount of THF.

L-Alanine t-butyl ester (0.7 g, 3.8 mmole) was added as a solid to an ice-cooled THF (7 mL) solution of the activated ester (above). Triethylamine (0.38 g, 3.8 mmole) in THF (0.5 mL) was added dropwise (10 min) to the fine suspension. The precipitated filtrate was filtered after 1 h and the filtrate evaporated under reduced pressure. The residue, containing the product, was chromatographed on 20 g of silica gel column (1 x 50 cm). Impurities were eliminated by first passing 50 mL of methylene chloride and subsequently the compound was eluted with 3% methanol in methylene chloride. Upon evaporation of the eluent solvent under reduced pressure an oil was obtained. The latter was crystallized from ethyl acetate/hexane to give 1.67 g (3.1 mmole) (82% yield from 11 of white crystalline powder, mp 155°-157°C. ¹H-NMR (CDCl₃) δ 1.36(3H, dd, J = 8Hz); 1.46(9H, s); 1.50-2.20(6H, m); 2.44(2H, app. t, J = 8Hz); 2.64(2H, app. t, J = 8Hz); 3.23(2H, m); 3.66(3H, s); 4.00-4.52(2H, m); 5.10(2H, s); 7.20(1H, m, rotamer of amide —NH); 7.40(5H, s); 8.13(1H, m, rotamer of amide—NH); 9.46(1H, s) ppm. IR(CHCl₃) 3320, 1720, 1705, 1660, 1535 cm⁻¹. Elemental analysis cal'd for C₁₃H₁₂N₂O₅: C, 57.87; H, 6.64; N, 7.10. Found: C, 57.87; H, 6.67; N, 7.09.

EX. 13
N₆-Methoxysuccinyl-N₆-carbobenzoxylysylalanine t-butyl ester (14)

10

CH₃O

CH₃(OCH₃)(9)

5

This compound was prepared from N₆-carbobenxoxy-L-lysine according to an analogous procedure to that described for 10. The product was crystallized from chloroform/hexane to give 1.35 g (3.42 mmole) (90% yield) of a crystalline powder, mp 85°-86°C. ¹H-NMR(CDCl₃) δ 1.50-2.00(6H, m); 2.44(2H, app. t, J = 8Hz); 2.64(2H, app. t, J = 8Hz); 3.23(2H, m); 3.66(3H, s); 4.30-4.80(1H, m); 5.10(2H, s); 7.20(1H, m, rotamer of amide —NH); 7.40(5H, s); 8.13(1H, m, rotamer of amide—NH); 9.46(1H, s) ppm. IR(CHCl₃) 3320, 1720, 1705, 1660, 1535 cm⁻¹. Elemental analysis cal'd for C₁₃H₁₁N₂O₅: C, 57.87; H, 6.64; N, 7.10. Found: C, 57.87; H, 6.67; N, 7.09.

EX. 14
N₆-Methoxysuccinyl-N₆-carbobenzoxylysylalanine (15)

10

CH₃O

CH₃(OCH₃)(9)

15

16
EX. 18
Preparation of an ether alcoholic solution of diazomethane, with a Diazaled distillation kit

Ethanol, 95% (8 mL) was added to a solution of potassium hydroxide (1.79 g) in water (2.7 mL) in a 50 mL distilling flask fitted with a dropping funnel and an efficient condenser set downward for distillation. The condenser was connected to two receiving flasks in series, the second containing 20 mL of diethyl ether. The inlet tube of the second receiver was dipped below the surface of the ether. Both receivers were cooled to 0° C. The flask containing the alkali solution was heated in a water bath to 65° C, and a solution of Diazalz (N-methyl-N-nitroso-p-toluenesulphonamide) (7.2 g, 33.0 mmole) in ether (70 mL) was added through the dropping funnel over about 30 min. The rate of distillation was approximately equal to the rate of addition. When the dropping funnel was empty, another 20 mL of ether was added slowly and the distillation was continued until the distilling ether was colorless. The combined ethereal distillate contained about 1 g (33.0 mmole) of diazomethane based on the amount of Diazalz used. Extreme care is warranted during and after the preparation of diazomethane. Care must be taken to avoid possible explosions by thoroughly checking the glassware for cracks and scratches, heating the alkali solution with a water bath (not to exceed 75° C) and keeping the generated diazomethane solutions at or below 5° C.

EX. 19
N-t-Boc-L-prolyl chloromethyl ketone (19)

The procedure of Digenis et al.39 was followed for the preparation of compound 19.

Isobutylchloroformate (2.1 g, 15.3 mmole) was added to a solution of triethylamine (1.55 g, 15.3 mmole) and t-Boc-L-proline (3.3 g, 15.3 mmole) in diethyl ether (30 mL) cooled to -15° C. The reaction mixture was stirred at this temperature for 10 min, at which time a cooled solution (0° C) of diazomethane (55 (1 g, 32.4 mmole) in diethyl ether (200 mL) was added. The vessel, equipped with a calcium sulfate drying tube, was stirred at 0° C in the hood overnight (14 h). The organic layer was washed subsequently with saturated bicharbonate solution (30 mL), water (30 mL) and brine (30 mL). After drying with 15 g of MgSO₄, the solvent was evaporated under reduced pressure to yield of yellow oil. Hydrogen chloride gas was bubbled through an ice cooled solution of the yellow oil (N-t-Boc-L-prolyl azoethyl ketone) in diethyl ether (50 mL) for 60s. The reaction was stopped after 10 min by diluting with cooled diethyl ether (50 mL) and evaporated under vacuum. The residue, containing the product, was chromatographed on 50 g of silica gel column (3 x 75 cm) Impurities were eliminated by first passing 100 mL of 10% hexane in chloroform and subsequently the compound was eluted with chloroform. Upon evaporation of the eluent solvent under reduced pressure a transparent oil was obtained. The latter (19) was then crystallized from chloroform/hexane to give 3.1 g (12.5 mmole) (82% yield) of colorless crystals m.p. 47°-49° C. 1H-NMR (CDCl3) δ 1.46 (9H, s); 1.86-2.16 (4H, m); 3.33-3.76 (2H, m); 4.36 (2H, s); 4.40-4.80 (1H, m) ppm. IR (Nujol) 1740, 1690 cm⁻¹. Elemental Analysis Cal’d. for C₁₁H₁₈CINO₂: C, 53.33; H, 7.32; N, 5.65. Found: C, 53.46; H, 7.35; N, 5.56.

The NMR, IR and Elemental analysis were found to be identical to those reported by Digenis et al.39.

EX. 20
N-{(N-t-Boc-L-propyl)methyl]isopropylamine (20)

This compound was synthesized by a modified procedure of Digenis et al.39.

Isopropylamine (7.4 g, 125.0 mmole) was slowly added to a cooled solution (0° C) of 19 (3.1 g, 12.5 mmole) in diethyl ether and stirred overnight at room temperature (14 h). The salt formed was filtered and the filtrate evaporated under vacuum. The oil was chromatographed on 30 g of silica gel column (2 x 25 cm). Impurities were eliminated by first passing 50 mL of 1% methanol in chloroform and subsequently the compound was eluted with 5% methanol in chloroform. Upon evaporation of the eluent solvent under reduced pressure 3.1 gm (11.4 mmole) (90.8% yield) of an oil was obtained. 1H-NMR (CDCl3) δ 0.93-1.31 (6H, app. d, J = 7Hz); 1.46(9H, s); 1.86-2.16 (4H, m); 2.83 (1H, m); 3.30-3.73 (5H, app. m); 4.30 (1H, app. t, J = 8Hz) ppm. IR (CHCl₃)3330, 1695 cm⁻¹.

The NMR and IR of this compound were found to be identical to those reported by Digenis et al.39.

EX. 21

the procedure of Digenis et al.39 was followed for the preparation of compound 21.
for 2 h, the formed triethylamine salt filtered and the filtrate evaporated under vacuum. The crude oil was dissolved in ethyl acetate and washed with water, 10% aqueous citric acid (30 mL), water (30 mL) and brine (30 mL); dried (10 g of MgSO₄) and evaporated under reduced pressure. The residue, containing the product, was dissolved in a small amount of chloroform and chromatographed on 40 g of silica gel column (2 x 30 cm). The impurities were eliminated by first passing chloroform and subsequently the compound was eluted with 50:1 chloroform in ethyl acetate. Upon evaporation of the eluent solvent under reduced pressure 4.6 g (10.7 mmoles) (94% yield) of a transparent oil was obtained. ¹H-NMR (CDCl₃) δ 1.13 (3H₂,app. d, J = 8Hz); 1.00 (3H₂,add. d, J = 8Hz); 1.06-1.16 (4H₂, m); 3.56-3.84 (2H₂, m); 4.20, 4.32 (2H₂, center of a set of dd, overlapping with another set, J = 20Hz, rotamers of the CH₂ system); 3.96 (2Happ. t, J = 8Hz); 4.53-4.80 (4H₂, m);

IR (Nujol) 1740, 1720, 1600, 1595, 1520 cm⁻¹. The NMR and IR for this compound were found to be identical to those reported by Digenis et al.²⁹

EX. 22
p-Nitrophenyl-N-[Methoxysuccinyl-(N-carbobenzyloxylysylalanyl prolyl)-methyl]-N-isopropylcarbamate (44b, the LLD diastereomer)

To a cooled solution (−15' C.) of 15 (0.85 mmoles) and N-methylmorpholine (0.85 mmoles) in THF (5 mL) isobutylchlororofomate (0.13 g, 0.94 mmoles) in acetonitrile (3 mL) was added. After 10 min, 22 (0.4 g, 1 mmole) was added as a solid and N-methylmorpholine (0.1 g, 1 mmole) as an acetonitrile solution (2 mL). The reaction mixture was allowed to warm to 5° C. in 30 min and stirred (3 h). The reaction mixture was filtered, the filtrate was evaporated under vacuum and the residue was redissolved in methylene chloride (25 mL). The organic solvent was subsequently washed with water (25 mL), 10% aqueous citric acid (25 mL), water (25 mL) and brine (25 mL); dried (5 g of MgSO₄) and evaporated under reduced pressure. The residue, containing 0.99 g (68% yield) of a tan powder, mp 179°-182° C. (lit. mp 190°-193° C.). ¹H-NMR (DMSO-d₆) δ 1.13 (3H₂,app. d, J = 8Hz); 1.20 (3H₂,app. d, J = 8Hz); 1.70-2.30 (4H₂, m); 3.20-3.56 (2H₂, m); 4.20 (2H₂, m); 4.50 (2H₂, m); 7.22 (1Happ. d, J = 10Hz); 7.26 (1Happ. d, J = 10Hz); 8.22 (1Happ. d, J = 10Hz); 8.24 (1Happ. d, J = 10Hz); ppm. IR (Nujol) 1740, 1720, 1610, 1595, 1520 cm⁻¹.
ethyl acetate. The two bands were separately scraped from the plate and extracted with 20% methanol in chloroform (3 x 25 mL). The eluent solvent was evaporated under reduced pressure to give an amorphous powder. The lower band was pure 44a. The desired product (44b) isolated as 0.23 g (0.3 mmol) (35% yield) of a white amorphous powder, mp 46°-47° C., was obtained from the upper TLC band. 1H-NMR (CDCl3) δ 1.13 (3H,app,d, J = 7Hz, rotamer of l); 1.20 (3H,app,d, I = 7Hz, rotamer of 1); 1.36 (3H,app,d, J = 8Hz); 1.40-2.22 (10H, sm); 2.44 (2H,app,t, I = 8Hz); 2.64 (2H,app.t, J = 8Hz); 3.18 (2H, sm); 3.33 (2H,app.t, J = 8Hz); 3.66 (2H,ps); 4.30 (2H, center of 2 sets of dd, J = 20Hz, rotamer of the CH3(N) geminal system); 4.53 (1H,app.t, J = 8Hz); 4.63 (1H,app,t, J = 8Hz); 4.80 (2H,bs, m); 5.10 (2H,sm, s); 5.42 (1H,sm, m); 6.70-7.10 (2H,sm, rotamer of amide —NH); 7.24 (1H,app.d, J = 10Hz, rotamer of 1); 7.28 (1H,app.d, J = 10Hz, rotamers of 1); 7.34 (5H,app.s); 8.20 (1H,app.d, J = 10Hz, rotamer of 1); 8.25 (1H,app.d, J = 10Hz, rotamer of 1); 8.25 (1H,app.d, J = 10Hz, rotamer of 1); 8.25 (1H,app.d, J = 10Hz, rotamer of 1); 8.25 (1H,app.d, J = 10Hz, rotamer of 1); 8.25 (1H,app.d, J = 10Hz, rotamer of 1); 8.25 (1H,app.d, J = 10Hz, rotamer of 1); 8.25 (1H,app.d, J = 10Hz, rotamer of 1); 8.25 (1H,app.d, J = 10Hz, rotamer of 1); 8.25 (1H,app.d, J = 10Hz, rotamer of 1); IR (CHCl3) 3310, 1730, 1650, 1520, 735, 700 cm⁻¹. Elemental analysis cal'd for C38H50N4O12: C, 58.30; H, 6.44; N, 10.74. Found: C, 58.19; H, 6.50; N, 10.64.

EX. 24

p-Nitrophenyl N-[Methoxy succinyl-(N6-carboxbenzoyl)lysylalanyl-prolylmethyl]-N-isopropycarbamate (44a, the LLL diastereomer)

The title compound was prepared from 15 and 22 following a procedure analogous to that described for

EX. 25

p-Nitrophenyl N-[Methoxy succinyl-(N6-benzoyl)lysylalanylprolylmethyl]-N-isopropycarbamate (45b, the LLD diastereomer)
The title compound was prepared from 18 and 22 following a procedure analogous to that described for 44b. The amorphous powder containing 45a and 45b was dissolved in chloroform and applied to a preparative TLC plate (100 mg/mL) and developed two or three times with 15% isopropanol in chloroform. The lower band contained 45a while the upper band contained 45b. The two bands were separately scraped from the plate, extracted with 20% methanol in chloroform (3×25 mL) and eluent solvent evaporated under reduced pressure. The desired product (45b) isolated as 0.26 g (0.34 mmole) (40% yield) of a white amorphous powder mp 63°–64° C. was obtained from the upper TLC band. ¹H-NMR (CDCl₃) δ 1.13 (3H₆, app.d, J = 7Hz, rotamers of I); 1.20 (3H₆, app.d, J = 7 Hz, rotamers of I); 1.36 (3H₂, d, J = 8 Hz); 1.40–2.22 (10H₄, m); 2.44 (2H₄, app.t, J = 8 Hz); 2.64 (2H₄, app.t, J = 8 Hz); 3.18 (2H₄, m); 3.33 (2H₄, app.t, J = 8Hz); 3.66 (2H₄, s); 4.30 (2H₄, center of 2 sets of dd, J = 20Hz, rotamers of the CH₂(10) geminal system); 4.53 (1H₄, app.t, J = 8Hz); 4.63 (1H₄, app.t, J = 8Hz); 4.80 (2H₄, m); 5.42 (1H₄, m); 6.70–7.10 (2H₄, m); 60 rotamers of amide —NH; 7.24 (1H₄, app.d, J = 10Hz, rotamers of I); 7.28 (1H₄, app.d, J = 10Hz, rotamers of I); 7.34 (2H₄, app.d, J_p=8Hz, J_p=2Hz); 7.43 (1H₄, app.d, J_p=8Hz, J_p=2Hz); 7.83 (2H₄, app.d, J_p=8Hz, J_p=2Hz); 8.20 (1H₄, app.d, J = 10Hz, rotamers of I); 8.25 (1H₄, app.d, J = 10Hz, rotamers of I); IR (CHCl₃) 3400, 1730, 1645, 1525, 1440, 1345, 1215, 750, 665 cm⁻¹. Elemental Analysis cal’d for C₁₆H₂₉NO₁₁: C, 58.90; H, 6.37; N, 11.14. Found: C, 58.59; H, 6.76; N, 10.24.

EX. 26
p-Nitrophenyl
N-[Methoxy succinyl-(N₂-benzyl)lysylalaninoprolyl methyl]-N-isopropylcarbamate (45a, the LLL diastereomer)
of 1); 7.28(1H,app.d, J=10Hz, rotamers of 1); 7.34(2H,app.dd, J<sub>qq</sub>=8Hz, J<sub>qq</sub>=2Hz); 7.43(1H,app.dd, J<sub>qq</sub>=8Hz, J<sub>qq</sub>=2Hz); 7.83(2H,app.dd, J<sub>qq</sub>=8Hz, J<sub>qq</sub>=2Hz); 8.20(1H,app.d, J=10Hz, rotamers of 1); 8.25(1H,app.d, J=10Hz, rotamers of 1). IR (CHCl<sub>3</sub>) 3400, 1730, 1645, 1525, 1440, 1345, 1215, 750, 665 cm<sup>-1</sup>. Elemental Analysis cal'd for C<sub>37</sub>H<sub>48</sub>N<sub>9</sub>O<sub>11</sub>: C, 58.90; H, 6.37; N, 11.14. Found: C, 59.18; H, 6.76; N, 11.24.

EX. 27
p-Nitrophenyl
N-[Methoxysuccinyl-(N<sub>6</sub>-carbzenoxy)ornithylanylprolylmethyl]-N-isopropylcarbamate (46b, the LLD diastereomer)

The title compound was prepared from 12 and 22 following a procedure analogous to that described for 44b. The amorphous powder containing 46a and 46b was dissolved in chloroform and applied to a preparative TLC plate (100 mg/plate) and developed two or three times with 10% isopropanol in chloroform. The lower band contained 46a while the upper band contained 46b. The two bands were separately scraped from the plate, extracted with 20% methanol in chloroform (3 x 25 mL) and eluent solvent evaporated under reduced pressure. The desired product (46b) isolated as 0.08 g (0.13 mmoles) (15% yield) of a white amorphous powder, mp 50°-51° C., was obtained from the upper TLC band. <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ 1.13(3H,app.d, J=7Hz, rotamers of 1); 1.20(3H,app.d, J=7Hz, rotamers of 1); 1.36(3H,app.d, J=8Hz); 1.40-2.22(8H,m); 2.44(2H,app.t, J=8Hz); 2.64(2H,app.t, J=8Hz); 3.18(2H,m); 3.33(2H,app.t, J=8Hz); 3.66(2H,s); 4.30(2H, center of 2 sets of dd, J=20Hz, rotamers of the CH<sub>2</sub>(a) geminal system); 4.53(1H,app.t, J=8Hz); 4.63(1H,app.t, J=8Hz); 4.80(2H,app.m); 5.10(2H,m); 5.42(1H,m); 6.70-7.10(2H,m, rotamers of amide -NH); 7.24(1H,app.d, J=10Hz, rotamers of 1); 7.28(1H,p, app.d, J=10Hz, rotamers of 1); 7.34(5H,app.s); 8.20(1H,app.d, J=10Hz, rotamers of 1);

EX. 28
p-Nitrophenyl
N-[Methoxysuccinyl-(N<sub>6</sub>-carbzenoxy)ornithylanylprolylmethyl]-N-isopropylcarbamate (46a, the LLD diastereomer)
The title compound was prepared from 12 and 22 following a procedure analogous to that described for 44b. The amorphous powder containing 46a and 46b was dissolved in chloroform and applied to a preparative TLC plate (100 mg/plate) and developed two or three times with 10% isopropanol in chloroform. The lower band contained 46a while the upper band contained 46b. The two bands were separately scraped from the plate, extracted with 20% methanol in chloroform (3 × 25 mL) and eluent solvent evaporated under reduced pressure. The desired product (46a) isolated as 0.31 g (0.41 mmole) (48% yield) of a white amorphous powder, mp 55°-56°C, was obtained from the lower TLC band. 1H-NMR (CDCl3) δ 1.13(3H,app. d, J = 7Hz, rotamer of 1); 1.20(3H,app. d, J = 7Hz, rotamer of 1); 1.36(3H,app. d, J = 8Hz); 1.40-2.22(8H,M); 2.44(2H,app. t, J = 8Hz); 2.64(2H,app. t, J = 8Hz); 3.18(2H,m); 3.33(2H,app. t, J = 8Hz); 3.66(2H,s); 4.30(2H,a, center of 2 sets of dd, J = 20Hz, rotamers of the CH2(a) geminal system); 4.53(1H,app. t, J = 8Hz); 4.63(1H,app. t, J = 8Hz); 4.80(2H,app. m); 5.10(2H,m,b); 5.42(1H,app. m); 6.70-7.10(2H,m, rotamers of amide —NH); 7.24(1H,app. d, J = 10 Hz, rotamers of 1); 7.28(1H,app. d, J = 10Hz, rotamers of 1); 7.34(5H,app. s); 8.20(1H,app. d, J = 10Hz, rotamers of 1); 8.25(1H,app. d, J = 10Hz, rotamers of 1)ppm. IR 65 (CHCl3) 3310, 1730, 1650, 1520, 735, 700 cm⁻¹. Elemental Analysis cal’d for C27H44N6O12: C, 57.80; H, 6.25; N, 10.94. Found: C, 57.81; H, 6.36; N, 10.68.

EX. 29
p-Nitrophenyl
N-[Methoxy succinyl-[(N5-benzoyl)ornithylalanyl]prolyl methyl]-N-isopropyl carbamate (47, the LLD diastereomer)

A solution of the diastereomers 46a and 46b (0.16 g, 0.2 mmole) in glacial acetic acid (0.3 mL) was diluted with 30% hydrogen bromide in acetic acid (0.3 mL) and stirred at room temperature for 90 min. The reaction was stopped by the addition of dry diethyl ether (10 mL) and the suspension decanted 4 times to produce a hygroscopic brown powder. The brown powder was dried under a flow of nitrogen and dissolved in acetonitrile (2 mL).

Carbonyldiimidazole (0.09 g, 0.6 mmole) was added to a stirred solution of benzoic acid (0.07 g, 0.6 mmole) in acetonitrile (1 mL), and stirring continued for 10 min at room temperature. This solution was then added dropwise to a solution N-methylmorpholine (0.06 g, 0.8 mmole) and the hydrobromide salt described above. The mixture was stirred for 24 h at room temperature. The reaction mixture was evaporated under vacuum and the residue was dissolved in methylene chloride (20 mL). The solution was washed with water (20 mL), 10% aqueous citric acid solution (20 mL), water (20 mL), brine (20 mL), dried (2 g of MgSO4) and evaporated under reduced pressure. The residue, containing 47a and 47b, was applied to a preparative TLC plate and developed two or three times with 15% isopropanol in chloroform. The lower band contained 47a while the upper band contained 47b. The two bands were separately scraped from the plate, extracted with 20%...
methanol in chloroform (3×25 mL) and eluent solvent evaporated under reduced pressure. The desired product (47b) isolated as 0.04 g (0.05 mmol) (25% yield) of a white amorphous powder, mp 65°-66° C., was obtained from the upper TLC band. 1H-NMR (CDCl3) δ 1.13(3H,app.d, J = 7Hz, rotamers of l); 1.20(3H,app.d, J = 7Hz); 1.40-2.22(8H,m); 2.44(2H,app.t, J = 8Hz); 2.64(2H,app.t, J = 8Hz); 3.18(2H,m); 3.33(2H,app.t, J = 8Hz); 3.66(2H,s); 4.30(2H,center of 2 sets of dd, J = 20Hz, rotamers of the CH2(α) geminal system); 4.53(1H,app.t, J = 8Hz); 4.63(1H,app.t, J = 8Hz); 4.80(2H,app.m); 5.42(1H,m); 6.70-7.10(2H,m, rotamers of amide –NH); 7.24(1H,app.d, J = 10Hz, rotamers of l); 7.28(1H,app.d, J = 10Hz, rotamers of l); 7.34(2H,app.d, J = 8Hz, J = 2Hz); 7.43(1H,app.d, J = 8Hz, J = 2Hz); 7.83(2H,app.d, J = 8Hz, J = 2Hz); 8.20(1H,app.d, J = 10Hz, rotamers of l); 8.25(1H,app.d, J = 10Hz, rotamers of l); 5.42(1H,m); 6.70-7.10(2H,m, rotamers of amide –NH); 7.24(1H,app.d, J = 10Hz, rotamers of l); 7.28(1H,app.d, J = 10Hz, rotamers of l); 7.34(2H,app.d, J = 8Hz, J = 2Hz); 7.43(1H,app.d, J = 8Hz, J = 2Hz); 7.83(2H,app.d, J = 8Hz, J = 2Hz); 8.20(1H,app.d, J = 10Hz, rotamers of l); 8.25(1H,app.d, J = 10Hz, rotamers of l). Elemental Analysis cal’d for C36H46N2O11: C, 58.50; H, 6.20; N, 11.38. Found: C, 58.53; H, 6.37; N, 11.31.

EX. 30
p-Nitrophenyl
N-[Methoxysuccinyl-(N2-benzoyl)ornithylalanylpropylmethyl]-N-isopropyl carbamate (47a, the LLL diastereomer)

The title compound was prepared from 46a and 46b following a procedure analogous to that described for 47b. The residue, containing 47a and 47b, was applied to a preparative TLC plate and developed two or three times with 15% isopropanol in chloroform. The lower band contained 47a while the upper band contained 47b. The two bands were separately scraped from the plate, extracted with 20% methanol in chloroform (3×25 mL) and eluent solvent evaporated under reduced pressure. The desired product (47a) isolated as 0.06 g (0.09 mmol) (43% yield) of a white amorphous powder, mp 55°-56° C., was obtained from the lower TLC band. 1H-NMR (CDCl3) δ 1.06-1.28(6H,app.m, rotamer of l); 1.32(3H,app.d, J = 8Hz); 1.38-2.32(8H,m); 2.46(2H,app.t, J = 8Hz); 2.64(2H,app.t, J = 8Hz); 3.20(2H,m); 3.53-3.88(2H,m); 3.66(2H,s); 4.20,4.32(2H,center of a set of dd, overlapping with another set, J = 20Hz, rotamers of CH2(α) geminal system); 4.53(1H,app.t, J = 8Hz); 4.63(1H,app.t, J = 8Hz); 4.80(2H,app.m); 5.42(1H,m); 6.70-7.10(2H,m, rotamers of amide –NH); 7.24(1H,app.d, J = 10Hz, rotamers of l); 7.28(1H,app.d, J = 10Hz, rotamers of l); 7.34(2H,app.d, J = 8Hz, J = 2Hz); 7.43(1H,app.d, J = 8Hz, J = 2Hz); 7.83(2H,app.d, J = 8Hz, J = 2Hz); 8.20(1H,app.d, J = 10Hz, rotamers of l); 8.25(1H,app.d, J = 10Hz, rotamers of l). Elemental Analysis cal’d for C36H46N2O11: C, 58.50; H, 6.20; N, 11.38. Found: C, 58.54; H, 6.36; N, 11.18.

EX. 31
p-Nitrophenyl
N-[Methoxysuccinylalanyl-(Nε-carbobenzoxy)lysylpropylmethyl]-N-isopropyl carbamate (48b, the LLD diastereomer)
The title compound was prepared from 7 and 22 following a procedure analogous to that described for 44b. The amorphous powder containing 48a and 48b was dissolved in chloroform and applied to a preparative 20 TLC plate (100 mg/plate) and developed two or three times with 4% methanol in ethyl acetate. The upper band contained 48a while the lower band contained 48b.

The two bands were separately scraped from the plate, extracted with 20% methanol in chloroform (3×25 mL) and eluent solvent evaporated under reduced pressure. The desired product (48b) isolated as 0.24 g (0.31 mmoles) (36% yield) of a white amorphous powder, mp 44°-45°C, was obtained from the upper TLC band. 1H-NMR (CDCl3) δ 1.13(3H, app.d, J=7Hz, rotamer of l); 1.20(3H, app.d, J=7Hz, rotamer of l); 1.36(3H, d, J=8Hz); 1.40-2.22(10H, m); 2.44(2H, app.t, J=8Hz); 2.64(2H, app.t, J=8Hz); 3.18(2H, m); 3.56-3.84(2H, m); 3.66(3H, s); 4.23(2H, center of 2 sets of dd, J=20Hz, rotamers of the CH2(α) geminal system); 4.34-4.58(4H, m); 4.60-4.64(1H, m); 5.10(2H, s); 6.34-7.16(2H, m, rotamer of amide --NH--); 7.22(1H, app.d, J=10Hz, rotamers of l); 7.26(1H, app.d, J=10Hz, rotamers of l); 8.22(1H, app.d, J=10Hz, rotamers of l); 8.24(1H, app.d, J=10Hz, rotamers of l); 7.34(5H, app.s) ppm. IR (CHCl3) 3305, 1720, 1645, 1520, 735, 700 cm⁻¹. Elemental Analysis cal’d for C36H40N6O12: C, 58.30; H, 6.40; N, 10.70. Found: C, 58.49; H, 6.47; N, 10.63.

EX 32
p-Nitrophenyl
N-[Methoxyssucinyl-(Nα-carbobenzoxy) lysylprolyl methyl]-N-isopropylcarbamate (48a, the LLL diastereomer)
5.008,245

5.10(2H$_3$s); 5.42(1H$_m$,m); 6.36–7.08(2H$_o$,m, rotamers of amide —NH); 7.24(1H$_o$,app,d, J = 10Hz, rotamers of l); 7.28(1H$_o$,app,d, J = 10Hz, rotamers of l); 7.35(5H$_p$,app,s); 8.20(1H$_o$,app,d, J = 10Hz, rotamers of l); 8.25(1H$_o$,app,d, J = 10Hz, rotamers of l) ppm. IR (CHCl$_3$) 3305, 1720, 1645; 1520, 735, 700 cm$^{-1}$. Elemental Analysis cal’d for C$_{38}$H$_{52}$N$_2$O$_7$: C, 58.30; H, 6.40; N, 10.70. Found: C, 58.12; H, 6.55; N, 10.64.

EX. 33
p-Nitrophenyl N-[Methoxysuccinyl]alanyl-(N$_7$-benzoyl)lysylprolyl-methyl]-N-isopropylcarbamate (49b, the LLD diastereomer)

0.29 g (0.39 mmoles) (46% yield) of a white amorphous powder, mp 54°–55° C, was obtained from the lower TLC band. 1H-NMR (CDCl$_3$) δ 1.13(3H$_o$,app,d, J = 7Hz, rotamer of l); 1.20(3H$_o$,app,d, J = 7Hz, rotamer of l); 1.36(3H$_o$,d, J = 8Hz); 1.40–2.22(10H$_m$,m); 2.44(2H$_d$,app,t, J = 8Hz); 2.64(2H$_d$,app,t, J = 8Hz); 3.18(2H$_m$,m); 3.56–3.84(2H$_m$,m); 3.66(3H$_s$,s); 4.23(2H$_o$, center of 2 sets of dd, J = 20Hz, rotamers of the CH$_2$(b) geminal system); 4.34–4.58(4H$_m$,m); 4.60–4.64(1H$_m$,m); 6.34–7.16(2H$_m$,m, rotamer of amide —NH); 7.22(1H$_o$,app,d, J = 10Hz, rotamers of l); 7.26(1H$_o$,app,d, J = 10Hz, rotamers of l); 7.34(2H$_o$,app,d, J$_{oc}$=8Hz, J$_{oc}$=2Hz); 7.43(1H$_o$,app,d, J$_{oc}$=8Hz); 7.83(2H$_o$,app,d, J$_{oc}$=8Hz, J$_{oc}$=2Hz); 8.22(1H$_o$,app,d, J = 10Hz, rotamers of l) ppm. IR (CHCl$_3$) 3340, 1735, 1645, 1520, 1435, 1345, 1260, 735, 700 cm$^{-1}$. Elemental Analysis cal’d for C$_{37}$H$_{54}$N$_2$O$_7$: C, 58.92; H, 6.37; N, 11.14. Found: C, 58.56; H, 6.47; N, 11.30.

The title compound was prepared from 5 and 22 following a procedure analogous to that described for 44b. The amorphous powder containing 49a and 49b 60 was dissolved in chloroform and applied to a preparative TLC plate (100 mg/plate) and developed two or three times with 15% isopropanol in chloroform. The upper band contained 49a while the lower band contained 49b. The two bands were separately scraped from the plate, extracted with 20% methanol in chloroform (3×25 mL) and eluent solvent evaporated under reduced pressure. The desired product (49b) isolated as 8.24(1H$_o$,app,d, J = 10Hz, rotamers of l) ppm. IR (CHCl$_3$) 3340, 1735, 1645, 1520, 1435, 1345, 1260, 735, 700 cm$^{-1}$. Elemental Analysis cal’d for C$_{37}$H$_{54}$N$_2$O$_7$: C, 58.92; H, 6.37; N, 11.14. Found: C, 58.56; H, 6.47; N, 11.30.
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EX. 34
p-Nitrophenyl
N-[Methoxysuccinyl]alanyl-[(N-β-benzoyl)lysylpropylmethy]-N-isopropylcarbamate (49a, the LLL
diastereomer)

The title compound was prepared from 5 and 22 following a procedure analogous to that described for
44b. The amorphous powder containing 49a and 49b was dissolved in chloroform and applied to a prepara-
tive TLC plate (100 mg/plate) and developed two or three times with 15% isopropanol in chloroform. The
upper band contained 49a while the lower band contained 49b. The two bands were separately scraped
from the plate, extracted with 20% methanol in chloroform (3 x 25 mL) and eluent solvent evaporated under
reduced pressure. The desired product (49a) isolated as 0.29 g (0.38 mmole) (45% yield) of a white amorphous
powder, mp 61°-62° C., was obtained from the upper TLC band. 1H-NMR (CDCl3) δ 1.06-1.28 (6H,6,7,9,
30 7.34(2H,9,app.d, J = 8Hz, Jpp = 2Hz); 7.43(1H,app.d, J = 8Hz); 7.83(2H,app.d, J = 8Hz, Jpp = 2Hz); 8.20(1H,app.
d, J = 10Hz, rotamers of I); 8.25(1H,app.d, J = 10Hz, rotamers of I); 1345, 1345, 1260, 735, 700 cm−1. Elemental Analysis

EX. 35
p-Nitrophenyl
N-[Methoxysuccinylalanyl-[(N-β-carbobenzoxy)ornithylpropylmethyl]-N-isopropylcarbamate (50b, the LLD
diastereomer)

rotamers of I); 1.32(3H,d, J = 8Hz); 1.38-2.32(10H,m); 2.46(2H,app.t, J = 8Hz); 2.64(2H,app.t, J = 8Hz);
3.20(2H,m); 3.52-3.88(2H,m); 3.66(3H,s); 5.008,245

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4.20-4.32(2Hb, center of a set of dd, overlapping with
another set, J = 20Hz, rotamer of the CH3(6) geminal
system); 4.50(2H,app.t, J = 8Hz); 4.50-4.80(2H,d,m);
5.42(1H,m); 6.36-7.08(2H,m, rotamers of amide
5 —NH); 7.24(1H,app.d, J = 10Hz, rotamers of I);
7.28(1H,app.d, J = 10Hz, rotamers of I);
44b. The amorphous powder containing 50a and 50b was dissolved in chloroform and applied to a preparative TLC plate (100 mg/plate) and developed two or three times with 4% methanol in ethyl acetate. The upper band contained 50a while the lower band contained 50b. The two bands were separately scraped from the plate, extracted with 20% methanol in chloroform (3 x 25 mL) and eluent solvent evaporated under reduced pressure. The desired product (50b) isolated as 0.1 g (0.13 mmoles) (15% yield) of a white amorphous powder, mp 65°-66°C, was obtained from the lower TLC band. 1H-NMR (CDCl3) δ 1.13 (3H,app.d, J = 7Hz, rotamer of l); 1.20 (3H,app.d, J = 7Hz, rotamer of l); 1.36 (3H,app.d, J = 8Hz); 1.40-2.22 (8H,m); 2.44 (2H,app.t, J = 8Hz); 2.64 (2H,app.t, J = 8Hz); 3.18 (2H,m); 3.56-3.84 (2H,m); 3.66 (3H,s); 4.23 (2H, center of 2 sets of dd, J = 20Hz, rotamers of the CH2(2)geminal system); 4.34-4.58 (4H,app.m); 4.60-4.64 (1H,app.m); 5.10 (2H,s); 6.34-7.16 (2H,app.m, rotamer of amide -NH); 7.22 (1H,app.d, J = 10Hz, rotamers of l); 7.26 (1H,app.d, J = 10Hz, rotamers of l); 8.22 (1H,app.d, J = 10Hz, rotamers of l); 8.24 (1H,app.d, J = 10Hz, rotamers of l); 7.34 (5H,app.s) ppm. IR (CHCl3) 3305, 1720, 1645, 1520, 735, 700 cm⁻¹. Elemental Analysis cal’d for C25H24N2O2: C, 57.80; H, 6.25; N, 10.94. Found: C, 58.12; H, 6.55; N, 10.64.

EX. 36
p-Nitrophenyl
N-[3-Methoxy-2-cyclohexen-1-yl-N-carboxy]ornithyl-30
prolyl methyl-N-isopropylcarbamate (50a, the LLL
diastereomer)

The title compound was prepared from 9 and 22 following a procedure analogous to that described for 44b. The amorphous powder containing 50a and 50b was dissolved in chloroform and applied to a preparative TLC plate (100 mg/plate) and developed two or three times with 4% methanol in ethyl acetate. The upper band contained 50a while the lower band contained 50b. The two bands were separately scraped from the plate, extracted with 20% methanol in chloroform (3 x 25 mL) and eluent solvent evaporated under reduced pressure. The desired product (50a) isolated as 0.11 g (0.14 mmoles) (16% yield) of a white amorphous powder, mp 55°-56°C, was obtained from the upper TLC band. 1H-NMR (CDCl3) δ 1.06-1.28 (6H,app.m, rotamers of l); 1.32 (3H,app.d, J = 8Hz); 1.38-2.32 (8H,m); 2.46 (2H,app.t, J = 8Hz); 2.64 (2H,app.t, J = 8Hz); 3.20 (2H,m); 3.52-3.88 (2H,m); 3.66 (3H,s); 4.20, 4.32 (2H, center of a set of dd, overlapping with another set, J = 20Hz, rotamers of the CH2(2)geminal system); 4.50 (2H,app.t, J = 8Hz); 4.50-4.80 (2H,app.m); 5.10 (2H,s); 5.42 (1H,app.m); 6.36-7.08 (2H,m, rotamers of amide -NH); 7.24 (1H,app.d, J = 10Hz, rotamers of l); 7.28 (1H,app.d, J = 10Hz, rotamers of l); 7.35 (5H,app.s) 8.20 (1H,app.d, J = 10Hz, rotamers of l); 8.25 (1H,app.d, J = 10Hz, rotamers of l) ppm. IR (CHCl3) 3305, 1720, 1645, 1520, 735, 700. Elemental Analysis cal’d for C39H34N2O2: C, 57.80; H, 6.25; N, 10.94. Found: C, 57.68; H, 6.39; N, 10.89.
EX. 37
p-Nitrophenyl
N-[Methoxysuccinylalanyl-(N<sub>3</sub>-benzoyl)ornithyl(prolyl-
methyl]-N-isopropylcarbamate (51b, the LLD
diastereomer)

The title compound was prepared from 50a and 50b
following a procedure analogous to that described for
47b. The residue, containing 51a and 51b, was applied to
a preparative TLC plate and developed two or three
times with 4% methanol in ethyl acetate. The upper
band contained 51a while the lower band contained 51b.
The two bands were separately scraped from the plate,
extracted with 20% methanol in chloroform (3 × 25
mL) and eluent solvent evaporated under reduced pres-
sure. The desired product (51b) isolated as 0.04 g (0.05
mmoles) (26% yield) of a white amorphous powder, mp
54°-55° C., was obtained from the lower TLC band.

EX. 38
p-Nitrophenyl
N-[Methoxysuccinylalanyl-(N<sub>3</sub>-benzoyl)ornithyl(prolyl-
methyl]-N-isopropylcarbamate (51a, the LLL
diastereomer)

The title compound was prepared from 50a and 50b
following a procedure analogous to that described for
47b. The residue, containing 51a and 51b, was applied to
a preparative TLC plate and developed two or three
times with 4% methanol in ethyl acetate. The upper
band contained 51a while the lower band contained 51b.
The two bands were separately scraped from the plate,
extracted with 20% methanol in chloroform (3 × 25
mL) and eluent solvent evaporated under reduced pres-
sure. The desired product (51a) isolated as 0.05 g (0.06
molees) (32% yield) of a white amorphous solid, mp 55°-56° C., was obtained from the upper TLC band.  

1H-NMR (CDCl₃) δ 1.06-1.28 (6H, d, rotamers of l); 1.32 (3H, d, J = 8Hz); 1.38-2.32 (8H, m); 2.46 (2H, app, t, J = 8Hz); 2.64 (2H, app, t, J = 8Hz); 3.20 (2H, m); 5.27-3.88 (2H, c, rotamers of the CH₃ group); 6.55 (4H, s); 6.02 (2H, center of a set of dd, overlapping with another set, J = 20Hz, rotamers of the CH₂(α) geminal system); 4.40 (2H, app, t, J = 8Hz); 7.45-7.50 (2H, m); 5.42 (1H, m); 6.36-7.08 (2H, m, rotamers of amide —NH); 7.74 (1H, app, d, J = 10Hz, rotamers of l); 7.28 (1H, app, d, J = 10Hz, rotamers of l); 7.34 (2H, app, d, J = 8Hz, J = 2Hz); 7.43 (1H, app, d, J = 8Hz, J = 2Hz); 7.53 (2H, app, d, J = 8Hz, J = 2Hz); 8.20 (1H, app, d, J = 10Hz, rotamers of l); 8.25 (1H, app, d, J = 10Hz, rotamers of l) ppm. IR (CHCl₃) 3340, 1730, 1645, 1520, 1435, 1345, 1260, 735, 700 cm⁻¹. Elemental Analysis calcd. for C₁₉H₂₉NO₂: C, 85.5; H, 11.8; N, 3.7. Found: C, 85.8; H, 11.3; N, 3.6.

2. Stereospecific synthesis of desmosine-like peptide carbamate 48a and 48b

EX. 39

N,α-Methoxy succinyl alaninyl-Nα-carbobenoxysulphyl proline phenacyl ester (23, the LLD diastereomer)

50

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55° C.) Impurities were eliminated by first passing methylene chloride and subsequently the compound was eluted with 4% methanol in methylene chloride. Upon evaporation of the eluant solvent under reduced pressure an oil was obtained. The latter was crystallized form ethyl acetate/hexane to give a white crystalline powder. Incorporation of 41 into the product gave 0.5 g (0.77 mmole) (96% yield) of crystalline powder, mp 114°-116° C.

Procedural 2

Isobutyl chloroformate (0.43 g, 3.2 mmole) in acetonitrile (4 ml) was added to a dry-ice/carbon tetrachloride cooled solution of 7 (1.3 g, 2.9 mmole) and N-methylmorpholine (0.3 g, 2.9 mmole) in THF (10 ml). After 10 min, D-proline phenacyl ester hydrochloride (41) (0.9 g, 1.5 mmole) as a solid and N-methylmorpholine (0.35 g, 3.5 mmole) in acetonitrile (6 ml) were added to the reaction mixture maintained at −15° C. The reaction mixture was allowed to warm to 5° C. in 30 min, was filtered after 90 min, and the filtrate was evaporated in vacuo. The residue was redissolved in methylene chloride (20 ml) and washed with water (20 ml), 10% aqueous citric acid (20 ml), water (20 ml) and brine (20 ml); dried (2 g of MgSO₄), and evaporated in vacuo. The residue, containing the product,

23

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was purified similar to the methodology in procedure 1. The transparent oil was crystalized from ethyl acetate/hexane. The incorporation of 41 gave 0.85 g (1.25 mmole) (43% yield) of white crystalline powder (23), mp 114°-116° C. 1H-NMR (CDCl₃) δ 1.36 (3H, d, J = 8Hz); 2.40-2.22 (10H, m); 2.44 (2H, app, t, J = 8Hz); 2.64 (2H, app, t, J = 8Hz); 3.28 (2H, m); 3.56 (2H, m); 3.66 (3H, s); 4.50 (2H, app, t, J = 8Hz); 4.53-4.80 (1H, m); 5.43 (2H, s); 5.76 (1H, rotamer of amide—NH); 6.70-7.30 (2H, m, rotamer of amide—NH); 7.36 (5H, s); 7.60 (3H, d); 8.06 (2H, app, d, J = 8Hz, J = 2Hz); 8.06 (2H, app, d, J = 8Hz, J = 2Hz) ppm. IR (CHCl₃) 3280, 1725, 1680, 1630, 725, 635 cm⁻¹.

Procedure 1

A concentrated solution of N,N′-dicyclohexylcarbodiimide (0.17 g, 0.8 mmole) in THF was added dropwise to an ice cooled solution containing 7 (0.4 g, 0.8 mmole) and N-hydroxy succinimide (0.1 g, 0.8 mmole) in THF (4 ml). The solution was stirred at 5° C. for 14 h and the precipitated urea formed was filtered under vacuum. The filtrate was immediately used in the next reaction without further work-up.

A mixture of the above cooled (0° C.) solution and D-proline phenacyl ester hydrochloride (41) (0.22 g, 0.8 mmole) was stirred and triethylamine (0.08 g, 0.8 mmole) in THF (0.5 ml) was added dropwise. Upon completion of the reaction (5 h), the precipitate was filtered, washed with ethyl acetate and the filtrate evaporated under vacuum. The residue, containing the product, was chromatographed on 20 g of silica gel column
The title compound was prepared by an analogous procedure to that described in the preparation of compound 23. L-Proline phenacyl ester hydrochloride 42 was substituted in place of the D-enantiomer to form the LLL diastereomer product 24. Following procedure 1, the incorporation of 42 into 24 gave rise to 0.4 g (0.54 mmole) (74% yield) of crystalline powder mp 116°-118° C. The LLL diastereomer 24 demonstrated a lower Rf value than the LLD diastereomer 23 on silica gel TLC plates eluted with 4% ethyl acetate in methylene chloride and 3% methanol in methylene chloride. Procedure 2, produced 1.3 g (1.94 mmole) (66.8% yield) of white crystalline powder (24), mp 116°-118° C., with similar Rf values. 1H-NMR (CDCl3) δ 1.36(3H, d, J = 8Hz); 1.40-2.22(10H, m); 2.44(2H, app. t, J = 8Hz); 2.64(2H, d, H2, app. t, J = 8Hz); 3.18(2H, m); 65 3.66(3H, s); 4.34-4.58(3H, h, m); 4.60-4.64(1H, m); 5.10(2H, s); 5.43(2H, s); 6.70-7.30(2H, m, rotamer of amide -NH); 7.36(5H, s); 7.60(3H, app. dd, J = 8Hz, J = 2Hz); 8.06(2H, app. dd, J = 8Hz, J = 2Hz).

Small portions of zinc metal (total 2 g) were added over a 1 h period to the phenacyl ester (23) (0.5 g, 0.8 mmole) dissolved in glacial acetic acid (10 mL). After 3 h, the suspension was diluted with 20% methanol in chloroform (50 mL), filtered under vacuum and evaporated to a crude solid. Work-up was performed in a similar manner to that described in the preparation of 7. The chromatographically pure oil required two crystallizations from chloroform/petroleum ether to yield a white solid. Hydrolysis of the phenacyl ester of 23 produced 0.26 g (0.46 mmole) (57% yield) of a white powder (25) mp 119°-120° C. 1H-NMR (CDCl3) δ 1.36(3H, d, J = 8Hz); 1.40-2.22(10H, m); 2.44(2H, app. t, J = 8Hz); 2.64(2H, app. t, J = 8Hz); 3.18(2H, m); 3.56(2H, m); 3.66(3H, s); 4.50(2H, app. t, J = 8Hz); 4.83(1H, m); 5.10(2H, s); 5.60(1H, m, rotamer of amide -NH); 6.70-7.26(2H, m, rotamer of amide -NH);
EX. 42

Nα-Methoxy succinylalanyl-N3-carbenzoxylserylproline (26, the LLD diastereomer)

(see 25 for structure)

The title compound was prepared by an analogous procedure to that described in the preparation of compound 25. Hydrolysis of the phenacyl ester of 24 produced 0.44 g (0.78 mmole) (98% yield) of a white powder (26), mp 119°-120° C. The LLD diastereomer 26 demonstrated a lower RF value than the LLD diastereomer 25 on silica gel TLC plates eluted with 4% methanol in ethyl acetate. 1H-NMR (CDCl3) δ 1.36(3H, m, J=8Hz); 1.40-2.22(10H, m); 2.44(2H, app. t, J=8Hz); 2.64(2H, app. t, J=8Hz); 3.18(2H, m); 3.56(2H, m); 3.66(3H, s); 4.34-4.58(3H, m); 4.60-4.56(1H, m, rotamer of amide —NH); 5.10(2H, s); 6.70-7.26(2H, m, rotamer of amide —NH); 7.36(5H, s); 10.20(1H, s).

EX. 43

Nα-Methoxy succinylalanyl-N3-carbenzoxylserylprolyl chloromethyl ketone (27, the LLD diastereomer)

50

Isobutylchlooroformate (0.11 g, 0.8 mmole) in THF (2 mL) was added to a cooled solution of 25 (0.45 g, 0.8 mmole) and N-methylmorpholine (0.08 g, 0.8 mmole) in THF (4.5 mL) and the mixture was stirred for 10 min at −15° C. A cold solution (0° C.) of diazomethane (ca. 0.13 gm, 3.2 mmoles) in diethyl ether was added and the mixture stirred at −10° C. for 30 min, then at 5° C. for 90 min. The reaction mixture was then diluted with ethyl acetate (40 mL), and washed with saturated aqueous sodium bicarbonate (2×30 mL), water (30 mL) and brine (30 mL); dried (5 g of MgSO4) and evaporated in vacuo to give a yellow oil. 1H-NMR for the α-azomethyketone intermediate (CDCl3) δ 0.86-2.23(1H, m), 2.56(4H, t, J=7Hz)m 3.03-3.40(2H, m), 3.46-4.00(5H, m), 4.40-4.83(3H, m), 5.13(2H, s), 5.40-5.80(2H, m), 6.53-7.20(2H, m), 7.40(5H, s). IR (CHCl3) 3280,2215,1735,1635,1535 cm−1.

EX. 44

Nα-Methoxy succinylalanyl-N3-carbenzoxylserylprolyl chloromethyl ketone (28, the LLD diastereomer)

(see 27 for structure)

The title compound was prepared by an analogous procedure to that described in the preparation of compound 27. The presence of impurities in the IR and NMR for the α-azomethyketone intermediate produced during the reaction did not demonstrate clear differences between the LLD and LLL diastereomers. The RF value of the LLD and LLL diastereomers were significantly different when eluted twice on silica gel plates with 4% ethyl acetate in methylene chloride or methylene chloride. The α-chloromethyketone 28 was crystallized from chloroform/petroleum ether to give 0.25 g (0.4 mmole) (54% yield) of crystalline powder,
EX. 48

N-[Methoxysuccinylalanyl-(N\textsubscript{\textepsilon}-carbobenzoxy)lysyl-prolylmethyl]-N-isopropylcarbamate (48a, the LLL diastereomer)

(see page 46 for structure)

The title compound was prepared by an analogous procedure to that described in the preparation of compound 48b. The reaction pathway which incorporated L-proline into the final peptidyl carbamate 48a gave rise to 153.9 mg (196.0 moles) (39% yield) of amorphous solid (48a), mp 56°-57° C. 1H-NMR (CDCl\textsubscript{3}) (see 48a above). Elemental analysis calcd. for C\textsubscript{36}H\textsubscript{43}N\textsubscript{2}O\textsubscript{12}: C, 58.30; H, 6.44; N, 10.73. Found: C, 58.12; H, 6.55; N, 10.64%.

3. Synthesis of protected amino acids

EX. 49

N\textsubscript{\textepsilon}-t-Boc-N\textepsilon-carbobenzoxylysine (31)

This compound was synthesized by a modified procedure of Hendrickson et al.\textsuperscript{85} t-BOC-ON (2.2 g, 8.9 mmoles) was added as a solid to a solution of triethylamine (0.9 g, 8.9 mmoles) and N\textepsilon-carbobenzoxy-L-lysine (2.5 g, 8.9 mmoles) in DMF (20 mL) at room temperature and the components allowed to react for 24 h. The resulting precipitate was filtered and the filtrate evaporated under vacuum. The crude oil was dissolved in methylene chloride and chromatographed on 50 g of silica gel column (3×50 cm). Impurities were eliminated by first passing 100 mL of methylene chloride and subsequently the compound was eluted with 10% methanol in methylene chloride. Upon evaporation of the eluent solvent under reduced pressure 3.6 g (7.2 mmoles) (81% yield) of a transparent oil was obtained. 1H-NMR (CDCl\textsubscript{3}) δ 1.46(9H\textsubscript{nat}); 1.50-2.20(6H\textsubscript{nat}); 3.23(2H\textsubscript{nat}); 4.00-4.30(1H\textsubscript{nat}); 5.10(2H\textsubscript{nat}); 5.56(2H\textsubscript{nat}); 6.56-7.10(2H\textsubscript{nat}), rotamers of carbamate-NH); 7.36(5H\textsubscript{nat}); 7.63(3H\textsubscript{nat}, appd. dd, J\textsubscript{d} = 8Hz, J\textsubscript{a} = 2Hz); 8.06(2H\textsubscript{nat},appd. dd, J\textsubscript{d} = 8Hz, J\textsubscript{a} = 2Hz)ppm.

EX. 50

N\textsubscript{\textepsilon}-t-Boc-N\textepsilon-carbobenzoxy-L-lysine phenacyl ester (32)

This compound was synthesized by a modified procedure of Itoh et al.\textsuperscript{91} t-BOC-ON (2.2 g, 8.9 mmoles) was added as a solid to a solution of triethylamine (0.9 g, 8.9 mmoles) and N\textepsilon-carbobenzoxy-L-lysine (2.5 g, 8.9 mmoles) in DMF (20 mL) at room temperature and the components allowed to react for 24 h. The resulting precipitate was filtered and the filtrate evaporated under vacuum. The crude oil was dissolved in methylene chloride and chromatographed on 50 g of silica gel column (3×50 cm). Impurities were eliminated by first passing 100 mL of methylene chloride and subsequently the compound was eluted with 10% methanol in methylene chloride. Upon evaporation of the eluent solvent under reduced pressure 3.25 g (8.6 mmoles) (96% yield) of an oil was obtained. 1H-NMR (CDCl\textsubscript{3}) δ 1.46(9H\textsubscript{nat}); 1.50-2.20(6H\textsubscript{nat}); 3.23(2H\textsubscript{nat}); 4.00-4.30(1H\textsubscript{nat}); 5.10(2H\textsubscript{nat}); 6.56-7.10(2H\textsubscript{nat}), rotamers of carbamate-NH); 7.36(5H\textsubscript{nat}); 9.80(1H\textsubscript{nat})ppm.
5,008,245

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mp 58°-60° C. 1H-NMR (CDCl3) δ 1.36 (3H, d, J=8Hz); 3.20 (2H, s); 3.56 (2H, s); 3.66 (3H, s); 4.23 (2H, s), center of 2 sets of dd, J=20Hz, rotamers of the CH2(g) geminal system; 4.34-4.58 (3H, m); 4.60-4.64 (1H, m); 5.10 (2H, s); 5.42 (1H, s), rotamer of amide —NH; 6.36-7.08 (2H, m, rotamers of amide —NH); 7.35 (5H, s) ppm.

EX. 45

N-[Methoxysuccinylalanyl-(N-carbobenzoxy)lysylprolylmethyl]-N-isopropylamine (29, the LLD diastereomer)

Isopropylamine (1.4 g, 24.4 mmoles) was added to a cold solution (0° C.) of 27 (1.45 g, 2.44 mmoles) in THF (5 ml). The reaction mixture was stirred at 0° C. for 12 h, filtered under vacuum and the filtrate evaporated to a crude oil. The residue, containing the product, was redissolved in a small amount of hexane, chloroform, and chromatographed on 25 g of silica gel column (1×50 cm). The product was eluted with 7% methanol in methylene chloride. The eluent solvent was evaporated under reduced pressure. The reaction product 0.31 g (0.5 mmole) (21% yield) of a yellow oil. 1H-NMR (CDCl3) δ 1.06-1.28 (6H, d, J=8Hz); 1.38-2.32 (10H, m); 2.46 (2H, s, J=8Hz); 2.64 (2H, s, t, J=8Hz); 3.20 (2H, m); 3.52-3.88 (3H, m); 3.66 (3H, s); 4.20, 4.32 (2H, m), center of a set of dd, overlapping with another set, J=20Hz, rotamers of the CH2(g) geminal system; 4.50 (2H, s, t, J=8Hz); 4.50-4.80 (2H, m); 5.10 (2H, s); 5.42 (1H, m, rotamer of amide —NH); 6.36-7.08 (2H, m, rotamer of amide —NH); 7.35 (5H, s) ppm.

EX. 47

p-Nitrophenyl N-[Methoxysuccinylalanyl-(N-carbobenzoxy)lysylprolylmethyl]-N-isopropyl carbamate (48b, the LLD diastereomer)

An ice cooled solution of 29 (0.3 g, 0.5 mmole), N-methylmorpholine (0.07 g, 0.7 mmole) and 4-nitrophenyl chloroformate (0.15 g, 0.75 mmole) in THF (3 ml) was stirred for 2 h at 0° C. The reaction mixture was diluted with methylene chloride (15 ml) and washed subsequently with water (15 ml), 10% aqueous citric acid (15 ml), water (15 ml) and brine (15 ml); dried (2 g of MgSO4) and evaporated in vacuo to an oil. The oil was chromatographed on 5 g of silica gel column (1×25 cm). The impurities were eliminated by first passing 10 ml of methylene chloride and 20 ml of 2% methanol in methylene chloride. Subsequently the compound was eluted with 4% methanol in methylene chloride. Upon evaporation of the eluent solvent under pressure an amorphous solid was obtained. The product was further purified by preparative TLC (100 mg/plate). The TLC plates were developed twice using 4% methanol in ethyl acetate in order to see if two bands were present. The one band observed was scraped from the plate and the product was extracted with 20% methanol in chloroform. An amorphous white solid was obtained by evaporating the extraction solvent under reduced pressure. The incorporation of D-proline into the synthetic scheme gave rise to 89.9 mg (115.0 moles) (23% yield) of amorphous solid (48b), mp 44°-45° C. 1H-NMR (CDCl3) (see 48b above). Elemental analysis cal'd. for C34H39N3O12: C, 58.3; H, 6.44; N, 10.73. Found: C, 58.49; H, 6.47; N, 10.63.

(see page 45 for structure)

EX. 46

N-[Methoxysuccinylalanyl-(N-carbobenzoxy)lysylprolylmethyl]-N-isopropylamine (30, the LLD diastereomer)

The title compound was prepared by an analogous procedure to that described in the preparation of compound 29. The reaction produced 0.31 g (0.5 mmole) (21% yield) of a yellow oil. 1H-NMR (CDCl3) δ 1.06-1.28 (6H, d, J=8Hz); 1.32 (3H, s, J=8Hz); 1.38-2.32 (10H, m); 2.46 (2H, s, J=8Hz); 2.64 (2H, s, t, J=8Hz); 3.20 (2H, m); 3.52-3.88 (3H, m); 3.66 (3H, s); 4.23 (2H, m), center of 2 sets
Formic acid (98%) (6 mL) was added to a 10% solution of 32 (3.3 g, 6.9 mmoles) in ethyl acetate (50 mL) cooled to 5° C. Hydrogen chloride gas was slowly bubbled through the cooled solution in three 30s intervals, 10 min apart. The solution was stirred at 5° C for 30 min and was then allowed to equilibrate to room temperature. The progress of the reaction was monitored by TLC (10% methanol in chloroform). Upon completion of the reaction (3 h at 22° C), the suspension was filtered. The latter was recrystallized from absolute ethanol/hexane to give 2.2 g (5.1 mmoles) (73% yield) of a crystalline powder, m.p. 149°-151° C. 1H-NMR (DMSO-d6) δ 1.50-2.26(6H, m); 3.23(2H, m); 4.03-4.43(1H, m); 5.10(2H, d); 5.80(2H, s); 7.10(1H, m); 7.36(5H, s); 7.63(3H, K, K, app. dd, JHα=8Hz, JHβ=2Hz); 8.06(2H, J, app. dd, Jα=8Hz, Jβ=2Hz); 8.46(2H, m)ppm. IR(Nujol) 3350, 1750, 1700, 1680, 1585, 1535 cm^{-1}.  

EX. 52  
Nα-t-Boc-Nβ-carbomethoxy-L-ornithine (34)  

The title compound was synthesized and purified by an analogous procedure to that described in the preparation of compound 32. The reaction produced 3.7 g (7.7 mmoles) (86% yield) of a transparent oil. 1H-NMR (CDCl3) δ 1.46(9H, s); 1.50-2.20(4H, m); 3.23(2H, m); 4.00-4.30(1H, m); 5.10(2H, s); 5.56(2H, s); 6.56-7.10(2H, m, rotamer of carbamate -NH); 7.36(5H, s); 7.63(3H, J, app. dd, Jα=8Hz, Jβ=2Hz); 8.06(2H, K, app. dd, Jα=8Hz, Jβ=2Hz)ppm.  

EX. 54  
Nβ-Carbomethoxy-L-ornithine phenacyl ester hydrochloride (36)  

The title compound was prepared by an analogous procedure to that described in the preparation of compound 33. The resulting white powder was recrystallized from ethanol/hexane to give 2.4 g (5.7 mmoles) (76% yield) of a crystalline powder mp 166°-167° C. 1H-NMR (DMSO-d6) δ 1.50-2.26(4H, m); 3.23(2H, m);
The procedure of Itoh et al.91 was followed for the preparation of compound 37. Triethylamine (0.2 g, 2 mmole) was slowly added to a solution of (D-proline (0.23 g, 2 mmole) and t-BOC-ON (0.5 g, 2 mmole) in DMF (3 mL) at room temperature and the mixture stirred for 36 h. The DMF was coevaporated with toluene under vacuum. Aqueous hydrochloric acid (1 × 10−4M) (20 mL) and ethyl acetate (20 mL) were added to the flask to dissolve the residue. The organic layer was washed with water (3 × 20 mL) and brine (2 × 20 mL); dried (5 g of MgSO4) and evaporated under reduced pressure. The residue, containing the product, was chromatographed on 5 g of silica gel column (1 × 25 cm). Upon evaporation of the eluent solvent under reduced pressure a white solid was obtained. The latter was recrystallized from ethyl acetate/hexane. The incorporation of 37 into the product gave 0.58 g (1.75 mmole) (87.5% yield) of crystalline powder (39), mp 78°~79° C. 1H-NMR (CDCl3) δ 1.46(9H, s); 1.86~2.40(4H, m); 3.20~3.60(2H, m); 4.30(1H, m); 5.80(2H, s); 7.63(3H, f, f, app. dd, Jα=8Hz, Jγ=2Hz); 8.06(2H, H, app. dd, Jα=8Hz, Jγ=2Hz); ppm.

EX. 58
N-t-Boc-L-proline phenacetyl ester (40)
(see 39 for structure)

The title compound was prepared by an analogous procedure to that described in the preparation of compound 39. t-Boc-L-proline (38) was substituted into the reaction to produce 0.59 g (1.78 mmole) (88.8% yield) of crystalline powder (40), mp 76°~78° C.

EX. 59
D-Proline phenacetyl ester hydrochloride (41)

Formic acid (98%) (0.6 mL) was added to a cooled solution (0° C.) of t-Boc-D-proline phenacetyl ester (0.6 g, 1.75 mmole) in ethyl acetate (10 mL). Hydrogen chloride gas was slowly bubbled through the above solution in two 30s intervals, 10 min apart. The reaction mixture was stirred for 30 min at 0~5°C, then allowed to equilibrate to room temperature and stirred until the reaction was completed (approximately 1 h). The suspension was diluted with ethyl acetate and filtered under vacuum. The product was washed with ethyl acetate, air dried and recrystallized from absolute ethanol/diethyl ether. The use of 39 (the D isomer) in this reaction gave 0.3 g (1.1 mmole) (63.5% yield) of crystalline powder (41), mp 154°~156° C. 1H-NMR (DMSO-d6) δ 1.86~2.76(4H, m); 3.16~3.63(2H, m); 4.40~4.86(1H, m); 5.80(2H, s); 7.63(3H, f, f, app. dd, Jα=8Hz, Jγ=2Hz); 8.06(2H, H, app. dd, Jα=8Hz, Jγ=2Hz); 9.30(1H, m) ppm. IR (Nujol) 1755, 1600, 1500 cm−1.

EX. 60
L-Proline phenacetyl ester hydrochloride (42)
(see 41 for structure)

The title compound was prepared by an analogous procedure to that described in the preparation of compound 41. t-Boc-L-proline phenacetyl ester was substi-
This compound was synthesized by a modified procedure of Kurtz et al. Copper (II) carbonate, basic (16.3 g, 74.0 mmole) was added to a hot (80°C) solution of L-lysine (15 g, 82.0 mmole) in distilled water (250 mL). Excess copper carbonate was removed by gravity filtration while the mixture was still warm. The epsilon amino group of L-lysine was benzoylated by the dropwise addition of benzoyl chloride (14 g, 99.0 mmole) in THF (35 mL) to the above solution after being cooled to 5°C. Sodium bicarbonate was added in small portions (total 14 g, 167.0 mmole) to maintain the pH of the aqueous solution above 7 (pH was monitored with neutral litmus paper). The cold solution was maintained at 5°C for 4 h and 2 d at room temperature. A hot (80°C) solution (1 L) of EDTA (14.6 g, 0.25 moles) was added to the reaction mixture, stirred for 1 h and cooled to 10°C. The precipitate collected by vacuum filtration was washed with water and 30 mL of 95% ethanol. The powder, containing the product was purified by first adding the powder to 1 × 10−3 M hydrochloric acid (300 mL) and filtering off the sediment. The filtrate was slowly neutralized with 0.01N sodium hydroxide and the precipitate collected by gravity filtration. The latter was then air-dried to give 13 g (57 mmole) (69.5% yield) of a white powder, mp 232°–234°C (lit.93 mp 230°–233°C). 1H-NMR (DMSO-d6) δ 1.36–190(–3H, m); 3.20(2H, m); 4.20–4.53(1H, m); 7.16(1H, m, rotamer of amide -NH); 7.43(2H, t, app. dd, J6,H = 8Hz, J7,CH = 2Hz); 7.63(1H, t, app. dd, J7,CH = 8Hz, J8,CH = 2Hz); 8.03(2H, t, app. dd, J7,CH = 8Hz, J8,CH = 2Hz); 9.50(1H, s). ppm. IR (Nujol) 3330, 3030, 1687, 1270, 735, 695 cm−1.

Compound 52 (1.30 g, 5.40 mmole) was added to a solution of compound 20 (0.8 g, 2.96 mmole) and triethylamine (0.6 ml, 4.30 mmol) 5°C. The mixture was stirred for 2 h at 5°C. The reaction mixture was diluted with chloroform. The organic layer was washed with water, dried over MgSO4 and evaporated to give an oil. The oil obtained was purified by column chromatography (silica gel, CHCl3:1% MeOH in CHCl3 - 2% MeOH in CHCl3) to give white crystals (0.6 g, 42.8%). 1H-NMR (CDCl3) δ 8.12 (6Ha, d), 1.49(9Ha, s), 1.95(4Hb, m), 3.45 (4Hc, t) 4.2 (4Hc, m), 7.55(5Hb, s). Anal. Calcd for C22H33N6O4S: C, 55.69; H, 6.37; N, 17.71; S, 6.74. Found: C, 55.50; H, 6.43; N, 17.66; S, 6.68.
EX. 64

S-(1-Phenyl-5-tetrazoyl)-N-prolylmethyl-N-isopropyl thiocarbamate hydrochloride 54

Hydrogen chloride gas was passed through a solution of 53 (0.6 g) in ethyl acetate (5 ml) at 5°C for 3 min. The solution was allowed to stand at 5°C for 10 min and then evaporated in vacuo. The residue was triturated with ethyl ether to give white powder (0.24 g, 46%). 1H NMR 61.2 (6Ha, d), 2.0 (4H6, m), 3.4-4.9 (7Hc, m), 7.5 (5Hd, s).

EX. 65

S-(1-Phenyl-5-tetrazoyl)-N-[methoxysuccinyl-alanyl-(N6-carbonbenzoxyl)lysylpropylmethyl]-N-isopropyl thiocarbamate (PC5)

PC6 was prepared following a similar procedure as for the preparation of PC5 using compound 12 instead of 7. PC6 was obtained as a white powder (37.3%). m.p. 75°C. 1H NMR (CDCl3) 81.2 (9Ha, m), 1.6 (4Ha, m), 2.0 (4Hb, m), 2.5 (4Hb', s), 2.9-3.2 (4H6', m), 3.6 (3Hc, s), 3.7-4.8 (6Hd, m), 5.05 (2He, s), 5.3-6.9 (3Hf, m) 7.25 (5Hg, s), 7.5 (5Hg, s). Anal. calc'd for C38H39N5O8S: C, 56.30; H, 6.11; N, 15.60; S, 3.96. Found: C, 56.59; H, 6.16; N, 15.53; S, 3.92.

EX. 67

Elastase Enzyme Inhibitory Studies

These studies are conducted to show the activities of some of the compounds of this invention as inhibitors of...
various forms of the enzyme elastase. The results are shown in the following Tables.

Inhibition of PPE and HLE by Novel Peptidyl Carbamates; Variations at P₃

<table>
<thead>
<tr>
<th>Compound</th>
<th>P₃ᵃ</th>
<th>Isomerᵇ</th>
<th>Kᵢ (µM)</th>
<th>PPE</th>
<th>HLE</th>
</tr>
</thead>
<tbody>
<tr>
<td>5 (a,b)</td>
<td>N₄-CH₂-Lys</td>
<td>a (D)</td>
<td>N.I.ᶜ</td>
<td>3.40</td>
<td>0.22</td>
</tr>
<tr>
<td>6 (a,b)</td>
<td>N-ε-Bz-Lys</td>
<td>a (D)</td>
<td>N.I.</td>
<td>3.80</td>
<td>0.31</td>
</tr>
<tr>
<td>7 (a,b)</td>
<td>N-δ-CH₂-Orn</td>
<td>a (D)</td>
<td>N.I.</td>
<td>19.25</td>
<td>0.08</td>
</tr>
<tr>
<td>8 (a,b)</td>
<td>N-ε-Bz-Orn</td>
<td>a (D)</td>
<td>N.I.</td>
<td>14.30</td>
<td>2.14</td>
</tr>
</tbody>
</table>

ᵃ CH₂ = COOCH₂Ph; Bz = COPh; Lys = lysine; Orn = ornithine
ᵇ (L) and (D) refer to the configuration at the prolyl α-carbon
ᶜ N.I. refers to no inhibition at I/E = 100

Inhibition of PPE and HLE by Novel Peptidyl Carbamates Variations at P₄

<table>
<thead>
<tr>
<th>Compound</th>
<th>P₄ᵃ</th>
<th>Isomerᵇ</th>
<th>Kᵢ (µM)</th>
<th>PPE</th>
<th>HLE</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 (a,b)</td>
<td>N-ε-CH₂-Lys</td>
<td>a (L)</td>
<td>N.I.ᶜ</td>
<td>0.47</td>
<td>0.73</td>
</tr>
<tr>
<td>2 (a,b)</td>
<td>N-ε-Bz-Lys</td>
<td>a (L)</td>
<td>N.I.</td>
<td>0.38</td>
<td>0.08</td>
</tr>
<tr>
<td>3 (a,b)</td>
<td>N-δ-CH₂-Orn</td>
<td>a (L)</td>
<td>N.I.</td>
<td>0.65</td>
<td>0.70</td>
</tr>
<tr>
<td>4 (a,b)</td>
<td>N-ε-Bz-Orn</td>
<td>a (L)</td>
<td>N.I.</td>
<td>17.70</td>
<td>0.10</td>
</tr>
</tbody>
</table>

ᵃ CH₂ = COOCH₂Ph; Bz = COPh; Lys = lysine; Orn = ornithine
ᵇ (L) and (D) refer to the configuration at the prolyl α-carbon
ᶜ N.I. refers to no inhibition at I/E = 100

The invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the invention as set forth herein.

What is claimed herein is:

1. A compound selected from the group consisting of a compound of the formula

\[ \text{MeO} \stackrel{\text{P₃}}{\longrightarrow} \text{N} \stackrel{\text{P₄}}{\longrightarrow} \text{H} \stackrel{\text{P₅}}{\longrightarrow} \text{NH} \stackrel{\text{Y}}{\longrightarrow} \text{N} \stackrel{\text{P₆}}{\longrightarrow} \text{H} \stackrel{\text{P₇}}{\longrightarrow} \text{N} \]

wherein

- x is 1 or 2;
- Y is carbobenzoxy or benzyol; and XR is

\[ \text{NO₂ or } \text{SO₂} \]

and
2. The compound of claim 1 having the formula

3. The compound of claim 1 having the formula

4. The compound of claim 1 being selected from the group consisting of

1. p-Nitrophenyl N-[(Methoxysuccinyl)-L-alanyl-L-allyl-L-prolylmethyl]-N-isopropylcarbamate,
2. Methyl succinimide succinate
3. t-Butyl Methoxysuccinyl-L-alanine ester
4. Methoxysuccinyl-L-alanine,
5. Nα-Methoxysuccinyl-L-alanyl-Nε-benzoyl-L-lysine,
6. Nα-Methoxysuccinyl-L-alanyl-Nε-carboxbenzoxyl-L-lysine phenacetyl ester,
7. Nα-Methoxysuccinyl-L-alanyl-Nε-carboxbenzoxyl-L-lysine
8. Nα-Methoxysuccinyl-L-alanyl-Nε-carboxbenzoxyl-L-lysine phenacetyl ester,
9. Nα-Methoxysuccinyl-L-alanyl-Nε-carboxbenzoxyl-L-lysine
10. Nα-Methoxysuccinyl-Nε-carboxbenzoxyl-L-lysine
11. Nα-Methoxysuccinyl-Nε-carboxbenzoxyl-L-lysine phenacetyl ester,
12. Nα-Methoxysuccinyl-Nε-carboxbenzoxyl-L-lysine
13. Nα-Methoxysuccinyl-Nε-carboxbenzoxyl-L-lysine
14. Nα-Methoxysuccinyl-Nε-carboxbenzoxyl-L-lysine
15. Nα-Methoxysuccinyl-Nε-carboxbenzoxyl-L-lysine
16. Nα-Methoxysuccinyl-Nε-benzoyl-L-lysine
17. Nα-Methoxysuccinyl-Nε-benzoyl-L-lysine
18. Nα-Methoxysuccinyl-Nε-benzoyl-L-lysine
19. Nα-Methoxysuccinyl-Nε-benzoyl-L-lysine
20. Nα-Methoxysuccinyl-Nε-benzoyl-L-lysine
21. Nα-Methoxysuccinyl-Nε-benzoyl-L-lysine
22. p-Nitrophenyl N-[(L-prolylmethyl)-N-isopropylcarbamate hydrochloride,
p-Nitrophenyl N-[Methoxysuccinyl-(Nα-carbobenzoxyl)-L-lysyl-L-alanyl-D-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-(Nα-benzoyl)-L-lysyl-L-alanyl-L-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-(Nα-benzoyl)-L-lysyl-L-alanyl-D-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-(Nα-carbobenzoxyl)-L-ornithyl-L-alanyl-L-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-(Nα-carbobenzoxyl)-L-ornithyl-L-alanyl-D-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-(Nα-carbobenzoxyl)-L-ornithyl-L-alanyl-L-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-(Nα-carbobenzoxyl)-L-lysyl-L-prolylmethyl]-N-isopropylcarbamate, and


6. The compound of claim 1 selected from the group consisting of

p-Nitrophenyl N-[Methoxysuccinyl-L-alanyl-(Nα-carbobenzoxyl)-L-lysyl-D-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-L-alanyl-(Nα-benzoyl)-L-lysyl-L-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-L-alanyl-(Nα-benzoyl)-L-lysyl-D-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-L-alanyl-(Nα-carbobenzoxyl)-L-ornithyl-L-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-L-alanyl-(Nα-carbobenzoxyl)-L-ornithyl-D-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-L-alanyl-(Nα-carbobenzoxyl)-L-ornithyl-L-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-L-alanyl-(Nα-carbobenzoxyl)-L-ornithyl-D-prolylmethyl]-N-isopropylcarbamate,

p-Nitrophenyl N-[Methoxysuccinyl-L-alanyl-(Nα-benzoyl)-L-ornithyl-D-prolylmethyl]-N-isopropylcarbamate,

8. The compound of claim 1 having the formula

wherein

x is 1 or 2;

Y is benzoyl, and

XR is

9. The compound of claim 1 having the formula

wherein

x is 1 or 2;

Y is carbobenzoxy; and

XR is

10. The compound of claim 1 having the formula

wherein

x is 1 or 2;

Y is carbobenzoxy; and

XR is
11. The compound of claim 1 having the formula

wherein
-\text{x} = 1 or 2;
-\text{Y} is carboxybenzoyl or benzoyl; and
-\text{XR} is

12. The compound of claim 1 having the formula

wherein
-\text{x} = 1 or 2;
-\text{Y} is carboxybenzoyl or benzoyl, and
-\text{XR} is

13. The compound of claim 1 having the formula

wherein
-\text{x} = 1 or 2;
-\text{Y} is carboxybenzoyl or benzoyl, and
-\text{XR} is

14. The compound of claim 1 having the formula

wherein
-\text{x} = 1 or 2;
-\text{Y} is carboxybenzoyl or benzoyl; and
-\text{XR} is

15. An enzyme elastase inhibitory composition, comprising

an enzyme elastase inhibitory amount of the compound of claim 1; and

a carrier.

16. The composition of claim 14, wherein

the carrier is a pharmaceutically-acceptable carrier.

17. A method of selectively inhibiting the enzyme

elastase in an animal or a human in need of such treatment comprising administering to said animal or human an enzyme elastase inhibitory amount of the compound of claim 1.
18. A method of selectively inhibiting the enzyme elastase in an animal or a human in need of such treatment comprising administering to said animal or human an enzyme elastase inhibiting amount of the composition of claim 15.

19. The compound of claim 1 being selected from the group consisting of