A Comprehensive Selection of Reference Genes for RT-qPCR Analysis in a Predatory Lady Beetle, *Hippodamia convergens* (Coleoptera: Coccinellidae)

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A Comprehensive Selection of Reference Genes for RT-qPCR Analysis in a Predatory Lady Beetle, *Hippodamia convergens* (Coleoptera: Coccinellidae)

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Abstract

Reverse transcriptase-quantitative polymerase chain reaction (RT-qPCR) is a reliable, rapid, and reproducible technique for measuring and evaluating changes in gene expression. To facilitate gene expression studies and obtain more accurate RT-qPCR data, normalization relative to stable reference genes is required. In this study, expression profiles of seven candidate reference genes, including β-actin (*Actin*), elongation factor 1α (*EF1A*), glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*), cyclophilins A (*CypA*), vacuolar-type H+–ATPase (*ATPase*), 28S ribosomal RNA (*28S*), and 18S ribosomal RNA (*18S*) from *Hippodamia convergens* were investigated. *H. convergens* is an abundant predatory species in the New World, and has been widely used as a biological control agent against sapsucking insect pests, primarily aphids. A total of four analytical methods, geNorm, Normfinder, BestKeeper, and the ΔCt method, were employed to evaluate the performance of these seven genes as endogenous controls under diverse experimental conditions. Additionally, RefFinder, a comprehensive evaluation platform integrating the four above mentioned algorithms, ranked the overall stability of these candidate genes. A suite of reference genes were specifically recommended for each experimental condition. Among them, *28S*, *EF1A*, and *CypA* were the best reference genes across different development stages; *GAPDH*, *28S*, and *CypA* were most stable in different tissues. *GAPDH* and *CypA* were most stable in female and male adults and photoperiod conditions, *28S* and *EF1A* were most stable under a range of temperatures, *Actin* and *CypA* were most stable under dietary RNAi condition.

This work establishes a standardized RT-qPCR analysis in *H. convergens*. Additionally, this study lays a foundation for functional genomics research in *H. convergens* and sheds light on the ecological risk assessment of RNAi-based biopesticides on this non-target biological control agent.
Introduction

Reverse transcriptase-quantitative polymerase chain reaction (RT-qPCR) is a rapid, reliable, and reproducible method measuring gene expression during different biological processes [1]. Although RT-qPCR is considered as a major breakthrough in PCR technology, limitations exist, including variation in RNA extraction, reverse transcription, cDNA concentration, normalization, and PCR efficiency [2, 3, 4]. A commonly used technique in RT-qPCR to normalize the gene expression data is to simultaneously measure the expression of an internal reference gene in the same sample [1]. Reference genes, constitutively expressed to maintain cellular function, are the conventional choice for a standardized reference [1], although it is impractical to find a single reference gene expressed at constant levels under all biotic and abiotic conditions [5, 6].

The convergent lady beetle, *Hippodamia convergens* (Coleoptera: Coccinellidae), is a common lady beetle species in the New World. Although *H. convergens* larvae and adults are polyphagous, they prey primarily on aphids, including cotton, pea, melon, cabbage, potato, green peach, and corn leaf aphids [7]. As an effective biological control agent, *H. convergens* can reduce aphid densities in greenhouses [8, 9]. When aphids are limited, however, *H. convergens* feeds on thrips, small insect larvae and eggs, mites, and honeydew secreted by aphids and other sap-sucking insects, as well as plant based foods, such as pollen, nectar, and petals [10]. Nevertheless, convergent lady beetles must consume preys to reproduce [11].

The advent of the Genomics Era provides unprecedented opportunity to develop novel biopesticides with new modes of action to complement existing biological control agents. Most recently, RNA interference (RNAi)-based transgenic technology has been developed and offers a new approach for the management of insect pests [12–14]. For example, Baum and colleagues (2007) developed transgenetic corn plants that resisted the western corn rootworm, *Diabrotica virgifera virgifera*. By reducing translation of vacuolar H+-ATPase subunit A in the pest, the plant increased pest mortality and larval stunting and experienced less root damage as a result [12]. Although technical difficulties and regulatory hurdles still exist [15], the commercialization of a new generation of genetically modified crops is likely [16]. One major ecological concern regarding the biosafety of RNAi-based biopesticides and transgenic crops on the ecosystems is their potential effects on non-target organisms, especially biological control agents which play an important role in current pest management practices [17, 18]. Given the nature of RNAi mechanisms, the non-target effects will likely be through the modulation of gene expressions in non-target organisms [19]. Therefore, RT-qPCR will be a major research tool to evaluate potential non-target effects of this new biotechnology. Despite the demonstrated necessity for systematic validation of reference genes in RT-qPCR studies, normalization procedures for biological control agents have not yet received attention. These natural enemies are a major group of non-target organisms that will be exposed to the RNAi-based biopesticides and transgenic crops in the field.

The objective of this study is to evaluate and select appropriate reference genes with stable expression across various biotic and abiotic conditions in *H. convergens*. Seven candidate reference genes including β-actin (*Actin*), elongation factor 1 α (*EF1A*), glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*), cyclophilins A (*CypA*), vacuolar-type H⁺-ATPase (*ATPase*), 28S ribosomal RNA (*28S*), and 18S ribosomal RNA (*18S*) from *H. convergens* were tested. The stability of these candidate genes was investigated under three biotic (developmental stage, tissue type, and sex) and three abiotic conditions (temperature, photoperiod, and dietary RNAi). As a result, different sets of reference genes were recommended based on the respective experimental conditions.
Materials and Methods

Insects

*Hippodamia convergens* (Coleoptera: Coccinellidae) was purchased from a commercial vendor in California (High Sierra Ladybugs, http://www.highsierraladybugs.com). *Hippodamia convergens* larvae and adults were maintained in a growth chamber and provisioned with pea aphids, *Acyrthosiphon pisum*, at 23 ± 1°C temperature, 12L: 12D photoperiod, and 50% relative humidity. *A. pisum* clones were kindly provided by Dr. John Obrycki (University of Kentucky), and aphids were maintained at 20–28°C on seedlings of fava beans, *Vicia faba* (Fabales, Fabaceae) in a greenhouse.

Treatments

**Biotic factors.** The different developmental stages including eggs, four larval instars (collected at the first day of each instar), pupae, and adults. Tissues, including head, midgut, and carcass (body with its head and viscera removed) were dissected from the fourth instars. For the sex treatment, one adult female and male were collected separately and placed in 1.5 ml centrifuge tubes, respectively.

**Abiotic factors.** To examine the influence of temperature, third instars were exposed to 10, 22, and 30°C for 3 h. For photoperiod, third instars were exposed to 16:8, 12:12, and 8:16 h light: dark period for 2 d. For the dietary RNAi treatment, neonate first instars were fed with 15% sucrose solution containing chemically synthesized dsRNAs for ATPase, which causes mortality in *H. convergens* larvae (ATPase-dsRNA) (HP Pan unpublished data), water and β-glucuronidase dsRNA (GUS-dsRNA) as controls. At the start of the experiment, *H. convergens* neonate larvae (< 24 h old) were kept individually in a petri dish. Each neonate was provisioned with a 2 µl droplet containing 1 µl of dsRNA (8 µg/µl) and 1 µl of 30% sucrose solution on a daily basis. In 2 d, 16 µg of dsRNA were supplied to each *H. convergens* neonate. At day 3, five individuals in each treatment were collected as one sample for the subsequent RT-qPCR analysis.

For the developmental stage, a total of 15 eggs were collected as one biological replicate, while one pupa was collected individually as one replicate. For the remaining developmental stages, and all other biotic and abiotic conditions, five individuals were collected for each treatment. Each experiment was repeated three times. All collected samples were snap frozen in liquid nitrogen and stored at -80°C in 1.5 ml centrifuge tubes for the subsequent total RNA extraction.

RNA extraction and cDNA synthesis

Total RNA was extracted using TRIzol reagent (Invitrogen, Carlsbad, CA) following the manufacturer’s instruction. The whole body homogenates were centrifuged at 12000 g for 15 min and then supernatant was transferred to a new 1.5 ml microcentrifuge tube. A volume of 200 µl chloroform was added to the supernatant and then the mixture was incubated at room temperature for 10 min and then centrifuged at 4°C, 12000 g for 15 min. After that, the supernatant was transferred to a new 1.5 ml microcentrifuge tube, 400 µl Isopropyl alcohol was added to it, and the mixture precipitated at room temperature for 10 min. Then, the supernatant was discarded after the mixture was centrifuged at 4°C, 12000 g for 8 min, and then 1 ml 75% alcohol was added and centrifuged at 4°C, 7500 g for 5 min to wash the pellet. Finally, the pellet was air dried for 8 min and then dissolved in 30–80 µl ddH2O. DNase treated total RNA was denatured at 75°C for 5 min and immediately chilled on ice. The concentration of RNA was quantified with a NanoDrop 2000c spectrophotometer with the result for eggs (530.1 ± 28.2 ng/µl),
first instars (549.3 ± 35.6 ng/μl), second instars (1377.7 ± 164.7 ng/μl), third instars (2387.9 ± 369.6 ng/μl), fourth instars (1376.5 ± 332.0 ng/μl), pupae (733.8 ± 226.1 ng/μl), adults (914.9 ± 377.5 ng/μl), heads (1038.2 ± 78.9 ng/μl), carcasses (3147.9 ± 372.7 ng/μl), and guts (2591.7 ± 1620.0 ng/μl). First-strand cDNA was synthesized from 2.0 μg of total RNA using the M-MLV reverse transcription kit (Invitrogen, Carlsbad, CA) according to the manufacturer’s recommendations. The cDNA was diluted 10-fold for the subsequent RT-qPCR studies.

### Reference Gene Selection and Primer Design

Seven reference genes were selected (Table 1). Primers for 18S and 28S were designed based on the sequences obtained from NCBI (accession number: EU164617 and EU164644, respectively). For the remaining five reference genes, degenerate primers were designed using CODEHOP (http://blocks.fhcrc.org/codehop.html) according to the conserved amino acid residues among Coleopteran (S1 Table). PCR amplifications were performed in 50 μl reactions containing 10 μl 5×PCR Buffer (Mg²⁺ Plus), 1 μl dNTP mix (10 mM of each nucleotide), 5 μl of each primer (10 μM each), and 0.25 μl of Go Taq (5u/μl) (Promega). The PCR parameters were as follows: one cycle of 94°C for 3 min; 35 cycles of 94°C for 30 s, 59°C for 30 s, 72°C for 1 min; a final cycle of 72°C for 10 min. Amplicons of the expected sizes were purified, cloned into the pCR4-TOPO vector (Invitrogen, Carlsbad, CA), and sent out for sequencing (S2 Table). After the identity of these reference genes was confirmed, primers for the subsequent RT-qPCR analysis were designed online, https://www.idtdna.com/Primerquest/Home/Index.

### Reverse Transcriptase-Quantitative Polymerase Chain Reaction (RT-qPCR)

PCR reaction (20 μl) contained 7 μl of ddH₂O, 10 μl of 2×SYBR Green MasterMix (BioRad), 1 μl of each specific primer (10 μM), and 1 μl of the first-strand cDNA template. The RT-qPCR program included an initial denaturation for 3 min at 95°C followed by 40 cycles of denaturation at 95°C for 10 s, annealing for 30 s at 55°C, and extension for 30 s at 72°C. For melting curve analysis, a dissociation step cycle (55°C for 10 s, and then 0.5°C for 10 s until 95°C) was
added. Reactions were set up in 96-well format Microseal PCR plates (Bio-Rad) in triplicates, and carried out in a MyiQ single Color Real-Time PCR Detection System (BioRad). Existence of a single peak in melting curve analysis was used to confirm gene-specific amplification and rule out non-specific amplification and primer-dimer generation. A standard curve for each primer pair was constructed with serial dilutions of cDNA (1, 1/5, 1/25, 1/125, 1/625, and 1/3125). PCR amplification efficiency (E) was calculated according to the equation: $E = (10^{-1/\text{slope}} - 1) \times 100$.

Expression stability analysis of candidate reference genes

The stability of each candidate reference gene was evaluated by algorithms geNorm [1], Norm-Finder [20], BestKeeper [21], and the ΔCt method [22]. Finally, we compared and ranked the tested candidates based on the comprehensive web-based analysis tool RefFinder (http://www.leonxie.com/referencegene.php).

Results

Primer specificity and efficiency

Seven candidate reference genes were visualized as a single amplicon on a 1.5% agarose gel. Furthermore, gene-specific amplification of these genes was confirmed by a single peak in melting-curve analysis (S1 Fig). The correlation coefficient ($R^2$) and PCR efficiency for each standard curve were shown in Table 1.

$C_t$ values of candidate reference genes

For the developmental stage experiment, the $C_t$ values ranged from 10 to 25. 18S and 28S showed average $C_t$ values below 15 cycles. Actin was less expressed in the egg stage with the $C_t$ values above 28. 18S and ATPase were the most and the least expressed reference gene, respectively (Fig 1A).

For the sex experiment, the $C_t$ values ranged from 13 to 28. 18S and 28S showed average $C_t$ values below 15 cycles. All of the seven candidate reference genes were equally expressed in both female and male adults. 18S and ATPase were the most and the least expressed reference gene, respectively (Fig 1B).

For the tissue experiment, the $C_t$ values ranged from 11 to 24. 18S and 28S showed average $C_t$ values below 12 cycles. Actin showed average $C_t$ values below 18. 18S and ATPase were the most and the least expressed reference gene, respectively (Fig 1C).

For the temperature experiment, the $C_t$ values ranged from 9 to 24. 18S and 28S showed average $C_t$ values below 12 cycles. Actin showed average $C_t$ values below 18. EF1A, GAPDH, and CypA showed average $C_t$ values below 23. 18S and ATPase were the most and the least expressed reference gene, respectively (Fig 1D).

For the photoperiod experiment, the $C_t$ values ranged from 10 to 26. 18S, 28S, and Actin showed average $C_t$ values below 18 cycles. EF1A showed the $C_t$ values around 20, GAPDH and CypA were around 23. 18S and ATPase were the most and the least expressed reference gene, respectively (Fig 1E).

For the dietary RNAi experiment, the $C_t$ values ranged from 10 to 24. 18S, 28S, Actin, and EF1A showed average $C_t$ values below 20 cycles, GAPDH and CypA were around 22. The expression of ATPase was significantly depressed in the ATPase-dsRNA supplied samples compared to the water and GUS-dsRNA fed controls. 18S and ATPase were the most and the least expressed reference gene, respectively (Fig 1F).
Stability of candidate reference genes under biotic conditions

*GeNorm* bases its ranking on the geometric mean of the standard deviation (SD) of each transformed gene set of pair combinations (M-value). The lower the M-value is, the higher the
ranking. Under the impact of developmental stage, CypA and EF1A were co-ranked as the most stable genes. The overall order based on geNorm from the most stable to least stable reference gene was: CypA = EF1A, 28S, ATPase, 18S, GAPDH, Actin (Table 2). For the tissue experiment, 28S and GAPDH were co-ranked as the most stable genes. The overall order based on geNorm from the most stable to least stable reference gene was: 28S = GAPDH, CypA, EF1A, ATPase, 18S, Actin (Table 2). For the sex experiment, CypA and GAPDH were co-ranked as the most stable genes. The overall order based on geNorm from the most stable to least stable reference gene was: CypA = GAPDH, 28S, EF1A, ATPase, 18S, Actin (Table 2).

A low stability value suggests a more stable gene by NormFinder. Under the impact of developmental stage, ATPase was the most stable gene. The overall order based on NormFinder from the most stable to least stable reference gene was: ATPase, 28S, GAPDH, CypA, EF1A, 18S, Actin (Table 2). For the tissue experiment, 28S and GAPDH were co-ranked as the most stable genes. The overall order based on NormFinder from the most stable to least stable reference gene was: 28S = GAPDH, CypA, EF1A, ATPase, 18S, Actin (Table 2). For the sex experiment, GAPDH was the most stable gene. The overall order based on NormFinder from the most stable to least stable reference gene was: GAPDH, 28S, CypA, EF1A, ATPase, 18S, Actin (Table 2).

The stability of a gene is inversely proportional to the BestKeeper computed standard deviation (SD) value. Under the impact of developmental stage, EF1A was the most stable gene. The overall order based on BestKeeper from the most stable to least stable reference gene was: EF1A, CypA, 28S, ATPase, 18S, GAPDH, Actin (Table 2). For the tissue experiment, GAPDH was the most stable gene. The overall order based on BestKeeper from the most stable to least stable reference gene was: GAPDH, 28S, 18S, CypA, Actin, EF1A, ATPase (Table 2). For the sex

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experiment, GAPDH was the most stable gene. The overall order based on BestKeeper from the most stable to least stable reference gene was: GAPDH, CypA, 28S, ATPase, EF1A, 18S, Actin (Table 2).

The ΔCt method relies on relative pair-wise comparisons. Using raw Ct values, the average SD of each gene set is inversely proportional to its stability. Under the impact of developmental stage, 28S was the most stable gene. The overall order based on ΔCt method from the most stable to least stable reference gene was: 28S, ATPase, CypA, EF1A, GAPDH, 18S, Actin (Table 2). For the tissue experiment, GAPDH was the most stable gene. The overall order based on ΔCt method from the most stable to least stable reference gene was: GAPDH, 28S, CypA, EF1A, 18S, ATPase, Actin (Table 2). For the sex experiment, GAPDH was the most stable gene. The overall order based on ΔCt method from the most stable to least stable reference gene was: GAPDH, 28S, CypA, EF1A, ATPase, 18S, Actin (Table 2).

Under the impact of developmental stage, according to Reffinder, which integrates the above-mentioned four software tools to compare and rank the candidates, the comprehensive ranking of candidate reference gene from the most to the least stable was: 28S, EF1A, CypA, ATPase, GAPDH, 18S, Actin (Fig 2A). For the tissue experiment, the comprehensive ranking of candidate reference gene from the most to the least stable was: GAPDH, 28S, CypA, EF1A, 18S, ATPase, Actin (Fig 2B). For the sex experiment, the overall ranking from the most to the least stable reference gene in both adult females and males was: GAPDH, CypA, 28S, EF1A, ATPase, 18S, Actin (Fig 2C). To summarize, the overall ranking from the most to the least stable reference gene under the biotic condition was: 28S, CypA, EF1A, ATPase, GAPDH, 18S, Actin (Fig 2G).

Stability of candidate reference genes under abiotic conditions

Based on GeNorm, for the temperature experiment, 28S and EF1A were co-ranked as the most stable genes. The overall order based on geNorm from the most stable to least stable reference gene was: 28S = EF1A, ATPase, 18S, Actin, CypA, GAPDH (Table 3). For the photoperiod experiment, ATPase and GAPDH were co-ranked as the most stable genes. The overall order based on geNorm from the most stable to least stable reference gene was: ATPase = GAPDH, CypA, 28S, Actin, EF1A, 18S (Table 3). For the dietary RNAi experiment, Actin and GAPDH were co-ranked as the most stable genes. The overall order based on geNorm from the most stable to least stable reference gene was: Actin = GAPDH, CypA, 28S, 18S, EF1A, ATPase (Table 3).

Based on NormFinder, for the temperature experiment, EF1A was the most stable gene. The overall order based on NormFinder from the most stable to least stable reference gene was: EF1A, ATPase, 28S, 18S, CypA, Actin, GAPDH (Table 3). For the photoperiod experiment, GAPDH was the most stable gene. The overall order based on NormFinder from the most stable to least stable reference gene was: GAPDH, CypA, ATPase, 28S, Actin, EF1A, 18S (Table 3). For the dietary RNAi experiment, CypA was the most stable gene. The overall order based on NormFinder from the most stable to least stable reference gene was: CypA, Actin, 28S, GAPDH, 18S, EF1A, ATPase (Table 3).

Based on BestKeeper, for the temperature experiment, 28S was the most stable gene. The overall order based on BestKeeper from the most stable to least stable reference gene was: 28S, EF1A = ATPase, 18S, Actin, CypA, GAPDH (Table 3). For the photoperiod experiment, 28S was the most stable gene. The overall order based on BestKeeper from the most stable to least stable reference gene was: 28S, GAPDH, ATPase, EF1A, CypA, Actin, 18S (Table 3). For the dietary RNAi experiment, CypA was the most stable gene. The overall order based on BestKeeper from the most stable to least stable reference gene was: CypA, GAPDH, 18S, Actin, 28S, EF1A, ATPase (Table 3).
Fig 2. Stability of candidate reference genes expression under different treatment according to their stability value by RefFinder. A lower Geomean value indicates more stable expression.

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Based on \(\Delta C_t\) method, for the temperature experiment, \(EF1A\) was the most stable gene. The overall order based on \(\Delta C_t\) method from the most stable to least stable reference gene was: 
\[ EF1A, 28S, ATPase, 18S, CypA, Actin, GAPDH (Table 3) \]. For the photoperiod experiment, 
\[ GAPDH\] was the most stable gene. The overall order based on \(\Delta C_t\) method from the most stable to least stable reference gene was: 
\[ GAPDH, CypA, ATPase, 28S, Actin, EF1A, 18S (Table 3) \]. For the dietary RNAi experiment, 
\[ CypA\] was the most stable gene. The overall order based on \(\Delta C_t\) method from the most stable to least stable reference gene was: 
\[ CypA, Actin, 28S, 18S, GAPDH, EF1A, ATPase (Table 3) \].

Based on \(RefFinder\), for the temperature experiment, the comprehensive ranking of candidate reference gene from the most to the least stable was: 
\[ EF1A, 28S, ATPase, 18S, CypA, Actin, GAPDH (Fig 2D)\]. For the photoperiod experiment, the overall ranking from the most to the least stable reference gene under the photoperiod stress was: 
\[ CypA, GAPDH, ATPase, 28S, 18S, Actin, EF1A (Fig 2E) \]. For the dietary RNAi experiment, the overall ranking from the most to the least stable reference gene under the dsRNA stress was: 
\[ CypA, Actin, GAPDH, 28S, 18S, EF1A, ATPase (Fig 2F) \]. To summarize, the overall ranking from the most to the least stable reference gene under the abiotic condition was: 
\[ Actin, EF1A, 28S, CypA, GAPDH, 18S, ATPase (Fig 2H) \].

### Quantitative analysis of candidate reference genes based on \(geNorm\)

To decide the minimal number of reference genes mandatory for normalization, the V-value was computed by \(geNorm\). Beginning with two genes, the software continuously adds another
gene and recalculates the normalization factor ratio. If the added gene does not elevate the normalization factor ratio over the proposed 0.15 cut-off value, then the starting pair of genes is sufficient for the normalization. Otherwise, more genes should be incorporated. However, if the new ratio is above 0.15, then more genes should be included. For the developmental stage experiment, although none of the V-value was less than 0.15, the lowest value appeared at V3/4, suggesting that three reference genes were desirable for reliable normalization throughout developmental stages (Fig 3). For the tissue experiment, the first V-value < 0.15 emerged at V3/4, suggesting that three reference genes were required for reliable normalization in different tissue types (Fig 3). For the sex, temperature, photoperiod, and dietary RNAi experiments, the first V-value < 0.15 showed at V2/3, recommending that two reference genes were sufficient for reliable normalization (Fig 3).

Fig 3. Pairwise variation (V) analysis of the candidate reference genes. The geNorm first calculates an expression stability value (M) for each gene and then compares the pair-wise variation (V) of this gene with the others. A threshold of V<0.15 was suggested for valid normalization. Starting with two genes, the software sequentially adds another gene and recalculates the normalization factor ratio. If the added gene does not increase the normalization factor ratio above the proposed 0.15 cut-off value, then the original pair of genes is enough for normalization. However, if the new ratio is above 0.15, then more genes should be included [1].

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Discussion

RT-qPCR quantification requires a comprehensive normalization by reference genes to counteract confounding variation in experimental data. Generally considered to be expressed in all cell types of an organism at a constant level to maintain basic cellular functions, reference genes have been widely adopted as internal controls for various analyses involving gene quantification, including Western blot, Northern blot, and RT-qPCR. Each candidate reference gene, however, should be evaluated under specific experimental conditions for gene profiling to ensure a constant level of expression [23]. Our results demonstrate that the best suited reference genes can be different in response to different biotic and abiotic conditions. This is consistent with previous reports showing that reference genes are differentially expressed under specific experimental conditions in the sweet potato whitefly, *Bemisia tabaci* (Hemiptera: Aleyrodidae), diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae), brown planthopper, *Nilaparvata lugens* (Hemiptera: Delphacidae), beet armyworm, *Spodoptera exigua* (Hübner) (Lepidoptera: Noctuidae), oriental leafworm moth, *Spodoptera littoralis* (Lepidoptera: Noctuidae), oriental fruit fly, *Bactrocera dorsalis* (Diptera: Tephritidae), pea aphid, *Acyrthosiphon pisum* (Harris) (Hemiptera, Aphidiae), and Colorado potato beetle, *Leptinotarsa decemlineata* (Say) (Coleoptera: Chrysomelidae) [5, 6, 24–29]. Even under the same insecticide treatment, different classes of insecticides warranted different sets of reference genes to normalize target gene expression in *B. tabaci*, indicating the necessity for custom reference gene selection [30].

The large body of recent works clearly suggested that there are no “universal” reference genes that are stably expressed and applicable for all cell and tissue types and various experimental conditions [5, 6, 27–29]. For example, as a major component of the protein scaffold which supports the cell and determines its shape, *Actin* has been widely regarded as the universal reference gene and used extensively as the internal reference without any validation. For *H. convergens*, however, *Actin* was the least stable reference gene under different biotic conditions (developmental stage, tissue, and sex). Recently, Lin and Redies [31] compared histological expression profile of *GAPDH* and *Actin* in the developing chicken embryo. Neither *GAPDH* nor *Actin* was expressed in all cell types or tissues at high levels, and in some organs, these two reference genes exhibited partially complementary expression patterns. Specifically, *Actin* is highly expressed in the gizzard, but it was virtually non-existent in cardiac muscle cells.

There has been ongoing discussion about the optimal number of reference genes required for RT-qPCR analysis. Previously, gene expression studies have predominantly used a single endogenous control, however, this will profoundly influence the statistical outcome and may lead to inaccurate data interpretation [32] or it is simply insufficient to normalize the expression of target genes [33]. To avoid biased normalization, more and more researchers have moved away from a single endogenous control and started to embrace the idea of using multiple reference genes to analyze gene expression [5, 6, 27–29]. Determination of the optimal number of reference genes, however, has always being a trade-off between accuracy and practicality. A minimum of three most stably expressed reference genes is recommended by Thomas and colleagues [34]. In this study, two reference genes are sufficient to normalize the expression and provide a more conservative estimation of target gene expression under abiotic conditions, including temperature, photoperiod, and dietary RNAi. In contrast, three stable reference genes are generally required under biotic conditions, including developmental stage and tissue.

This is the first study to investigate candidate reference genes for gene expression analyses in the predatory species *H. convergens*. Based on a comprehensive analysis integrating five commonly used analytical methods to compare and rank the candidate reference genes under an array of biotic (developmental stage, tissue, and sex) and abiotic (temperature, photoperiod, and dietary RNAi) conditions, a suite of candidate reference genes were specifically
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Supporting Information
S1 Fig. Melting curve of the seven candidate reference genes. (TIFF)

S1 Table. Degenerate primers used for RT-qPCR. (DOCX)

S2 Table. The sequencing results of these five genes including Actin, EF1A, GAPDH, CypA, ATPase using the degenerate primers. (DOCX)

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Author Contributions
Conceived and designed the experiments: XGZ HPP XWY. Performed the experiments: XWY HPP. Analyzed the data: XWY HPP. Contributed reagents/materials/analysis tools: XGZ. Wrote the paper: HPP BDS XGZ.

References


