Programmable folding of fusion RNA \textit{in vivo} and \textit{in vitro} driven by pRNA 3WJ motif of phi29 DNA packaging motor

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Shu, Dan; Khisamutdinov, Emil F.; Zhang, Le; and Guo, Peixuan, "Programmable folding of fusion RNA \textit{in vivo} and \textit{in vitro} driven by pRNA 3WJ motif of phi29 DNA packaging motor" (2014). Pharmaceutical Sciences Faculty Publications. 17.  
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Notes/Citation Information
Published in Nucleic Acids Research, v. 42, issue 2, no. e10.

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Digital Object Identifier (DOI)
http://dx.doi.org/10.1093/nar/gkt885

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Programmable folding of fusion RNA in vivo and in vitro driven by pRNA 3WJ motif of phi29 DNA packaging motor

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Received July 30, 2013; Revised September 6, 2013; Accepted September 9, 2013

ABSTRACT

Misfolding and associated loss of function are common problems in constructing fusion RNA complexes due to changes in energy landscape and the nearest-neighbor principle. Here we report the incorporation and application of the pRNA-3WJ motif of the phi29 DNA packaging motor into fusion RNA with controllable and predictable folding. The motif included three discontinuous ~18 nucleotide (nt) fragments, displayed a distinct low folding energy (Shu D et al., Nature Nanotechnology, 2011, 6:658–667), and folded spontaneously into a leading core that enabled the correct folding of other functionalities fused to the RNA complex. Three individual fragments dispersed at any location within the sequence allowed the other RNA functional modules to fold into their original structures with authentic functions, as tested by Hepatitis B virus ribozyme, siRNA, and aptamers for malachite green (MG), spinach, and streptavidin (STV). Only nine complementary nucleotides were present for any two of the three 18-nt fragments, but the three 9 bp branches were so powerful that they disrupted other double strands with more than 15bp within the fusion RNA. This system enabled the production of fusion complexes harboring multiple RNA functionalities with correct folding for potential applications in biotechnology, nanomedicine and nanotechnology. We also applied this system to investigate the principles governing the folding of RNA in vivo and in vitro. Temporal production of RNA sequences during in vivo transcription caused RNA to fold into different conformations that could not be predicted with routine principles derived from in vitro studies.

INTRODUCTION

RNA nanotechnology involves programmable and addressable designs of RNA 3D nanoparticles by fusing or grafting individual RNA structural moieties or functionalities to one another (1,2). Many RNA folding programs are available (3–5) for RNA secondary structure prediction. However, manipulating tertiary folding of the reconstructed RNA is very challenging due to changes in the nearest-neighbor and the redistribution of energy landscapes. Development of systems to fold RNA functionalities in a controllable and predictable manner is therefore very desirable. It has been reported that RNA junction motifs, such as the three-way junction (3WJ) (6–9), four-way junction (4WJ) (10), and five-way junction (5WJ) (11) can form stable conformations composed of discontinuous RNA fragments. For example, the stable properties of the 3WJ structural motif derived from pRNA of bacteriophage phi29 DNA packaging motor has been utilized to generate functional RNA nanoparticles (6–8,12). Other examples of in silico generated RNA nanostructures based on RNA structural motifs include RNAI/II inverse kissing loops (13), kink-turn structural motifs (14,15), receptor-loop/loop-receptor interacting motifs (16,17), and phi29 pRNA hand-in-hand and foot-to-foot interactions (1,12,18–22). In addition, tightly folded RNA motifs have been reported in vivo that served as vectors to carry exogenous functional RNA modules (23).

More and more non-coding RNA molecules have been discovered to play essential roles in the regulation of a variety of diverse biological functions in cells (24–27). Often, the function of an RNA molecule is dependent on its tertiary structure, as well as information encoded in its canonical or non-canonical base pairing (28–30). Thus, prior to in vitro or in vivo design of artificial RNA nanoparticles, one needs to consider if the desired fusion RNA will fold into the predicted functional structure and retain its correct functionality. An example is the RNA paranemic motif coupled with malachite green (MG)
binding RNA aptamer (31) to confer rigidity and enhance fluorescence emission (6,7,8,32).

The structural properties encoded in intracellular functional RNAs are extremely important in RNA nanotechnology. Several methods have been applied to determine structural properties of small and large RNAs (33–37), including physical and chemical approaches to probe RNA/ribonucleoprotein structures in vitro (38). Additionally, several chemical reagents that are sensitive to secondary and/or tertiary structures have been used for elucidating RNA structure in vivo (34). Dimethyl sulfate (DMS) has been extensively applied for probing RNA structure in a variety of organisms ranging from bacteria to eukaryotes (39,40). Lead-(II)-acetate has also been used to probe RNA structures in bacteria (41); this ion easily enters bacterial cells and primarily induces specific cleavages at positions of tight metal ion binding. Another chemical reagent is the hydroxyl radical (OH), which has been widely used in nucleic acid footprinting (42). This technique can be applied to probe RNA tertiary structure and its intermolecular interactions.

Recently, we discovered that the pRNA-3WJ motif (Figure 1a) exhibits unusually robust properties (6,8,43) and can serve as a core to drive the folding of other RNA molecules fused to the complex. The pRNA-3WJ domain can be assembled from three pieces of RNA oligonucleotides resulting in a structure with: (i) an unusually high thermodynamic stability; (ii) highly efficient complex assembly even in the absence of metal salts; (iii) resistance to denaturation even in the presence of 8 M urea; (iv) the ability to stay intact without dissociating at ultra-low concentrations; and (v) coordination of two divalent metal ions (6,43). Herein, we demonstrate that this robust pRNA-3WJ motif can provide a leading core and drive the correct folding of other functionalities fused into the RNA complex with controllable and predictable consequences. This system will enable the production of fusion RNA with incorporated functionalities folded into their authentic structure and function for applications in biotechnology and nanotechnology.

MATERIALS AND METHODS

In vitro synthesis and purification of RNAs

All pRNA-3WJ constructs were prepared by in vitro transcription using T7 RNA polymerase from DNA templates amplified by polymerase chain reaction (PCR), as described previously (44). The DNA templates and primers were synthesized chemically by IDT (Iowa). RNAs were used directly for MG fluorescence assays, or purified by 8 M urea 8% polyacrylamide gel electrophoresis (PAGE).

Folate binding assay

Human nasopharyngeal carcinoma KB cells [American Type Culture Collection (ATCC)] were grown on glass coverslides in folate-free medium overnight. Cy3-labeled pRNA-3WJ-FA-siRNA(survivin)-Ribozyme was incubated with the cells at 37°C for 2 h. After washing with phosphate buffered saline (PBS), the cells were fixed by 4% paraformaldehyde and stained by Alexa Fluor® 488 phalloidin (Invitrogen) for cytoskeleton and TO-PRO®-3 iodide (642/661) (Invitrogen) for nuclei. The cells were then assayed for binding and cell entry by Zeiss LSM 510 laser scanning confocal microscope.

Assay for the silencing of genes in cancer cell model

KB cells were transfected with 25 nM of individual fusion pRNA-3WJ complex using Lipofectamine 2000 Transfection Reagent (Invitrogen). After 48 h, cells were collected and target gene silencing effects were assessed by western blot assay, as previously described (6). Briefly, cells were lysed by Radio-Immunoprecipitation Assay (RIPA) lysis buffer (Sigma) and the cell total protein was extracted for the assay. Equal amounts of proteins were then loaded onto 15% sodium dodecyl sulfate (SDS)–PAGE and electrophoretically transferred to Immun-Blot PVDF membranes (Bio-rad). The membrane was probed with survivin antibody (R&D) (1:4000 diluted) and β-actin antibody (Sigma) (1:5000 diluted) overnight, followed by 1:10 000 anti-rabbit secondary antibody conjugated with horseradish peroxidase (Millipore) for 1 h. Membranes were blotted by ECL kits (Millipore) and exposed to film for autoradiography.

Binding assay for the Streptavidin-binding aptamer

Tritium-labeled [3H] RNA nanostructures (pRNA-3WJ-MG-HBV-STV) were preassembled in binding buffer (PBS with 10 mM Mg2+) before incubation with streptavidin (STV) agarose resin (Thermo Scientific). A total of 50 µl aliquot of STV resin was equilibrated at room temperature (RT) following washing with binding buffer. In all, ~5 µg RNA samples were added to each tube and incubated with the resin for 1 h at RT. After incubation, the resin was spun at 500 x g for 1 min, and the supernatant (pass through) was removed. Then, 50 µl binding buffer was added to the mixture and incubated for 15 min to wash the resin several times. RNA was eluted by 5 mM biotin solution and samples were analyzed on a 1900 TR Liquid Scintillation Counter (Packard).

Hepatitis B virus ribozyme activity assay

Hepatitis B virus (HBV) substrate was labeled with Cy3 (Mirus Bio LLC) and incubated with RNA nanoparticle at 37°C for 1 hr in Tris buffer (50 mM Tris–HCl, pH = 7.5; 20 mM MgCl2, 20 mM NaCl). pRNA-HBV ribozyme was used as a positive control (45). After
incubation, the products were run in 8 M Urea, 10% PAGE gel for fluorescence imaging.

MG aptamer fluorescence assay

Gel-purified RNAs were mixed with 2 μM Malachite Green, trimethylmetane (MG) in binding buffer containing 100 mM KCl, 5 mM MgCl2 and 10 mM HEPES (pH 7.4) and incubated at RT for 30 min. The refolded RNA samples were heating to 95°C for 5 min followed by slow cooling to 37°C prior to MG staining. Fluorescence was measured using a fluorospectrometer (Horiba Jobin Yvon; SPEX Fluolog-3), excited at 615 nm, scanning from 625 to 800 nm for emission (7,32,46).

Spinach fluorescence measurements

Assembled pRNA-3WJ nanoparticles (0.2 μM) fused with Spinach aptamer in TMS buffer were mixed with DFHBI dye (2 μM) and incubated at room temperature for 30 min. Fluorescence was measured using a fluorospectrometer (Horiba Jobin Yvon), excited at 450 nm and scanning from 565 to 750 nm for emission.

Vector construction for in vivo expression

A sequence of cis-ribozyme was fused onto the 3'-end of DNA templates of corresponding RNA for terminal processing (45,46). The S′+0 (Reference), S′+6, S′+12, and S′+15 were inserted between BglII/NdeI sites of expression vector pET-3b. BglII cleavage removed the original T7 promoter in the vector to prevent any undesired sequence in the 5'-end of the RNA product. The insertion fragment contained the T7 promoter at the 5'-end. The clone was transformed into Escherichia coli strain DH5α and recombinant plasmids were verified by sequencing (GENEWIZ).

RNA in vivo expression and purification

Escherichia coli strain BL21 star (DE3; Invitrogen) was transformed by the recombinant plasmids, and the colony was inoculated by 5 ml LB medium containing 100 μg/ml ampicillin, grown at 37°C, and shaken at 250 rpm until A600 nm reached 0.5. IPTG solution of 50 μl (1 M) was added to 5 ml of cell culture. Cells were allowed to grow for 1.5 h, and then were pelleted and resuspended in 250 μl of 10 mM magnesium acetate, 1 mM Tris–HCl, pH 7.4 (47). The total soluble RNAs were extracted using 500 μl of water saturated phenol (pH 4.5; Fisher). The aqueous phase was ethanol precipitated then dissolved in 50 μl of 0.05% DEPC treated water.

Gel electrophoresis of in vivo prepared RNAs

In vivo expressed S′+0 (reference), S′+6, S′+12, and S′+15 RNAs were analyzed by 8% PAGE gel containing 8 M of urea by loading each of sample and run in 1x TBE (89 mM Tris base, 89 mM Boric acid, 2 mM ethylenediaminetetraacetic acid) at 120 V, at RT for 1 h. The 8% native PAGE gel was run in 1x TBM (89 mM Tris base, 200 mM Boric acid, 5 mM MgCl2, pH 8.0) at 70 V, at 4°C for 3 h. After electrophoresis, the gels were stained with 10 μM MG in 100 mM KCl, 5 mM MgCl2, and 10 mM HEPES (pH 7.4) for 15 min at RT. The MG fluorescence image was acquired by the Cy5 channel (635 nm excitation/670 nm emission) using a Typhoon scanner (48). The gels were then stained with EB and scanned in EB channel (532 nm excitation/580 nm emission).

RESULTS

pRNA-3WJ motif drives the folding of different RNA functionalities in fusion RNA

Figure 1b shows the design principles for constructing fusion RNA with multiple functionalities. The three pRNA-3WJ fragments (a3WJ, b3WJ, and c3WJ) are dispersed within the complex and serve as a driving force for the folding of other RNA molecules. Several RNA functionalities were incorporated and tested in vitro including: (i) siRNA targeting survivin gene (49,50); (ii) MG (Malachite Green dye, triphenylmethane) binding RNA aptamer (51); (iii) Spinach RNA aptamer (52); (iv) STV binding RNA aptamer (53); and (v) HBV ribosome (45). These RNA moieties are unique in their sequences and folding properties, and exhibit different Gibbs free energies (ΔG). When the functional therapeutic RNA moieties, i.e. siRNA, ribozymes or receptor-binding aptamers, were fused to any of the three branches of the pRNA-3WJ, the 3WJ and the incorporated RNA modules folded independently, as validated by AFM image (Figure 2a), and retained their authentic functions, as demonstrated by functional assays (Figure 2). The location of the three 3WJ fragments in the fusion RNA was not critical; but the order of appearance from S′-a3WJ, to b3WJ, to c3WJ -3' (Figure 1b) was the essential factor.

Assessment of cell binding and gene silencing using fusion pRNA-3WJ-FA-siRNA (survivin)-Ribozyme nanoparticles

A pRNA-3WJ complex was coupled with folate (FA) ligand, survivin siRNA, and ribozyme, denoted as pRNA-3WJ-Folate-siRNA(survivin)-Ribozyme nanoparticle (Figure 2a). FA serves as a cancer cell delivery agent via receptor mediated endocytosis (6,8,54). Cy3-labeled pRNA-3WJ complex was incubated with FA-receptor positive KB cells and tested for cell binding efficiency. Confocal microscopy indicated strong binding of RNA nanoparticles as demonstrated by excellent co-localization of cytoplasm (green) and RNA nanoparticles (red) (Figure 2b).

Western blot was performed to assay the gene silencing effects of survivin siRNA in KB cells. After 48-h transfection, reduced survivin protein expression level was observed compared to scramble controls (Figure 2c). β-actin was used as an endogenous control. We previously demonstrated that the mechanism of siRNA release from pRNA nanoparticles is by Dicer processing (21).

Assessment of aptamer functionalities using fusion pRNA-3WJ-MG-Spinach nanoparticles

MG binding RNA aptamers (51) and Spinach RNA aptamers (52) were fused to the pRNA-3WJ scaffold (denoted pRNA-3WJ-MG-Spinach nanoparticles) for structure and function verification (Figure 2d). Free
MG and DFHBI (3,5-difluoro-4-hydroxybenzylidene imidazolinone; Spinach aptamer-binding dye) are not fluorescent by themselves, and only emit fluorescence after binding to the correctly folded RNA aptamers. Both MG binding aptamer and Spinach aptamers incorporated in the 3WJ motif retained their capacity to bind MG and DFHBI dyes, respectively, as demonstrated by their fluorescence emission in native PAGE (Figure 2) and in solution (Figure 2f).

Assessment of ribozyme catalytic activity and STV binding using fusion pRNA-3WJ fragments

Tetravalent RNA nanoparticles were constructed using pRNA-3WJ motif harboring MG binding aptamer (6,8,32), HBV ribozyme (45), and STV aptamer (53,48) (denoted pRNA-3WJ-MG-HBV-STV nanoparticles) (Figure 2g). HBV ribozyme (45) was able to cleave its 135-nt RNA genome substrate into two fragments (60 nt and 75 nt) (Figure 2h). The results are comparable to optimized positive controls, thereby confirming that the fused ribozyme retained its authentic structure and function. MG aptamer also retained its capacity to bind MG dye, as revealed by fluorescence emissions in urea PAGE (Figure 2i) and verified by total RNA staining with EB (Figure 2j).

For assessment of STV binding, [3H]-UTP whole chain labeled pRNA-3WJ-MG-HBV-STV nanoparticles were incubated with STV resin in an affinity column. The column was washed, and the RNA nanoparticles were then eluted with biotin (Figure 2k), indicating correct folding of the incorporated STV aptamer. As a negative control, RNA nanoparticles without STV aptamer did not appear in the elution fractions.

Design principles for evaluating the power of three pRNA-3WJ fragments to override other complementary sequences within fusion RNA constructs

The energy landscape and nearest-neighbor principle are key factors governing RNA folding. Due to the free-energy minimization principle, complementary sequences within RNA also play a critical role in the formation of RNA 3D structures, e.g. pseudoknots, multi-way junctions, and hairpins. To apply the pRNA-3WJ for fabricating RNA nanostructures, the power of three pRNA-3WJ fragments to override other complementary sequences within fusion RNA complexes have to be tested. Accordingly, fusion RNA complexes were designed based on: (i) three fragments of pRNA-3WJ to direct the folding of the RNA; (ii) additional 5'-end interfering sequences complementary to the internal sequence of the 3WJ; and (iii) MG-aptamer to serve as a reporter for verification of authentic folding (7). If the RNA is degraded or misfolded, the MG fluorescence will disappear, and therefore can be used as an indicator for assessing the folding of fusion RNA complexes.

RNA constructs containing interfering sequences have a designated nomenclature. For instance, ‘5’ + 12 means that 12 overhanging nucleotides complementary to the MG-aptamer sequence were added at the 5’-end of the RNA complex. Similarly, ‘5’N+12 (NM = non-matching) means that 12 nucleotides non-complementary to the MG-aptamer sequences were added at the 5’-end of the RNA complex. All RNA nanostructures were constructed to contain the pRNA-3WJ and an MG aptamer (Figure 3a). Because the sequence at the 5’-end appeared earlier during transcription than the sequence at the 3’-end, single-stranded (ss) nucleotides of different lengths (6, 12, and 15 nt) were added to the 5’-end, complementary to the MG-aptamer sequence, in order to interfere with the original folding driven by the pRNA-3WJ (Figure 3b). Several controls were designed in which overhanging NM nucleotides of differing lengths complementary to the MG-aptamer sequence were added (Figure 3c). All these
RNA constructs contain discontinuous motifs, such as those observed in a pseudoknot (55–59).

We then investigated whether the folding was affected by the discontinuous motifs prior to or after refolding subsequent to transcription. This is particularly important in the context of in vivo folding of RNA and for transcribing therapeutic RNA nanoparticles directly in the cell with unprocessed additional sequences at the 5′-end (top panel) and corresponding fluorescence spectra (bottom panel). In (b) and (c), black and red curves represent fluorescence spectra after transcription and after refolding, respectively.

**Comparison of pRNA-3WJ folding after transcription and after refolding**

To unravel the folding differences of in vitro RNA transcription products before and after refolding, pRNA-3WJ-MG emission spectra were measured. RNA transcription reactions were performed, as described previously (60). The reactions were stopped by the addition of DNase I. Fluorescence spectra of the transcribed pRNA-3WJ-MG complex was then measured in the presence of MG fluorophore. The pRNA-3WJ-MG complex was then heated to 95°C and slowly cooled to 37°C (refolding). The fluorescence spectra of the refolded pRNA-3WJ-MG complex was then compared with the transcribed complex.

The fluorescence intensity was similar for all the control RNA samples, 5′ + 0, 5′NM + 6, 5′NM + 9, and 5′NM + 12, regardless of the refolding step (Figure 3c). However, RNA constructs 5′ + 12 and 5′ + 15 with 12 and 15 overhanging nucleotides, respectively, showed significant differences (Figure 3b). Upon increasing the ssRNA length at the 5′-end from 6 to 12nt, as exemplified in molecule 5′ + 12, the fluorescence intensity after transcription was found to be lower than after refolding. Increasing the length of the ssRNA to 15nt in RNA 5′ + 15 resulted in a larger difference (Figure 3b). These results demonstrate that the complementary interfering sequences at the 5′-end partially disrupted the folding of pRNA-3WJ assembly during the transcription reaction. In contrast, if the 15-nt complementary interfering sequence is instead placed at the 3′-end, no differences in fluorescence intensity are observed (Figure 3C, complex 3′ + 15), which is consistent with our interpretations.
In vivo assessment of the effect of temporal folding after transcription

RNA constructs with interfering sequences of 6, 12, and 15 nt, similar to the in vitro design of the 5’+0, 5’+6, 5’+12, and 5’+15 RNA were expressed in E. coli cells. The principle of this design was based on the following ideas: (i) a thermodynamically stable and tightly folded pRNA-3WJ-MG aptamer would drive the correct folding of the MG-aptamer; (ii) tightly folded RNA are relatively resistant to RNase degradation in the cell (most cellular RNases degrade ssRNA, but not dsRNA) and misfolded RNA would be trimmed or truncated by RNase, leading to a loss in fluorescence from the MG dye that only binds to the appropriately folded and intact MG aptamer; and (iii) the assay would be based on the comparison of the sequences that affect the folding of the core of the pRNA-3WJ-MG aptamer. Thus, a cis-acting ribozyme sequence was placed at the 3’-end to cleave the undesirable sequences arising from uncontrolled termination of transcription at the 3’-end (Figure 3a) (45,61).

The RNA products from E. coli cells were first assayed by 8% denaturing PAGE (Figure 6b). As reported previously, the pRNA-3WJ is resistant to 8 M urea denaturation (6,8), and as such, denaturing gel can be used to detect the presence of the pRNA-3WJ-MG aptamer complex. Upon staining with either ethidium bromide (EB) (Figure 6b, top) or MG dye (Figure 6b, bottom), the 5’+0 RNA, serving as a reference molecule, and the 5’+6 RNA were detected on the gel as distinct bands around 200 bp, indicating the presence of both the pRNA-3WJ and the MG aptamer in both constructs. The higher yield of 5’+0 RNA compared to 5’+6 construct indicated that the 6 complementary nucleotides at the 5’-end of 5’+6 were able to disrupt the folding of the pRNA-3WJ-MG aptamer complex only to some extent. However, MG-stained RNA was not detected in both 5’+12 and 5’+15 RNA lanes, indicating the misfolding of those pRNA-3WJ-MG aptamer complexes. Several degraded products were observed between 50 and 70 bp after EB staining (Figure 6b, top, lanes 5’+12 and 5’+15). This data agreed with the in vitro data, suggesting that RNA folding is temporally dependent during transcription, and that the 5’-end sequence appearing earlier during transcription influenced the RNA folding more than the sequence that appeared later during transcription.

To verify the denaturing PAGE results, fluorescence intensities of the RNA constructs obtained in vitro were measured further in presence of MG dye (Figure 6e), in a manner similar to that of the in vitro experiments (Figure 3). The fluorescence intensities were detected for both 5’+0 and 5’+6 constructs (Figure 6c, black curves). But, the 5’+0 RNA showed higher fluorescence value than the 5’+6 construct with six interfering nucleotides. There was almost no fluorescence detected for constructs 5’+12 and 5’+15. These data are consistent with MG-stained denaturing PAGE, suggesting that while six interfering nucleotides partially disrupted the folding of pRNA-3WJ-MG aptamer, 12 and 15 nt complementary

similar ‘r’ values close to 1, indicating the effect was not dependent on nucleotide length (Figure 5b). However, in the presence of overhanging nucleotides that interfered with the MG aptamer sequence there was a correlation: the longer the additional sequences at the 5’-end, the smaller the ‘r’ value, implying that there are fewer pRNA-3WJ-MG constructs that folded properly during temporal transcription (Figure 5a). The data leads to the conclusion that the RNA folding was temporally dependent. The 5’-end sequences that appeared earlier during transcription influenced the RNA folding more than the sequences that appeared later during transcription.
sequences completely interfered with the formation of pRNA-3WJ-MG-aptamer structures in vivo.

To refold the in vivo RNA products of pRNA-3WJ-MG-aptamer, the denaturation and refolding step was performed by heating the RNA solution to 95°C and slowly cooling to 37°C. Interestingly, the measured fluorescence intensities were significantly reduced or were not detected at all (Figure 6c, red curves). These data are in sharp contrast to that obtained from the in vitro experiments. The plot of ratio versus nucleotide length from the in vitro RNA folding experiment resulted in a descending trend (Figure 5a), which was not observed for in vivo samples. The results suggest that RNA folding in vivo and in vitro are not always identical. Correctly folded 5' +12 or 5' +15 RNA were significantly recovered after denaturation and refolding (Figure 3b). However, the misfolded in vivo 5' +12 and 5' +15 RNA constructs, if any, could not be recovered upon refolding (Figure 6c).

Comparison of the sequences with their complementary partners

The folding properties of discontinuous 3WJ-pRNA were further analyzed by a prediction algorithm using free energy minimization parameters and mfold (4). The goal was to investigate how different lengths at the 5’-end contributed to the disruption of the 3WJ-pRNA constructs, and whether there was a relationship between this influence and the predicted free energy of the added sequences on their partners. The following free energies for the duplexes of 5’- or 3’-end overhanging sequences and their complementary partners were calculated at 10^-4 M RNA in the presence of 1 M NaCl:

\[
\Delta G = -6.1 \text{kcal/mol for the 5'} + 6 \text{ RNA}
\]

\[
\Delta G = -19.4 \text{kcal/mol for the 5'} + 12 \text{ RNA}
\]

\[
\Delta G = -25.6 \text{kcal/mol for the 5'} + 15 \text{ RNA}
\]

\[
\Delta G = -26.6 \text{kcal/mol for the 3'} + 15 \text{ RNA} \text{ (control)}
\]

The longest construct with a 15-nt complementary sequence had the highest stability with a \( \Delta G \) of \(-25.6 \text{kcal/mol} \). This indicates that the folding of the 3WJ-pRNA complex 5’ +15 during in vitro transcription was strongly interfered. This theoretical data was consistent with the correlation observed by fluorescence measurements of ‘r’ values. The 3’-end interfering sequences resulted in the lowest free energy value, \( \Delta G = -26.6 \text{kcal/mol} \). This sequence did not interfere with the MG fluorescence signal since the transcription reaction occurs from the 5’ to the 3’-end, and the 15-nt region was produced after the 3WJ complex had folded.

**DISCUSSION**

With the recent emergence of RNA nanotechnology (2), development of new systems to investigate the folding of RNA nanostructures has become very desirable. It has been reported that RNA junction motifs, such as 3WJs, 4WJs, and 5WJs can form rigid structures composed of discontinued RNA fragments (6, 8,10,11). The application of robust RNA motifs as vectors to carry exogenous functional moieties has been reported (23,6,8,12,48). The novelty here is not the application of the RNA motif as vector, but the use of the unique pRNA-3WJ with favorable thermodynamic attributes to drive the assembly of fusion RNA complexes (6,8). The energy landscape in RNA folding remains fundamentally important. When many motifs with similar levels of folding energies are fused into one large RNA molecule, new complementary sequences will appear and redistribution of energy landscape following the nearest neighboring principle will occur.

The pRNA-3WJ motif can be assembled from three pieces of RNA oligonucleotides with a \( T_M \) slope close to 90° (6,8). The resulting three-component complex was resistant to 8 M Urea denaturation and does not dissociate.
at extremely low concentrations (6,8). The three components had a much higher affinity for favorable interactions compared to any of the two components, indicating cooperative simultaneous folding of the three helical stems. The incorporation of a pRNA-3WJ with distinct low folding energy will provide a leading motif to drive the folding of other functional motifs with weaker folding forces. Thus, functional modules with weaker folding forces will be able to fold independently into their original authentic structure when linked to the branches of the pRNA-3WJ.

CONCLUSIONS

The thermodynamically ultrastable pRNA-3WJ motif of phi29 motor pRNA can be used to drive the folding of fused RNA molecules with controllable and predictable consequences. When a large fusion RNA molecule is constructed containing three individual 18-nt fragments dispersed at any location within the sequence, the three fragments drive the correct folding of other RNA functionalities or motifs into their original 3D structures with authentic functions. This approach offers a new system for future investigations into the important topic of RNA folding in vivo.

ACKNOWLEDGEMENTS

We thank Dr. Farzin Haque for insightful comments and Jeannie Haak for assistance in preparing this article.

FUNDING

National Institutes of Health (NIH) grants [EB003730 and CA151648 to P.G.] and The Arnold and Mabel Beckman fund [1210 to D.S.]. Funding to Peixuan Guo’s Endowed Chair in Nanobiotechnology is by the William Fairish Endowment Fund. P.G. is a cofounder of Kylin Therapeutics, Inc., and Biomotor and Nucleic Acid Conflict of interest statement to P.G.

Nanotechnology Development Corp. Ltd. Funding for Therapeutics, Inc., and Biomotor and Nucleic Acid Fairish Endowment Fund. P.G. is a cofounder of Kylin Chair in Nanobiotechnology position is by the William Fairish Endowment Fund. P.G. is a cofounder of Kylin Therapeutics, Inc., and Biomotor and Nucleic Acid Conflict of interest statement to P.G.

National Institutes of Health (NIH) grants [EB003730 and CA151648 to P.G.] and The Arnold and Mabel Beckman fund [1210 to D.S.]. Funding to Peixuan Guo’s Endowed Chair in Nanobiotechnology position is by the William Fairish Endowment Fund. P.G. is a cofounder of Kylin Therapeutics, Inc., and Biomotor and Nucleic Acid Nanotechnology Development Corp. Ltd. Funding for open access charge: NIH [EB003730 and CA151648 to P.G.].

Conflict of interest statement. P.G. is a cofounder of Kylin Therapeutics, Inc. and Biomotor and Nucleic Acid Nanotechnology Development Corp. Ltd.

REFERENCES


