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Monoclonal Antibody 1A7 and Use for the Treatment of Melanoma and Small Cell Carcinoma

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The present invention relates to monoclonal antibody 1A7. This is an anti-idiotypic produced by immunizing with an antibody specific for ganglioside GD2, and identifying a hybridoma secreting antibody with immunogenic potential in a multi-step screening process. Also disclosed are polypeptide and polypeptide derivatives based on 1A7, including single chain variable region molecules and fusion proteins, and various pharmaceutical compositions. When administered to an individual, the 1A7 antibody overcomes immune tolerance and induces an immune response against GD2, which comprises a combination of anti-GD2 antibody and GD2-specific T cells. The invention further provides methods for treating a disease associated with altered GD2 expression, particularly melanoma, neuroblastoma, glioma, soft tissue sarcoma, and small cell carcinoma. Patients who are in remission as a result of traditional modes of cancer therapy may be treated with a composition of this invention in hopes of reducing the risk of recurrence.

5 Claims, 24 Drawing Sheets
Cheresh, D.A. et al. (1984). "Localization of the Gangliosides GD2 and GD3 in Adhesion Plaques and on the Surface of Human Mela-


Cheung, N-K.V. et al. (1994). "Antibody Response to Murine Anti-


* cited by examiner
Figure 1

M K L P V R L L V L M F W I P A
ATG AAG TGG CCT GTT AGG CTG TTG GTG ATG TTC TGG ATT CCT GCT
S S D
TCC AGC GAT (-1 to -19, leader)

D V L M T Q T P L S L P V S L G
GAT GTT TTG ATG ACC CAA ACT CCA CTC TCC CCT CTG GTC AGT CTT GGA
D Q A S I S C
GAT CAA GCC TCC ATC TCT TGC (1-23, Frame work 1)

R S S Q S I V H S N G N T Y L E
AGA TCT AGT CAG AGC ATT GTA CAT AGT AAT GGA AAC ACC TAT TTA GAA
(24-39, CDR 1)

W Y L Q K P G Q S P N L L I Y
TGG TAC CTA CAG AAA CCA GGC CAG TCT CCA AAC CTC CTG ATC TAC
(40-54, Frame work 2)

F V S N R F S
TTT GTT TCC AAC CGA TTT TCT (55-61, CDR 2)

G V P D R F S G S G S G T D F T
GGG GTC CCA GAC AGG TTC AGT GGC AGT GGA TCA GGG ACA GAT TTC ACA
L K I S R V E A E D L G V Y Y C
CTC AAG ATC AGC AGA GTG GAG GCT GAG GAT CTG GGA GTT TAT TAC TGC
(62-93, Frame work 3)

F Q G S H V P W T
TTT CAA GGT TCA CAT GTT CCG TGG ACG
(94-102, CDR 3)

F G G T K L E I K
TTC GTG GGA GGC ACC AAG CTG GAA ATC AAA
(103-112, Frame work 4)

R A D A A P T V S I F P P
CGG GCT GAT GCT GCA CCA ACT GTA TCC ATC TCC CCA CCA
S S K L G
TCC AGT AAG CTT GGG (Constant region)
Figure 2

MAV LGL LFL VTV FPS C
ATG GCT GTC TTG GGG CTG CTC TTC TGC CTG GTG ACA TTC CCA AGC TGT
VLS
GTC CTG TCC (-1 to -19, Leader)

QVKESGPFLVPSPQ
CAG GTG CAG CTG AAG GAG TCA GGA CCT TTC CTG GTG CCC CCC TCA CAG
SLSITCTVSFSLT
AGC CTG TCC ATC ACA TGC ACT GTC TCA GGG TTC TCA TTA ACC
(1-30, Frame work 1)

TVGS
ACC TAT GGT GTA AGC (31-35, CDR 1)

WIRQPPLKGLEWLG
TGG ATT CGC CAG CCT CCA GGA AAG GTT CTG GAG TGG CTG GGA
(36-49, Frame work 2)

AIWGDGTNTYNHSALIS
GCA ATT TGG GGT GAC GGG ACC ACA AAT TAT CAT TCA GCT CTC ATT TCC
(50-65, CDR 2)

RLSISKDNSKSSQVFLK
AGA CTG AGC ATC AGC AAG GAT AAG TCC AAG AGC CAA GTT TTC TTA AAA
LNSLTDDTDATAYTYCAK
CTG AAC AGT CTA GAA ACT GAT GAC AGC AGC GCC AGC AGC TAC TAC TGT GCC AAA
(66-97, Frame work 3)

LGNYDALDWC
CTG GGT AAC TAC GAT GCT CTG GAC TAC
(98-106, CDR 3)

WGQGTSVTVSS
TGG GGT CAA GGA ACC TCA GTC ACC GTC TCC TCA
(107-117, Frame work 4)

AKTTPPPVYPLVPGSL
GCC AAA AGC ACA CCC CCA CCC GTC TAT CCA TTG GTC CCT GGA AGC TGT GG
(Constant region)
Figure 3(A)

1A7:  

1  DVLMTQTPLSLPV5LGDAQASICRSSQSVH5NGNTYLEWLYLQPQGSPHNLILYFYVSNRF  50

1  K K  60
2  K K  60
3  K K  60
4  K K  60
5  K K  60
6  K K  60
7  K K  60
8  X K K  60
9  S . . . . . . . . . . . . . . . F . K K  64
10  K K  60
11  K K  60
12  K K  79
13  K K L  60
14  K K  60
15  S . . . . . . . . . . . . . . . F . K K  64

1A7:  

1  SGVPDRTSGSGLDFTLKISYVEAEDLGVYVCQGSHVPWTFGSGKLEIK  112

1  112
2  112
3  112
4  111
5  X 112
6  Y 112
7  C Y 111
8  Y 111
9  T Y 116
10  R Y 112
11  R 112
12  Y S 131
13  Y 112
14  W Y 112
15  Q T 116
Figure 3(B)

1A7:  

1  QVQVKEGFLVPSLITCTVGFLTTYGVSIRQPGKLGAEWLGQSDGTTNYH  60
2  .G.A..................S........V..............V........S........  52
3  1  .Q...G.A.............S..IT.V..............V........N........  60
8  23  .LQ...G.A.............G..N.V.............M...N.D.N  82
9  1  .L...G.A.............G..N.V.............M...N.D.N  60
10  133  .LQ...G.A.............G..N.V.............M...N.D.N  192
12  1  .L...G.A.............SR.S.H.V.............M...G.N.D.N  60
14  23  .LQ...G.A.............G..N.V.............M...N.D.N  82
15  1  .LQ...G.A.............G..N.V.............M...N.D.N  60

1A7:  

61  SALISRLSISKDNKQVFLKMSLQDOTATYCAKL---------------GNYDALMWCGSIVS  117
1  53  .---------------P-------YDYExxxxx.Y....TL..  109
2  61  .---------------X-------xxxxxxx.K.Y.............  120
3  80  .T.K. T.T...........M........R...SVSIYYGRSDK.FT.Y.............  144
5  61  .H..................M........H........R..........xxxxxxx.Y.H.Y.............  120
6  61  .H..................M........H........R..........xxxxx.Y.M.Y.............  118
7  61  .H..................M........H........R..........xxxxx.X.Y.M.Y.............  119
8  83  .K..................M........H........R..........RDYR.Y....T.............  138
9  61  .K..................M........H........R..........RDYR.Y....TL.............  116
10  193  .K..................M........H........R..........RDYR.Y....T.............  248
11  80  .K..................M........H........R..........RDYR.Y....TL.............  135
12  61  .K..................M........H........R..........GYYDX.M.Y.............  117
13  81  .M..................M........I........I..........xxxxx.Y.M.Y.............  139
14  83  .K..................M........H........R..........RDYR.Y....T.............  138
15  61  .K..................M........H........R..........RDYR.Y....T.............  116
**Figure 3(C)**

**VL consensus:**  
1 DVLHTQTPSLPVSLGDQASIRCRISSQISVSHSGNTYLEWQLQKQQPSKLLYFVSNRF  
1A7:  
1 ........................................P..........................N...............  

**VL consensus:**  
61 SGVPDRFSGSGGTDFTKISRVEAEDLGYYCFQGSHVPFWGKGTLKLEIK  
1A7:  
61 ..................................................  

**VH consensus:**  
1 QVQLKESGPGLVAPSQSLSITCTVSFGSFLTSYGYMVHRQPQPKFLLEFLVIMGDSNYN  
1A7:  
1 ........................................T...........S..................A...........T...  

**VH consensus:**  
61 SALKSRLSISKDNSQSQVLKKNLQTDARYYCAReXXXYYAHDFWQGTSVTSS  
1A7:  
61 ........................................T...........KL--GN.D.L.W...............  

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Figure 4

- PRO#685
- PRO#778

(cpm bound vs. μg Ab3 ADDED)
Figure 5

![Graph showing % inhibition vs μg Ab3 added]
Figure 6

% inhibition vs. \( \mu g \) Ab3/Ab1 added
Figure 7(A)

Fluorescence Intensity [log]

Relative number of cells
Figure 7(B)

relative number of cells

Fluorescence Intensity [log]
Figure 7(C)

relative number of cells

Fluorescence Intensity [log]

100

50

10^1 10^2 10^3 10^4
Figure 8
Figure 9

% inhibition

μg Ab3/Ab1 added

5 2.5 1.25 0.6 0.3 0.15 0.075 0.04
Figure 11
Figure 12

![Bar chart showing OD 405 nm for three patients: Patient #1, Patient #2, and Patient #3.]

- OD 405 nm for Patient #1:
- OD 405 nm for Patient #2:
- OD 405 nm for Patient #3:
Figure 13

% Inhibition vs. μg Ab3/Ab1

% Inhibition vs. μg Ab3
Figure 15

GCCGATATCACCCATATGGCTGTCTTGGGGCTGCTCTCTGTGCTTCGCTGGTGACATTCCCAAGGC
TGTTCTGGCTCCAGGCTGACGAGGATCAAGCCTCATTCTGCTCTGACCGGTGACTATTGCTATA
AGCTGGATTCCAGCCACCTCCAGGAAAGGGTCTGGAGTGGCTGGAGCAATTGGGG
TGACGGGACGACAAGATTATCATCCAGTCTCTCATATCCAGACTGACATCGACAAAGGA
TAATCAGCAAGACCAAAGTATTTCTTTAAACTGAACAGTCTGCAAACTGAGTCACGACGC
CACGTACTACTGTCGCAAAACTGGTAAACTACGATGCTCTGGACTACTGCTGGGTCAAGG
AACCTGGTCACCCCGTCTCCACGGGAGGAGGTGCTGCGCCTGGGCTGGGTGGGCTGGGG
CGGGGATCCGATTTTGATGACCCCAAACCTCCACCTCTCCCTGCTGCTGATCTCTGGGA
GATCAAGCCTCCACCTCTCTGGGAGGATCTAGTCAAGAGGAATTGGAATACTGACGAC
ACCTATTAGAATGGTACCTACAAGCAACCCAGGCGACTCCCTCTCAGTACAT
TTTGGGGAACGATTTTCGCTGGGCTCCAGAACGGTGTCAGTGCCAGTGGATCAGGG
ACAGATTTCCGACTCAAGATCGACGAGTGAGGCTGAGGATCTTGGGAGTTATTAC
TGCTTTCAAGGGTCACATGGTCGAGCGCCGAGCTGAGGACCAAAGCTGGAAATCT
AAATAATTCTAGAGATG

1  mavglfllcl  vtfpscvelsq  vqvkesgplf  vppsqslsit  ctvsgfsltt
51  ygyvswirqpp  gkglewlqai  wgddgtnyhs  alisrlsik  dnsksqvfllk
101  lnslqtdlda  tyycaklgnvy  dalwywggt  svtvssgggg  sgggsqgggg
151  sdvlmlqtpql  slpvsldgqda  siscrsqsqi  vhsngntyle  wylqkpgasp
201  nniliyfvsrne  fsgvpgdrgsg  sgsqtdfllk  isrvaeadlg  vyyctqfgshv
251  pwtfgggtkl  eik
Figure 16

pH1A7
4519 bps
Figure 17(A)

>gb|L22327|HUS1GKAVAA Mouse rearranged immunoglobulin kappa-chain mRNA V-J

1 GATTTTTTAGACCGAAGTGCTCTCCTGATTGAGACTGCTCTCCCTGCTGCT 60
61 ATCTGTTCCTATCAATGACACATTCTCTCATCTGCTCAATGATATG 120
121 TACCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCT 180
181 TCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCTGCT 240
241 AGACAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 300
301 TGAGCCTTCGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 336

>gb|L18941|HUS1GKAVAA Mouse rearranged immunoglobulin light chain Ab438 mRNA V-J

1 GATTTTTTAGACCGAAGTGCTCTCCTGATTGAGACTGCTCTCCCTGCTGCT 60
61 ATCTGTTCCTATCAATGACACATTCTCTCATCTGCTCAATGATATG 120
121 TACCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCT 180
181 TCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCTGCT 240
241 AGACAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 300
301 TGAGCCTTCGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 336

>gb|K34588|HUS1GKXBR Mouse Ig kappa-chain mRNA V-J region, partial cds.

1 GATTTTTTAGACCGAAGTGCTCTCCTGATTGAGACTGCTCTCCCTGCTGCT 60
61 ATCTGTTCCTATCAATGACACATTCTCTCATCTGCTCAATGATATG 120
121 TACCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCT 180
181 TCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCTGCT 240
241 AGACAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 300
301 TGAGCCTTCGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 336

>gb|K32857|HUS1GKXSP Mouse Ig rearranged kappa-chain mRNA V-region, partial

1 GATTTTTTAGACCGAAGTGCTCTCCTGATTGAGACTGCTCTCCCTGCTGCT 60
61 ATCTGTTCCTATCAATGACACATTCTCTCATCTGCTCAATGATATG 120
121 TACCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCT 180
181 TCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCTGCT 240
241 AGACAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 300
301 TGAGCCTTCGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 336

>gb|H37323|HUS1GKXDA Mouse monoclonal idiotypic antibody Ig kappa light

1 GATTTTTTAGACCGAAGTGCTCTCCTGATTGAGACTGCTCTCCCTGCTGCT 60
61 ATCTGTTCCTATCAATGACACATTCTCTCATCTGCTCAATGATATG 120
121 TACCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCT 180
181 TCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCTGCT 240
241 AGACAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 300
301 TGAGCCTTCGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 336

>emb|Z22035|NDIGKXAA M.domesticus Igk variable region.

1 GATTTTTTAGACCGAAGTGCTCTCCTGATTGAGACTGCTCTCCCTGCTGCT 60
61 ATCTGTTCCTATCAATGACACATTCTCTCATCTGCTCAATGATATG 120
121 TACCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCT 180
181 TCTGACGACCTAGGACTCCACCTGACTCTGCTGCTGCTGCTGCTGCTGCT 240
241 AGACAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 300
301 TGAGCCTTCGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 336
Figure 17(B)

**gb|M36589|** MUSIGKABS Mouse Ig kappa-chain mRNA V-J region, partial cDNA.

```
>gb|M36589|
1  GATGTTTTGATGACAAAACCCACTCTCTCCCTGGGAGACATGCCCTCCAGGTTGCTGGGTTG
61  ATCTCTTGGAGACTGATTCTAGAGGTAGGTTGCTGTCTTCTGGGAGACTGATTCTAGAG
121  CGAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
181  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
241  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
301  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
336  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
```

**gb|M32858|** MUSIGKCSQ Mouse Ig rearranged kappa-chain mRNA V-region, partial.

```
>gb|M32858|
1  GATGTTTTGATGACAAAACCCACTCTCTCCCTGGGAGACATGCCCTCCAGGTTGCTGGGTTG
61  ATCTCTTGGAGACTGATTCTAGAGGTAGGTTGCTGTCTTCTGGGAGACTGATTCTAGAG
121  CGAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
181  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
241  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
301  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
336  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
```

**emb|X87231|** MMKAPLI M.musculus mRNA for antibody light chain.

```
>emb|X87231|
89  GATGTTTTGATGACAAAACCCACTCTCTCCCTGGGAGACATGCCCTCCAGGTTGCTGGGTTG
149  ATCTCTTGGAGACTGATTCTAGAGGTAGGTTGCTGTCTTCTGGGAGACTGATTCTAGAG
209  CGAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
269  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
329  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
389  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
```

**gb|U29428|** MUS musculus anti-PC rearranged Ig kappa chain V-J region.

```
>gb|U29428|
13  GATGTTTTGATGACAAAACCCACTCTCTCCCTGGGAGACATGCCCTCCAGGTTGCTGGGTTG
73  ATCTCTTGGAGACTGATTCTAGAGGTAGGTTGCTGTCTTCTGGGAGACTGATTCTAGAG
133  CGAGGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
193  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
253  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
313  GCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
```

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U.S. Patent
Apr. 14, 2009
Sheet 22 of 24
US 7,517,967 B2
Figure 18(A)

>gb|U01185|44010185 Mus musculus BALB/c anti-glycophorin A type N

1 CAGGTGCAGCTGCAAGAGTCTCAAGACCTGCCTGTGCTGCCCTCCCATC 60
61 ACATGCACGCTCTGACCCTGTCTTTAAGCACTATGTTATTAACTGTTGCCAGAGCCCT 120
121 CCAGGAAAGGTTGCTGGAACCTCTGCCTGAAGATGATATGCCCTGGGCTGAGACCAAAATTATCAT 180
181 TCAGCTCTCATCTTCCAGTACAGTGTACATGAAGACACCCGTTCACCCTCAAATTTTCTTA 240
241 AAACCTGAAACAGTCTGCAAACAGTCAGACAGAGCCAGACTGTGCTGCAAACAA 301
292 ----------------------------------------------------------- 315
316 GCTAAGGACACTGCCTGCAAGGAACCTCAGTCACGGTCTCTCCTCA 360

>gb|M26985|MUSIGHPR Mus musculus productively rearranged IgH chain allele 1,

1 CAGGGTCACGTGGAAGAGTCAAGAGTCTGACTGCCTGTGCTGCCCTCCCATC 60
61 ACATGCACGCTCTGACCCTGTCTTTAAGCACTATGTTATTAACTGTTGCCAGAGCCCT 120
121 CCAGGAAAGGTTGCTGGAACCTCTGCCTGAAGATGATATGCCCTGGGCTGAGACCAAAATTATCAT 180
181 TCAGCTCTCATCTTCCAGTACAGTGTACATGAAGACACCCGTTCACCCTCAAATTTTCTTA 240
241 AAACCTGAAACAGTCTGCAAACAGTCAGACAGAGCCAGACTGTGCTGCAAACAA 301
292 ----------------------------------------------------------- 315
316 GCTAAGGACACTGCCTGCAAGGAACCTCAGTCACGGTCTCTCCTCA 360

>dbj|017387|PVYIB Potato virus Y immunoglobulin gene for monoclonal antibody

58 CAGGGTCACGTGGAAGAGTCAAGAGTCTGACTGCCTGTGCTGCCCTCCCATC 60
118 ACATGCACGCTCTGACCCTGTCTTTAAGCACTATGTTATTAACTGTTGCCAGAGCCCT 120
178 CCAGGAAAGGTTGCTGGAACCTCTGCCTGAAGATGATATGCCCTGGGCTGAGACCAAAATTATCAT 237
238 TCAGCTCTCATCTTCCAGTACAGTGTACATGAAGACACCCGTTCACCCTCAAATTTTCTTA 240
298 AAACCTGAAACAGTCTGCAAACAGTCAGACAGAGCCAGACTGTGCTGCAAACAA 301
358 TAC 360
361 TGCGGCAAGGACACACTGCTACAGTCTGCCTCA 393

>gb|K36228|MUSIGHMAEI Mouse Ig heavy-chain mRNA V region, partial cds from

1 CAGGGTCACGTGGAAGAGTCAAGAGTCTGACTGCCTGTGCTGCCCTCCCATC 60
61 ACATGCACGCTCTGACCCTGTCTTTAAGCACTATGTTATTAACTGTTGCCAGAGCCCT 120
121 CCAGGAAAGGTTGCTGGAACCTCTGCCTGAAGATGATATGCCCTGGGCTGAGACCAAAATTATCAT 180
181 TCAGCTCTCATCTTCCAGTACAGTGTACATGAAGACACCCGTTCACCCTCAAATTTTCTTA 240
241 AAACCTGAAACAGTCTGCAAACAGTCAGACAGAGCCAGACTGTGCTGCAAACAA 301
305 TAC 304
305 - 305
306 CTACCTATGCTATGGAACACTGCCTGCAAGGAACCTCAGTCACGGTCTCCTCA 354

>gb|L48671|MUSAB Mus musculus (cell line C57BL/F2-22) chromosome 12 anti-DNA

1 CAGGGTCACGTGGAAGAGTCAAGAGTCTGACTGCCTGTGCTGCCCTCCCATC 60
61 ACATGCACGCTCTGACCCTGTCTTTAAGCACTATGTTATTAACTGTTGCCAGAGCCCT 120
121 CCAGGAAAGGTTGCTGGAACCTCTGCCTGAAGATGATATGCCCTGGGCTGAGACCAAAATTATCAT 180
181 TCAGCTCTCATCTTCCAGTACAGTGTACATGAAGACACCCGTTCACCCTCAAATTTTCTTA 240
241 AAACCTGAAACAGTCTGCAAACAGTCAGACAGAGCCAGACTGTGCTGCAAACAA 292
292 ---------------------------------- 304
305 ACAATGCTATGGAACACTGCCTGCAAGGAACCTCAGTCACGGTCTCCTCA 354
MONOCLONAL ANTIBODY 1A7 AND USE FOR THE TREATMENT OF MELANOMA AND SMALL CELL CARCINOMA

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a continuation of U.S. Ser. No. 10/153,401, filed May 21, 2002, which is a continuation of U.S. Ser. No. 09/293,533, filed on Apr. 15, 1999, issued as U.S. Pat. No. 6,509,016, which is a continuation of U.S. Ser. No. 08/752,844, filed on Nov. 21, 1996, issued as U.S. Pat. No. 5,935,821, which is a continuation-in-part of U.S. Ser. No. 08/372,676, filed Jan. 17, 1995, issued as U.S. Pat. No. 5,612,030; and U.S. Ser. No. 08/591,196, filed Jan. 16, 1996; issued as U.S. Pat. No. 5,977,316; both of which are hereby incorporated herein in their entirety.

STATEMENT OF RIGHTS TO INVENTIONS MADE UNDER FEDERALLY SPONSORED RESEARCH

This invention was made in part during work supported by a grant from the United States Public Health Service (CA72018). The government has certain rights in the invention.

BACKGROUND

In spite of numerous advances in medical research, cancer remains the second leading cause of death in the United States. Traditional modes of clinical care, such as surgical resection, radiotherapy and chemotherapy, have a significant failure rate, especially for solid tumors. Failure occurs either because the initial tumor is unresponsive, or because of recurrence due to regrowth at the original site or metastasis. Cancer remains a central focus for medical research and development.

Under the hypothesis that neoplastic cells are normally regulated by immune surveillance, an attractive approach is to re-focus the immune system in affected individuals back towards the tumor. Many types of cancers should be susceptible to the immune system, because they express unusual antigens that reflect the oncogenic transformation of the cell. Antibodies or T cells directed against a target antigen specifically expressed on tumor cells may be able to recruit immune effector functions, and thereby eliminate the tumor or mitigate the pathological consequences.

There are several potential pitfalls in this approach. The first is that the target tumor antigen may be shed from the cell, and thereby block the approach of tumor-specific immune components. The second is that the expression of the target antigen may be heterologous. In this case, a specific immune response would be ineffective in a substantial subclass of affected individuals, or the risk of escape variants would be high. The third difficulty is that tumor-specific antigens are generally-antigenically related to autoantigens, or comprise autoantigens in an atypical mode of expression. This means that tumor-associated antigens are often poorly immunogenic; perhaps due to an active and ongoing immunosuppression against them. Furthermore, cancer patients tend to be immunosuppressed, and only respond to certain T-dependent antigens.

A number of features suggest that gangliosides may be preferable to other types of target antigens for antibody-mediated killing of certain tumor types. Gangliosides like GD2 have simple, well-defined structures, and the level of expression is not affected by antibody binding. In vitro studies have shown that monoclonal antibodies against gangliosides like GD2 and GD3 potentiate lymphocyte response which could potentially be directed towards tumor cells. In addition, certain gangliosides have been implicated in the adhesion of tumor cells to solid substrates, and antibodies inhibit this attachment. An immune response against them, even if not successful in eliminating the tumor cells, could reduce the extent of pathology.

In particular, glycosphingolipid GD2 is expressed at high density by tumors of human neuroectodermal origin; including malignant melanoma, neuroblastoma, glioma, soft tissue sarcoma and small cell carcinoma of the lung. The GD2 antigen is absent in most normal tissues, except for low levels in brain and peripheral nerve.

Melanoma is one of the human diseases for which there is an acute need of new therapeutic modalities. It is a particularly aggressive form of skin cancer, and occurs in increased frequency in individuals with regular unguarded sun exposure. In the early phases, melanoma is characterized by proliferation at the dermal-epidermal junction, which soon invades adjacent tissue and metastasizes widely. Worldwide, 70,000 patients are diagnosed and 25,000 deaths are reported each year. The American Cancer Society projects that by the year 2000, 1 out of every 75 Americans will be diagnosed with melanoma in their lifetime.

Fortunately, melanoma is one of the cancers for which gangliosides hold significant promise as a target antigen Livingstone (1995) Immunol. Rev. 145:147-166. Increased expression of GD2 has been observed in a majority of malignant melanoma cells. Several murine monoclonal anti-GD2 antibodies were reported to suppress the growth of tumors of neuroectodermal origin in athymic (nu/nu) mice or cause remission in patients with metastatic melanoma. A human-mouse chimeric anti-GD2 antibody remissions in patients with metastatic neuroblastoma. The mechanism is thought to involve antibody dependent cellular cytotoxicity (ADCC) or complement-mediated cytotoxicity (CMC). Clinical responses have been obtained by treating with monoclonal antibodies against GM2, GD2 and GD3. Active immunization with a ganglioside vaccine comprising GM2 produced anti-GM2 antibodies in 50/58 patients, who survived longer than antibody negative patients.

Neuroblastoma is a highly malignant tumor occurring during infancy and early childhood. Except for Wilms’s tumor, it is the most common retroperitoneal tumor in children. This tumor metastasizes early, with widespread involvement of lymph nodes, liver, bone, lung, and marrow. While the primary tumor is resolvable by resection, the recurrence rate is high.

Small cell lung cancer is the most malignant and fastest growing form of lung cancer. It accounts for 20-25% of new cases of lung cancer, and 60,000 cases will be diagnosed in the US in 1996. The primary tumor is generally responsive to chemotherapy, but is shortly followed by wide-spread metastasis. The median survival time at diagnosis is ~1 year, with a 5 year survival rate of only 5-10%.

If there was a simple and reliable therapeutic strategy for providing immune reactivity against GD2, then the clinical prospects for these types of cancers might improve.

Unfortunately, there are several reasons why GD2 is less than ideal as a component of an active vaccine. For one thing, GD2 is of limited supply, and is difficult to purify. Of course, because GD2 is a ganglioside, it cannot be generated by simple recombinant techniques. Secondly, gangliosides in general and GD2 in particular, are poorly immunogenic. In order to render them more immunogenic in humans, it has
been necessary to conjugate them to protein carriers like KLH (U.S. Pat. No. 5,308,614; WO 94/16731).

Similarly, the passive administration of anti-GD2 antibodies is less than ideal as an approach to long-term care. The amount of antibody that must be provided passively is substantial. It may lead to the formation of anti-immunoglobulin or anti-idiotype, which in turn may lead to diminished responsiveness with each succeeding dose. And perhaps most importantly, it fails to provide the host with components of the immune response which may be critical in tumor eradication; particularly tumor-directed cytotoxic T cells.

How else, then, could an active immune response against GD2 be obtained? The network hypothesis of Lindemann and Jerne suggests a way of overcoming both the natural immune tolerance against GD2, and the shortage of supply of GD2. It relies on the fact that antibodies comprise variable region epitopes that themselves may be immunogenic, leading to the generation of second-level antibodies called anti-idiotypes. According to the hypothesis, immunization with a given tumor-associated antigen will generate production of antibodies against this tumor-associated antigen (Ab1), which is purified and used in a second round of immunization to generate the anti-idiotype (Ab2). Some of these Ab2 molecules (called Ab2J) fit into the paratopes of Ab1, and can be used as surrogate tumor-associated antigens. Thus, immunization with Ab2J can lead to the generation of anti-anti-idiotype antibodies (Ab3) that recognize the corresponding original tumor-associated antigen identified by Ab1. The idea is that the Ab2 presents a feature that is structurally related to the tumor antigen in a different context that makes it more immunogenic.

Accordingly, efforts have been made elsewhere using anti-idiotypes to elicit a response against various tumor-associated antigens. One group of investigators raised an anti-idiotype related to the melanoma associated ganglioside GM3 (Kanda et al., Yamamoto et al., Hastings et al.) Saleh et al. and Cheung et al. have raised anti-idiotypes against GD2. Other anti-idiotypes have entered early clinical trials: for example, Mittelman et al. are using an anti-idiotype related to a high molecular weight melanoma associated antigen (MAA). Chattopadhyay et al. have developed anti-idiotypes against a melanoma-associated proteoglycan. Chapman et al are treating melanoma patients with an anti-idiotype related to ganglioside GD3. Most of these trials are still at about the phase I level with results still pending.

It is important to emphasize that every potential anti-idiotype for use in tumor therapy must be evaluated on a case-by-case basis. First, only a fraction of antibodies raised against an Ab1 are limited in their reactivity against the paratope of the Ab1 (i.e., non-reactive against features shared with other potential antibodies in the host individual). Second, anti-idiotypes are not necessarily immunogenic. Third, only a fraction of immunogenic anti-idiotypes elicit a response against the original tumor antigen and not against other antigens with less tissue specificity. These properties are related to the structure of the anti-idiotype, including the amino acid sequence of its variable regions. Different anti-idiotypes raised against the same Ab1 will be different in this respect, and must be evaluated separately. Furthermore, the anti-tumor response elicited will depend on the immunological status of the individual being immunized.

SUMMARY OF THE INVENTION

This disclosure outlines a particular monoclonal anti-idiotype antibody, designated 1A7. This antibody has been established as being capable of eliciting an anti-GD2 response. It has all the desirable properties that provide for escaping immune tolerance to GD2, and is appropriate for treating GD2-associated disease.

Also described are a number of other compounds derived from 1A7. These include in particular polynucleotides and polypeptides based on the 1A7 heavy and light chain variable regions, distinguishing 1A7 from other immunoglobulin molecules. A wide range of such products are contemplated. Preferred derivative compounds include a humanized version of intact 1A7, polypeptide fragments from the complementarity determining regions of the 1A7 heavy chain, and a vaccinia virus vector comprising an encoding region for a 1A7 variable region. Especially preferred compounds include a 1A7 antibody fusion molecule comprising a polypeptide region with GM-CSF or IL-2 attached to the heavy chain constant region, a single-chain V_{H}V_{L} or V_{H}V_{L} variable region, and polynucleotides encoding such polypeptides.

The compounds and compositions of this invention may be used inter alia for eliciting a humoral or cellular immune response in an individual. They may be used for detecting or treating a GD2-associated disease, including therapy of advanced disease, and prophylactic care, particularly for decreasing the risk of recurrence in an adjuvant setting.

Accordingly, this invention embodies monoclonal antibody 1A7, and a hybridoma cell line that produces it. Also included are antibodies having identifying characteristics of 1A7.

Another embodiment of this invention is a polynucleotide comprising a sequence encoding a polypeptide with immunological activity of monoclonal antibody 1A7, wherein the polypeptide comprises at least 5 consecutive amino acids from a variable region of monoclonal antibody 1A7. The variable region may be from either the 1A7 light chain or heavy chain. The 5 consecutive amino acids preferably play a role in 1A7 immunoreactivity, and may be from a complementarity determining region. Another embodiment is an isolated polynucleotide of at least 20 consecutive nucleotides capable of forming a stable duplex with the 1A7 light or heavy chain encoding sequence, but not with encoded sequences for other previously described immunoglobulin molecules. Any of these polynucleotides may be in the form of cloning vectors, expression vectors, or transfected into host cells.

Also embodied is a polypeptide having immunological activity of monoclonal antibody 1A7, wherein the polypeptide comprises at least 5 consecutive amino acids from a variable region of monoclonal antibody 1A7. The variable region may be from a light chain or heavy chain. The 5 consecutive amino acids preferably play a role in 1A7 immunological reactivity, and may be from a complementarity determining region. Intact 1A7, functionally active fragments of 1A7, fusion proteins, humanized antibodies, multiple antigen proteins, and other polypeptide derivatives of 1A7 are included. Of special interest are single-chain variable regions and fusion proteins comprising a cytokine.

Another embodiment is a pharmaceutical composition or vaccine, comprising monoclonal antibody 1A7, a polynucleotide of this invention, or a polypeptide of this invention, and a pharmaceutically acceptable excipient.

A further embodiment of this invention is a method of eliciting an immune response in an individual comprising administering to the individual an effective amount of monoclonal antibody 1A7, or a polynucleotide or polypeptide of this invention. The immune response may comprise either humoral or cellular immunity, preferably both.

Yet another embodiment is a method of treating a GD2-associated disease in an individual, comprising administering
monoclonal antibody 1A7, or a polynucleotide or polypeptide of this invention. The disease may be melanoma, neuroblastoma, glioma, soft tissue sarcoma, and small cell carcinoma. The individual may have a clinically detectable tumor, or the tumor may have been previously treated and rendered undetectable. The method may be for palliating the disease, or for reducing the risk of recurrence.

A further embodiment of this invention is a kit for detection or quantitation of an anti-GD2 antibody or a 1A7 polynucleotide in a sample, comprising monoclonal antibody 1A7 or a polynucleotide or polypeptide of this invention in suitable packaging. Also embodied are methods for detecting such antibodies or polynucleotides by employing a reagent or kit embodied in this invention.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 is a depiction of the cDNA sequence (SEQ ID NO:1) and the amino acid sequence (SEQ ID NO:2) of the light chain variable region of 1A7 and adjoining residues.

FIG. 2 is a depiction of the cDNA sequence (SEQ ID NO:3) and the amino acid sequence (SEQ ID NO:4) of the heavy chain variable region of 1A7 and adjoining residues.

FIG. 3 is a listing in which the amino acid sequences of the 1A7 variable region are compared with 15 light and heavy chain immunoglobulin sequences from the prior art. Panel A (amino acids 1 through 112 of SEQ. ID NO:2) shows the light chain comparison; Panel B (SEQ. ID NO:5:14) shows the heavy chain comparison. Panel C shows variable region consensus sequences for the light and heavy chains (SEQ. ID NO:15:16), and compares them with those of 1A7. The variable region of 1A7 shows splicing differences about the VDJ junction of the heavy chain, and an additional 16 point differences.

FIG. 4 is a graph depicting the binding of monkey Ab3 to 1A7 (1A7) by sandwich radioimmunoassay. Open squares denote Ab3 from monkey PR0#685; open circles denote Ab3 from monkey PR0#778.

FIG. 5 is a bar graph depicting inhibition of Ab1-A2 binding by monkey Ab3 or vice-versa. For each pair of bars, the left hand bar denotes Ab2-Ab3 with labeled Ab1; the right hand bar denotes Ab1-Ab3 with labeled Ab2.

FIG. 6 is a bar graph depicting inhibition of binding of 125I labeled 14G2a antibody to GD2 positive melanoma cell line M21/P6 in the presence of different concentrations of Ab1 and monkey Ab3. Parallel inhibition curves were obtained using either purified Ab1 or Ab3 from monkey sera. For each triad of bars, the left hand bar denotes control binding with 14G2a; the middle (hatched) bar denotes binding of Ab3 from monkey PRO#685; the right hand bar denotes binding of Ab3 from monkey PRO#778.

FIG. 7 is a graph from a FACS analysis of the binding of monkey Ab3 to tumor cells. Panel A shows the staining observed of GD2-expressing M21/P6 cells labeled with pre-immune and immune Ab3. Panel B shows the staining observed on another cell line not expressing GD2. Panel C shows control staining of M21/P6 cells using the GD2-specific antibody 14G2a, or no antibody.

FIG. 8 is a bar graph depicting binding of Ab1 and monkey Ab3 to different gangliosides by ELISA. For each pair of bars, the left hand (solid) bar denotes the binding of Ab3 from monkey PRO#685; the right hand (hatched) bar denotes control binding by anti-GD2 antibody 14G2a. This experiment shows the antibody induced upon immunization with the anti-idiotypic 1A7 is antigen specific.

FIG. 9 is a bar graph depicting inhibition of binding of 125I labeled 14G2a antibody to purified GD2 by 14G2a and monkey Ab3. For each triad of bars, the left hand (solid) bar denotes monkey PRO#778; the middle (open) bar denotes 14G2a, the right hand (hatched) bar denotes monkey PRO#685.

FIG. 10 is a half-tone reproduction of a blot analysis showing binding of 14G2a and monkey Ab3 to different gangliosides. Lane 1, monkey PRO#685; lane 2, monkey PRO#778; lane 3, 14G2a; lane 4, monkey anti-11D10; lane 5, PBS-BSA.

FIG. 11 is a bar graph depicting the proliferation of T cells obtained from monkeys immunized with monoclonal antibody 1A7, when they are stimulated in culture using either antibody 1A7 or a cancer cell line expressing GD2.

FIG. 12 is two bar graphs depicting the character of Ab3 antibody purified from the sera of human patients treated with antibody 1A7. Upper panel shows that the Ab3 response comprises specific antibody to ganglioside GD2 (hatched bars) but not GD3 (solid bars). Lower panel shows that the anti-GD2 response is predominantly IgG (hatched bars) rather than IgM (solid bars).

FIG. 13 is a two graphs further characterizing Ab3 from three human patients. The induced Ab3 inhibits the binding of anti-GD2 to purified ganglioside GD2 (upper panel) or a GD2 expressing cancer cell line (lower panel) in a dose-dependent fashion.

FIG. 14 is a map of a model plasmid construct for creating a vaccinia vector comprising a region encoding a 1A7 heavy or light chain variable region.

FIG. 15 is a listing of the nucleotide sequence (SEQ. ID NO:65) and amino acid translation (SEQ. ID NO:66) for a single-chain variable region molecule (scFv), developed from the variable region sequences of intact monoclonal antibody 1A7.

FIG. 16 is a map of a vector plasmid comprising an expression cassette for a scFv and adapted for administration as a human polynucleotide vaccine.

FIG. 17 depicts the 10 polynucleotide sequences that were most closely matched to the 1A7 light chain variable region encoding sequence in a database search. These sequences have the designations SEQ. ID NO:17-26.

FIG. 18 depicts the 10 polynucleotide sequences that were most closely matched to the 1A7 heavy chain variable region encoding sequence in a database search. These sequences have the designations SEQ. ID NO:27-44.

DETAILED DESCRIPTION

This invention relates to the discovery of an anti-idiotypic antibody that is capable of recruiting a tumor-specific response against GD2. The antibody is designated 1A7. The immune response elicited by 1A7 typically comprises both humoral and cellular components, and is therefore expected to be useful in palliating the clinical conditions related to GD2-associated tumors. The invention comprises the 1A7 antibody molecule, along with polynucleotide and polypeptide derivatives thereof, and methods for using these compounds in diagnosis and treatment.

Cancer patients are typically tolerant to various tumor associated antigens (TAA), including GD2. 1A7 successfully circumvents immune tolerance, and elicits an immune response against GD2. According to the network theory Ab1 represents anti-tumor monoclonal antibody; Ab2 represents anti-idiotypic monoclonal antibody; and Ab3 represents anti-idiotypic monoclonal antibody. 1A7 is an Ab2 that stimulates Ab3 with the same specificity as Ab1, and can therefore recruit effector functions against tumors bearing the tumor antigen. An Ab3 response reactive against the original TAA is referred to herein as Ab3'.
While not wishing to be bound by theory, one explanation is that the 1A7 combining site may present a region that at least partly resembles an epitope in GD2 in the context of one or more other epitopes which render it more immunogenic. The epitope of GD2 which may resemble that of 1A7 is identified by the Ab1 (14G2α) used to generate 1A7. As a result, 1A7 escapes the normal immune tolerance against GD2, and is able to elicit an anti-GD2 response.

FIGS. 1 and 2 show the nucleotide encoding region of the 1A7 light and heavy chain variable regions, respectively, along with the corresponding amino acid translation. These sequences were compared with those of other known immunoglobulin molecules (Example 2). Both the polynucleotide and polypeptide variable region sequences for 1A7 are unique.

Amongst the 50 database sequences matched most closely to that of the 1A7 light chain variable region, none was identical 1A7 differed from the five closest sequences by 2 substitution differences at residues 50 and 55, which are contained in the second CDR. The two differences at these positions were non-conservative substitutions, and persisted in comparisons with other light chain sequences.

Amongst the 50 database sequences matched most closely to that of the 1A7 heavy chain variable region, none was identical FIG. 3, Panels A and B). The following summarizes the main points deduced from the comparison.

The closest match was with a heavy chain fragment. There were 2 deletions and 12 substitution differences.

The closest match with a full length heavy chain variable region had the following features: There were 3 deletions and 17 substitution differences.

1A7 differed in length from all sequences but one, due to insertions or deletions of 1 to 8 residues about the VDJ junction. For the sequence of equal length, there were 20 substitution differences.

All other comparisons showed a total of at least 22 insertions, deletions and substitution differences. Differences appeared throughout the variable region.

FIG. 3 Panel C provides a comparison of the 1A7 amino acid light and heavy chain sequences with consensus sequences derived from the database sequences. Other than splicing differences about the heavy chain VDJ junction, there appear to be about 16 differences between 1A7 and the consensus sequences that have likely arisen from somatic mutation during antibody maturation.

Particularly of interest in developing 1A7 derivatives with 1A7 immunoactivity are regions of the polynucleotide or polypeptide sequence comprising part of the VDJ junction. Also of interest are regions spanning at least one, preferably 2, and more preferably 3 or more of the point differences between the 1A7 amino acid sequences and the consensus sequences.

The full sequences of the 1A7 light and heavy chain constant regions have not been determined, but are expected to be identical or nearly identical to those of other mouse immunoglobulin molecules.

For the mouse kappa light chain constant region, four genetic allotopes encoding two protein allotopes have been published by Solin et al. (1993) Immunogenetics 37:401-407, which is hereby incorporated herein by reference. FIG. 1 of Solin et al. depicts mouse and rat immunoglobulin kappa chain gene sequences, comparing the sequences within the kappa chain constant region for different strains and highlighting allotopic differences. Included are kappa chain constant region sequences for BALB/c, Pt., SJL, and M. spretrus. Other naturally occurring allotopes are possible.

The mouse γ1 heavy chain constant region DNA sequence from newborn mice has been published by Honjo et al. (1979) Cell 18:559-568, which is hereby incorporated herein by reference. FIG. 5 of Honjo et al. shows the germ-line DNA sequence, along with the encoded protein sequence. Shown in the line above is another protein sequence obtained from the mouse myeloma MOPC 21. Other naturally occurring allotopes are possible.

The 1A7 antibody and derivatives thereof are useful, for example, for eliciting an anti-GD2 immune response, for treating a GD2-associated disease, and as reagents for detecting the presence of anti GD2.

Certain compounds, compositions and methods described in this application relate generally to 1A7 and derivatives thereof which are routinely generated by classical techniques of immunochemistry. This includes 1A7 which has been coupled to another compound by chemical conjugation, or by mixing with an excipient or an adjuvant. It includes such immunoglobulin fragments as Fab, F(ab')2, Fab', and isolated heavy and light chains. First generation therapies are those based on such compounds and compositions.

Other compounds, compositions and method described in this application relate to polynucleotide and polypeptide derivatives of 1A7 that fall outside the preceding classification. The compositions are typically generated by genetic engineering, although they may alternatively be obtained by other methods and combinations of methods. This classification includes (but is not limited to) isolated polynucleotide fragments, recombinant polynucleotides, engineered peptide fragments and fusion peptides, and polynucleotides or polypeptides prepared by chemical synthesis based on the sequence data. Preferred compounds include polypeptide fragments of the 1A7 CDRs, antibody fusion proteins comprising cytokine effector components, single-chain variable region proteins and the polynucleotides encoding them, therapeutic plasmids comprising 1A7 polynucleotides, and vectors such as vaccinia vectors.

Unless explicitly stated otherwise or required by their rate, methods of making or using certain compounds embodied in this invention may be applied to other compounds of this invention, as appropriate.

Pharmaceutical compositions and treatment modalities of this invention may be brought to bear whenever it is desirable to elicit a response against GD2, especially in humans. Human patients with GD2-associated tumors, including melanoma, neuroblastoma, glioma, soft tissue sarcoma, and small cell carcinoma (including small cell lung cancer) are especially appropriate subjects. The subjects may have an advanced form of disease, in which case the objective may include mitigation or reversal of disease progression, and amelioration of side effects. The subjects may have had a history of the condition, for which they have already been treated, in which case the objective will typically include a decrease or delay in the risk of recurrence of clinical pathology.

Additionally, the antibodies and derivatives of this invention may also be used as probes, primers, and polypeptides for use as diagnostic reagents. These applications are described in more detail in the sections that follow.

Definitions

"1A7" is a particular anti-idiotypic antibody raised against the anti-GD2 monoclonal antibody with the designation 14G2α. The generation and characterization of 1A7 is described in Example 1.

"Immunological activity" of 1A7 refers to any of the following activities: ability to specifically bind monoclonal anti-
body 14G2a; ability to inhibit the binding of 1A7 to 14G2a or 14G2a to GD2 in a specific manner, and an ability to elicit an immune response against GD2. A specific immune response may comprise antibody, B cells, T cells, and any combination thereof; and effector functions resulting therefrom. Included are the antibody-mediated functions ADCC and complement-mediated cytosis (CMC). The T cell response includes T helper cell function, cytotoxic T cell function, inflammation/inducer T cell function, and T cell mediated suppression. A compound able to elicit a specific immune response according to any of these criteria is referred to as "immunogenic".

1A7 "activity" or "function" refers to any of the immunological activities of 1A7, or to any other biological activity ascribed to 1A7 in this disclosure, including the role of 1A7 in the amelioration or palliation of GD2-associated disease.

The "variable region" of 1A7 refers to the variable region of the 1A7 light chain or the variable region of the 1A7 heavy chain, either alone or in combination, and includes amino acids encoded by the V, D, and J encoding regions of the respective gene, as appropriate.

GM-CSF, IL-2, and other biologically active molecules referred to herein are meant to include fragments and derivatives based on the respective parent molecule that have the same biologic or physiologic function.

A "polynucleotide" is a polymeric form of nucleotides of any length, which contain deoxyribonucleotides, ribonucleotides, and analogs in any combination analogs. Polynucleotides may have any three-dimensional structure, and may perform any function, known or unknown. The term "polynucleotide" includes double-, single-stranded, and triple-helical molecules. Unless otherwise specified or required, any embodiment of the invention described herein that is a polynucleotide encompasses both the double-stranded form and each of two complementary single-stranded forms known or predicted to make up the double stranded form.

The following are non-limiting examples of polynucleotides: a gene or fragment, exons, introns, mRNA, tRNA, rRNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes, and primers. A polynucleotide may comprise modified nucleotides, such as methylated nucleotides and nucleotide analogs, unmodified, other sugars and linking groups such as fluororibose and thioate, and nucleotide branches. The sequence of nucleotides may be interrupted by non-nucleotide components. A polynucleotide may be further modified after polymerization, such as by conjugation with a labeling component. Other types of modifications included in this definition are caps, substitution of one or more of the naturally occurring nucleotides with an analog, and introduction of means for attaching to proteins, metal ions, labeling components, other polynucleotides, or a solid support.

The term "recombinant" polynucleotide as used herein intends a polynucleotide of genomic, cDNA, semisynthetic, or synthetic origin which either does not occur in nature or is linked to another polynucleotide in an arrangement not found in nature.

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to polymers of amino acids of any length. The polymer may be linear or branched, it may comprise modified amino acids or amino acid analogs, and it may be interrupted by non-amino acids. The terms also encompass an amino acid polymer that has been modified naturally or by intervention; for example, disulfide bond formation, glycosylation, lipoprotein, acetylation, phosphorylation, or any other manipulation or modification, such as conjugation with a labeling component. Unless stated or implied otherwise, the term 1A7 polypeptide includes any polypeptide monomer or polymer with 1A7 activity, including the intact 1A7 antibody, and smaller and larger functionally equivalent polypeptides, as described herein.

A "fusion polypeptide" is a polypeptide comprising regions in a different position in the sequence than occurs in nature. The regions may normally exist in separate proteins and are brought together in the fusion polypeptide; or they may normally exist in the same protein but are pieced in a new arrangement in the fusion polypeptide.

A "functionally equivalent fragment" of a 1A7 polypeptide or polynucleotide varies from the native sequence by any combination of additions deletions, or substitutions while preserving at least one functional property of the fragment relevant to the context in which it is being used. A functionally equivalent fragment of a 1A7 polynucleotide either encodes a polypeptide that is functionally equivalent to 1A7 when used in an expression system, or has similar hybridization specificity as a 1A7 polynucleotide when used in a hybridization assay. A functionally equivalent fragment of a 1A7 polypeptide typically has one or more of the following properties: ability to bind monoclonal antibody 14G2a and 1A7 to inhibit the binding of 1A7 to 14G2a or 14G2a to GD2 in a specific manner, and an ability to elicit an immune response with a similar antigen specificity as that elicted by 1A7.

A "cell line" or "cell culture" denotes higher eukaryotic cells grown or maintained in vitro. It is understood that the descendants of a cell may not be completely identical (either morphologically, genotypically, or phenotypically) to the parent cell.

A "vector" refers to a recombinant DNA or RNA plasmid or virus that comprises a heterologous polynucleotide to be delivered into a target cell, either in vitro or in vivo. The heterologous polynucleotide may comprise a sequence of interest for purposes of therapy, and may optionally be in the form of an expression cassette. As used herein, a vector need not be capable of replication in the ultimate target cell or subject. The term includes cloning vectors for the replication of a polynucleotide, and expression vectors for translation of a polynucleotide encoding sequence. Also included are viral vectors, which comprise a polynucleotide encapsidated or enveloped in a viral particle.

A "host cell" denotes a eukaryotic cell that has been genetically altered, or is capable of being genetically altered by administration of an exogenous polynucleotide, such as a recombinant plasmid or vector. When referring to genetically altered cells, the term refers both to the originally altered cell and to the progeny thereof.

"Heterologous" means derived from a genotypically distinct entity from the rest of the entity to which it is being compared. For example, a polynucleotide may be placed by genetic engineering techniques into a plasmid or vector derived from a different source, and is a heterologous polynucleotide. A promoter removed from its native coding sequence and operatively linked to a coding sequence with which it is not naturally found linked is a heterologous promoter.

A "signal sequence" or "leader sequence" is a short amino acid sequence that directs a newly synthesized secretory or membrane protein through a cellular membrane, usually the endoplasmic reticulum. Signal sequences are typically in the N-terminal portion of a polypeptide and are typically removed between biosynthesis and secretion of the polypeptide from the cell.

An "isolated" polynucleotide or polypeptide is one that is substantially free of the materials with which it is associated.
in nature. By substantially free is meant at least 50%, preferably at least 70%, more preferably at least 80%, and even more preferably at least 90% free of the materials with which it is associated in nature.

A "stable duplex" of polynucleotides, or a "stable complex" formed between any two or more components in a biochemical reaction, refers to a duplex or complex that is sufficiently long-lasting to persist between the formation of the duplex or complex and subsequent detection, including any optional washing steps or other manipulation that may take place in the interim.

A "biological sample" encompasses a variety of sample types, including blood and other liquid samples of biological origin, solid tissue samples such as a biopsy specimen or tissue cultures, or cells derived therefrom and the progeny thereof. The definition also includes samples that have been manipulated in any way after their procurement, such as by treatment with reagents, solubilization, or enrichment for certain components, such as proteins or polynucleotides. The term encompasses various kinds of clinical samples obtained from any species, and also includes cells in culture, cell supernatants, and cell lysates.

A "vaccine" is a pharmaceutical composition for human or animal use, which is administered with the intention of conferring the recipient with a degree of specific immunological reactivity against a particular target, or group of targets. The immunological reactivity may be antibodies or cells (particularly B cells, plasma cells, T helper cells, and cytotoxic T lymphocytes, and their precursors) that are immunologically reactive against the target, or any combination thereof. For purposes of this invention, the target is primarily tumor associated antigen GD2, but also includes any tumor associated antigen bound by 14G2a. The immunological reactivity may be desired for experimental purposes, for treatment of a particular condition, for the elimination of a particular substance, or for prophylaxis. An active vaccine is a vaccine intended to elicit an immune response in the recipient that persists in the absence of the vaccine components.

"Adjuvant" as used herein has two meanings, depending on the context in which it is used. In the context of a pharmaceutical preparation, an adjuvant is a chemical or biological agent given in combination with an antibody, polynucleotide or polypeptide to enhance its immunogenicity. In the context of cancer diagnosis or management, it refers to a class of subjects with no clinically detectable tumor mass, but who are suspected of being at risk of recurrence because of a history of cancer.

When referring to a type of cancer that normally manifests as a solid tumor, a "clinically detectable" tumor is one that is detectable on the basis of tumor mass; i.e., by such procedures as CAT scan, X-ray, or palpation. Positive biochemical, histological or immunological findings on their own are insufficient to meet this definition.

As used herein, "treatment" refers to clinical intervention in an attempt to alter the natural course of the individual or cell being treated, and may be performed either for prophylaxis or during the course of clinical pathology. Desirable effects include preventing occurrence or recurrence of disease, alleviation of symptoms, diminishment of any direct or indirect pathological consequences of the disease, preventing metastasis, lowering the rate of disease progression, amelioration or palliation of the disease state, and remission or improved prognosis.

The "pathology" associated with a disease condition is anything that compromises the well-being, normal physiology, or quality of life of the affected individual. This may involve (but is not limited to) destructive invasion of affected tissues into previously unaffected areas, growth at the expense of normal tissue function, irregular or suppressed biological activity, aggravation or suppression of an inflammatory or immunological response, increased susceptibility to other pathogenic organisms or agents, and undesirable clinical symptoms such as pain, fever, nausea, fatigue, mood alterations, and such other features as may be determined by an attending physician.

An "effective amount" is an amount sufficient to effect a beneficial or desired clinical result. An effective amount can be administered in one or more doses. For purposes of this invention, an effective amount of a 1A7 polynucleotide or polypeptide is an amount that induces an immune response, particularly an anti-GD2 response. In terms of treatment, an effective amount is amount that is sufficient to palliate, ameliorate, stabilize, reverse or slow the progression of the GD2-associated disease, or otherwise reduce the pathological consequences of the disease.

An "individual" is a vertebrate, preferably a mammal, more preferably a human. Mammals include, but are not limited to, farm animals, sport animals, and pets.

General Techniques
The practice of the present invention will employ, unless otherwise indicated, conventional techniques of molecular biology (including recombinant techniques), microbiology, cell biology, biochemistry and immunology, which are within the skill of the art. Such techniques are explained fully in the literature, such as, "Molecular Cloning: A Laboratory Manual", second edition (Sambrook et al., 1989); "Oligonucleotide Synthesis" (M. J. Gait, ed., 1984); "Animal Cell Culture" (R. J. Freshney, ed., 1987); "Methods in Enzymology" (Academic Press, Inc.); "Handbook of Experimental Immunology" (D. M. Weir & C. C. Blackwell, eds.); "Gene Transfer Vectors for Mammalian Cells" (J. M. Miller & M. P. Coligan, eds., 1987); "Current Protocols in Molecular Biology" (F. M. Ausubel et al., eds., 1987); "PCR: The Polymerase Chain Reaction", (Mullis et al., 1994); "Current Protocols in Immunology" (J. E. Coligan et al., eds., 1991).

All patents, patent applications, articles and publications mentioned herein, both supra and infra, are hereby incorporated herein by reference.

These techniques are applicable to the production of the polynucleotides and polypeptides of the invention, and, as such, may be considered in making and practicing the invention. Particularly useful techniques for particular embodiments will be discussed in the sections that follow.

How Monoclonal Antibody 1A7 Was Generated and Selected
1A7 was obtained by immunizing naive mice with 14G2a anti-GD2 antibody to obtain an anti-idiotypic response. 14G2a binds to a unique epitope of GD2. Syngeneic BALB/c mice were immunized four times with 14G2a (Ab1) and their spleen cells were fused with the non-secretory mouse myeloma P3-635 cells.

To obtain an anti-idiotypic with all the features desired, an extensive screening process was employed, comprising the following four important steps: (1) Positive selection for antibody binding to 14G2a; (2) Negative selection against antibody recognizing isotypic or allotypic determinants; (3) Positive selection for an ability to inhibit the binding of 14G2a to GD2; and (4) Positive selection for an ability to induce a humoral immune response against the original tumor-associated antigen (GD2) in both mice and rabbits.

Several anti-idiotypic (Ab2) hybridomas were obtained that were specific for the idioic component of the 14G2a immunogen, and did not react with its isotypic or allotypic determinants. To determine whether the anti-14G2a were
directed against the parotope of 14G2a, the binding of radio-
labeled 14G2a to the GI2-positive cell line M21/P6 was
studied in the presence of varying amounts of Ab2 hybridoma
culture supernatants. With as little as 5 pL of culture superna-
tant, three of the Ab2 tested inhibited the binding by at least
70%. 1A7 showed the highest level of inhibition. Accord-
ingly, 1A7 was grown and purified from ascites fluid for
further studies, while the others were kept in reserve.

The purified Ab2 was prepared as a vaccine and injected
into naive mice and rabbits. After 4 injections, serum samples
were titered for the presence of Ab3 that bound not only to the
immunizing Ab2, but also to GI2. The Ab2 passing all of
these screening stages was designated 1A7. Further details of
the method used to obtain 1A7 are provided in Example 1.

Ab3 produced in animals immunized with 1A7 has been
further characterized. The immune sera from both mice and
rabbits, prepared with 14G2a for binding to the GI2-associ-
ated cell line M21/P6 and inhibited the binding of radioiodi-
nated 14G2a to 1A7. This indicated that anti-anti-Id (Ab3) in
mice and rabbits may share idiotopes with 14G2a and they
probably bind to the same epitope as Ab1. Administration of
1A7 to non-human primates (cynomolgus monkeys) also
generated a specific immune response, comprising activity
against GI2 (Example 3).

The nucleic acid sequence encoding the light and heavy
chain variable regions of 1A7 have also been deduced, along
with the translated protein sequences (Example 2).

The invention also encompasses 1A7 conjugated to a label
capable of producing a detectable signal. These conjugated
antibodies are useful, for example, in detection systems such
as quantitation of anti-GI2 or tumor imaging. Such labels are
known in the art and include, but are not limited to,
radiotopes, enzymes, fluorescent compounds, chemiluminescent
compounds, and bioluminescent compounds. The labels may
be covalently linked to 1A7, or conjugated to the 1A7 through
a secondary reagent, such as a second antibody, protein A, or
a biotin-avidin complex.

Preparation of 1A7 Antibody

The 1A7 antibody of this invention can be prepared in
several ways.

It is most conveniently obtained from the hybridoma
deposited with the ATCC under Accession No. HB-11786,
or the progeny thereof. For example, the cells can be cultured in
a suitable medium, and spent medium can be used as an
antibody source. Optionally, matrix-coated channels or beads
and cell co-cultures may be included to enhance growth of
antibody-producing cells. For the production of large
amounts of antibody, it is generally more convenient to obtain
an ascites fluid. The method of raising ascites generally com-
prises injecting hybridoma cells into an immunologically
naive histocompatible or immunotolerant mammal, espe-
cially a mouse. The mammal is optionally primed for ascites
production by prior administration of a suitable composition;
for example, Pristane.

Alternatively, 1A7 can be chemically synthesized using
sequence data and other information provided in this disclo-
sure, in conjunction with standard methods of protein synthe-
sis. A suitable method is the solid-phase Merrifield technique.
Automated peptide synthesizers are commercially available,
such as those manufactured by Applied Biosystems, Inc.
(Foster City, Calif.).

1A7 may also be obtained by employing routine recombi-
nant methods such as described in Sambrook et al. (1989). For
instance, using the sequences and information provided
herein, a polynucleotide encoding either the 1A7 heavy or
light chain can be cloned into a suitable expression vector
(which contains control sequences for transcription, such as a
promoter). The expression vector is in turn introduced into a
host cell. The host cell is grown under suitable conditions
such that the polynucleotide is transcribed and translated into
a protein. Heavy and light chains of 1A7 may be produced
separately, and then combined by disulfide bond rearrange-
ment. Alternatively, vectors with separate polynucleotides
encoding each chain of 1A7, or a vector with a single poly-
nucleotide encoding both chains as separate transcripts, may
be transfected into a single host cell which may then produce
and assemble the entire molecule. Preferably, the host cell is
a higher eukaryotic cell that can provide the normal carbohy-
drate complement of the molecule. The 1A7 thus produced in
the host cell can be purified using standard techniques in the
art. A polynucleotide encoding 1A7 for use in the production
of 1A7 by any of these methods can in turn be obtained from
the hybridoma producing 1A7, or be produced synthetically
or recombinantly from the DNA sequence provided herein.

Methods of antibody isolation are well known in the art
See, for example, Harlow and Lane (1988) Antibodies: A
Laboratory Manual, Cold Spring Harbor Laboratory, New
York. The 1A7 antibody is a mouse immunoglobulin of the
IgG1 subclass, and may be isolated by any technique suitable
for immunoglobulins of this isotype. Purification methods
may include salt precipitation (for example, with ammonium
sulfate), ion exchange chromatography (for example, on a
cationic or anionic exchange column run at neutral pH and
eluted with step gradients of increasing ionic strength), gel
filtration chromatography (including gel filtration HPLC),
and chromatography on affinity resins such as protein A,
protein G, hydroxypatite, and anti-immunoglobulin. 1A7
may also be purified on affinity columns comprising the
14G2a paratope; for example, in the form of a purified Ab1 or
Ab3. Preferably, 1A7 is purified from BALB/c ascites using
Protein-A-CL-SEPHAROSE™ 4B chromatography followed
by chromatography on a DEAE-SEPHAROSE™ 4B
ion exchange column.

If 1A7 is to be administered to an individual, it is preferably
at least 90% pure, more preferably it is at least 90% pure; even
more preferably it is at least 95% pure and free of pyrogens
and other contaminants. In this context, the percent purity is
calculated as a weight percent of the total protein content of
the preparation, and does not include constituents which are
deliberately added to the composition after the 1A7 is puri-
fied.

Uses of 1A7 Antibody

The 1A7 antibody may be used for a number of purposes.
These include eliciting an antibody response to 1A7 or GI2,
eliciting a T cell response to 1A7 or GI2, and treating various
types of cancer. These uses are elaborated more fully in a later
section.

1A7 may also be used to purify anti-1A7 (Ab3), anti-GI2
(Ab1'), or 14G2a (Ab1). The method comprises contacting a
biological sample containing the antibody with a 1A7
polypeptide, producing a complex, and recovering the Ab3 or
Ab1 from the complex. Typically, the 1A7 polypeptide is
coupled to an affinity matrix for affinity column purification.
Such methods are routine in the art and need not be described
in detail herein.

The invention also encompasses methods of detecting anti-
1A7 or anti-GI2 in a biological sample. Anti-GI2 is detect-
able whenever (like 14G2a) it cross-reacts with 1A7. Anti-
GI2 with this activity may spontaneously arise during the
course of a GI2-associated disease. Anti-GI2 with this
activity is especially likely in individuals who have received
a course of therapy with 1A7, or a derivative thereof. These
methods are applicable in a clinical setting, for example, for monitoring antibody levels in an individual, as well as an industrial setting, in which commercial production of anti-

A7 or anti-GD2 is desired.

The assay methods entail coning any anti-A7 or anti-D2 target antibody in the sample with a A7 antibody or polypeptide under conditions suitable to allow the formation of a stable complex between the target and the A7, and detecting any stable complex formed. The sample is suitably prepared before conducting the assay, optionally by enriching for antibody activity. When using intact A7, it is generally preferable to deplete the sample of any anti-mouse immunoglobulin activity that may be present. Anti-mouse immunoglobulin antibody can be removed from a sample, for example, by precipitation with normal mouse IgG or adsorption with a mouse Ig adsorbent. Binding of anti-mouse immunoglobulin antibody, particularly that specific for the Fc region, can be minimized by judicious choice of the reagents of the assay. F(ab)'2 or Fab fragments of A7 and other reagents with fewer mouse determinants are appropriate.

After the sample is suitably prepared, it is mixed with a excess functional equivalent of A7 under conditions that permit formation of a complex between A7 and any target antibody that may be present. The amount of complex is then determined, and compared with complexes formed with standard samples containing known amounts of target antibody in the range expected. Complex formation may be observed by immunoprecipitation or nephelometry, but it is generally more sensitive to employ a reagent labeled with such labels as radioisotopes like 125I, enzymes like peroxidase and β-galactosidase, or fluorochromes like fluorescein.

Antibody assays may be conducted entirely in fluid phase. For example, anti-GD2 may be mixed with labeled A7. Alternatively, the anti-GD2 in the sample may be used to compete with a labeled anti-GD2 for binding sites on A7. Generally, bound and unbound label is separated to quantitate the percent bound. Suitable separation methods include gel filtration chromatography, and precipitation with antibody against immunoglobulin of the species from which the sample is obtained, optionally in the presence of polyethylene glycol. Alternatively, the proportion of bound and unbound label may be determined in situ, for example, using quenching/labeling pairs or enzyme/inhibitor labeling pairs. See, e.g., U.S. Pat. No. 3,996,345 (Ullman et al.).

It is generally more convenient to conduct a capture assay using a reagent linked to a solid phase, such as a polyethylene tube, microtiter plate well, or magnetic bead. In a competition-type capture assay, unlabeled target antibody in the sample competes with a labeled analog reagent for binding to A7. The A7 may be attached directly to the solid support, or captured later, for example, using an anti-immunoglobulin. In this assay, the amount of label associated with the solid phase is inversely related to the amount of anti-GD2 in the sample.

In the sandwich-type capture assay, target antibody is captured by A7 attached directly or through a secondary reagent to a solid phase. After washing, the anti-GD2 is detected using anti-immunoglobulin of the appropriate species, or a second A7 antibody, to which a label is directly or indirectly attached. Alternatively, the anti-immunoglobulin may be attached to the solid phase and labeled A7 is used to complete the sandwich. If the anti-immunoglobulin used is isotype-specific, then the class of the antibody may also be determined. In this type of assay, the amount of label associated with the solid phase correlates positively with the amount of anti-GD2 in the sample. Other methods of measuring specific antibody are known in the art, and may be adapted to measure anti-A7 or anti-GD2 by using A7 as the specific reagent

A7 may also be used to measure the level of cellular anti-A7 or anti-GD2 activity. In one example, A7 is used to identify anti-GD2 expressing cells in a cell suspension, perhaps B or T lymphocytes expressing a receptor that binds A7. A7 may be labeled and mixed into the cell suspension. Alternatively, unlabeled A7 may be mixed with the cells, and followed with a labeled secondary reagent such as labeled anti-mouse immunoglobulin or protein A. Suitable labels for this purpose include radioisotopes and fluorescent labels. The use of fluorescent labels also allows anti-GD2 cells to be separated from non-specific cells in a fluorescence-activated cell sorter. In a second example, anti-GD2 expressing cells are detected in a tissue section. Typically, the tissue is fixed and embedded in a suitable medium, overlaid with A7, and then developed with a secondary anti-immunoglobulin coupled with a fluorescent or enzyme marker.

Polynucleotide Derivatives of A7

The invention provides various polynucleotides encoding the anti-idiotypic antibody A7 or fragments of A7, based on the polynucleotide sequences provided herein. Various embodiments are described in this section, comprising a number of different combinations of the A7 heavy or light chain variable region sequences. In general, a A7 polynucleotide of this invention encodes at least one feature that is unique to the A7 molecule (in comparison with other immunoglobulins). Preferably, this feature is related in some way to an immunological reactivity of A7.

The invention encompasses a polynucleotide encoding a portion of the A7 heavy chain variable region, comprising at least about 70 consecutive nucleotides, preferably at least about 80 consecutive nucleotides, more preferably at least about 100 consecutive nucleotides, even more preferably at least about 150 nucleotides of SEQ ID NO:1. The invention also encompasses a polynucleotide encoding a portion of the A7 light chain variable region, comprising at least about 25 consecutive nucleotides, preferably at least about 30 consecutive nucleotides, even more preferably at least about 35 consecutive nucleotides of the CDR1 encoding sequence thereof. The invention also encompasses a polynucleotide encoding a portion of the A7 light chain variable region, comprising at least about 20 consecutive nucleotides, preferably at least about 25 consecutive nucleotides, even more preferably at least about 35 consecutive nucleotides of the CDR2 or CDR3 encoding sequence thereof.

The invention also encompasses a polynucleotide encoding a portion of the A7 heavy chain variable region, comprising at least about 70 consecutive nucleotides, preferably at least about 80 consecutive nucleotides, more preferably at least about 100 consecutive nucleotides, even more preferably at least about 150 nucleotides of SEQ ID NO:3. The invention also encompasses a polynucleotide encoding a portion of the A7 light chain variable region, comprising 15 consecutive nucleotides of the CDR1 encoding sequence thereof. The invention also encompasses a polynucleotide encoding a portion of the A7 light chain variable region, comprising at least about 20 consecutive nucleotides, preferably at least about 25 consecutive nucleotides, even more preferably at least about 35 consecutive nucleotides of the CDR2 or CDR3 encoding sequence thereof.

The invention includes isolated A7 polynucleotides encoding a polypeptide having immunological activity of A7, wherein the polypeptide encodes at least 5 amino acids of a variable light chain of A7 as depicted in SEQ ID NO:2.
In another embodiment, the invention includes isolated 1A7 polynucleotides encoding a polypeptide having immunological activity of 1A7, wherein the polynucleotide encodes at least 5 amino acids of a variable heavy chain of 1A7 as depicted in SEQ ID NO:4. The polynucleotide sequence may be similar to those depicted in SEQ ID NO:1 (Fig. 1) or SEQ ID NO:3 (Fig. 2) with changes designed to optimize codon usage, stability, or to facilitate cloning, or for some other purpose. It is within the skill of one in the art, given the amino acid sequence in SEQ ID NO:2 or SEQ ID NO:4, to design such polynucleotides. Preferred are polynucleotides encoding at least five amino acids of a 1A7 complementarity determining region (CDR).

The invention also encompasses polynucleotides encoding for functionally equivalent variants and derivatives of 1A7 and functionally equivalent fragments thereof which may enhance, decrease or not significantly affect properties of the polypeptides encoded thereby. These functionally equivalent variants, derivatives, and fragments display the ability to induce an immune response, preferably an anti-GD2 immune response. For instance, changes in a DNA sequence that do not change the encoded amino acid sequence, as well as those that result in conservative substitutions of amino acid residues, one or a few amino acid deletions or additions, and substitution of amino acid residues by amino acid analogs are those which will not significantly affect properties of the encoded polypeptide.

The 1A7 polynucleotides of the invention may comprise additional sequences, such as additional encoding sequences within the same transcription unit, controlling elements such as promoters, ribosome binding sites, and polyadenylation sites, additional transcription units under control of the same or a different promoter, sequences that permit cloning, expression, and transformation of a host cell, and any such construct as may be desirable to provide embodiments of this invention.

The invention includes a polynucleotide of at least about 15 consecutive nucleotides, preferably at least about 20 nucleotides, more preferably at least about 25 consecutive nucleotides, more preferably at least about 35 consecutive nucleotides, more preferably at least about 50 consecutive nucleotides, even more preferably at least about 75 nucleotides, still more preferably at least about 100 nucleotides, still more preferably at least about 200 nucleotides, and even more preferably at least about 300 nucleotides that forms a stable hybrid with a polynucleotide encoding the light chain or heavy chain variable region of 1A7, but not with other immunoglobulin encoding regions known at the time of filing of this application. Any set of conditions may be used for this test, as long as at least one set of conditions exist wherein the test polynucleotide demonstrates the required specificity.

Preferably, the 1A7 encoding sequences to which the test polynucleotide binds are those shown in SEQ ID NOS:1 and 3. Since the known immunoglobulin sequences fall into a hierarchy of similarity with that of 1A7, the test may be performed by comparing the hybridization of the test polynucleotide with the 1A7 sequence with the hybridization with the most closely related sequences. Preferably is a panel of about 10 of the most closely related sequences to SEQ ID NO:1 or 3, as appropriate. Sequences to which the test polynucleotide should not form a duplex are the light chain variable region encoding sequences listed in SEQ ID NOS: 17-26 and the heavy chain variable region encoding sequences listed in SEQ ID NOS: 27-44.

Hybridization reactions can be performed under conditions of different "stringency". Conditions that increase stringency of a hybridization reaction are published. See, for example, Sambrook and Maniatis. Examples of relevant conditions include (in order of increasing stringency): incubation temperatures of 25° C., 37° C., 50° C., and 68° C.; buffer concentrations of 10xSSC, 6xSSC, 1xSSC, 0.1xSSC (where SSC is 0.15 M NaCl and 15 mM citrate buffer) and their equivalent using other buffer systems; formamide concentrations of 0%, 25%, 50%, and 75%; incubation times from 5 minutes to 24 hours; 1, 2, or more washing steps; wash incubation times of 1, 2, or 15 minutes; and wash solutions of 6xSSC, 1xSSC, 0.1xSSC, or deionized water.

"Tₘ" is the temperature in degrees Centigrade at which 50% of a polynucleotide duplex made of complementary strands hydrogen bonded in antiparallel direction by Watson-Crick base pairing dissociates into single strands under conditions of the experiment. Tₘ may be predicted according to a standard formula, such as:

\[ Tₘ = 81.5 + 16.6 \log [Na⁺] + 0.41 \left( \% \text{GC} \right) - 0.1 \left( \% \text{AT} \right) \times 1000 \]

where [Na⁺] is the cation concentration (usually sodium ion) in mol/L; % (GC) is the number of G and C residues as a percentage of total residues in the duplex; (% F) is the percent formamide in solution (wt/vol); and L is the number of nucleotides in each strand of the duplex.

Useful 1A7 polynucleotides encoding fragments of 1A7 may be identified by generating polynucleotide fragments (based on SEQ ID NO:1 or SEQ ID NO:3, for example) and testing the polypeptides encoded thereby for a function of interest. Alternatively, the polypeptide fragment encoded by a particular polynucleotide may be prepared and tested for a function of interest. Alternatively, given a 1A7 polypeptide with desirable properties, a polynucleotide could be designed that encodes it.

Included in all these embodiments are polynucleotides encoding with regions for 1A7 polymers, fusion proteins, humanized immunoglobulins, single-chain variable regions, and other particular polypeptides of interest. These polypeptides are described in a later section.

The invention also provides polynucleotides covalently linked with a detectable label. Such polynucleotides are useful, for example, as probes for detection of related nucleotide sequences.

Preparation of 1A7 Polynucleotides:

The polynucleotides of this invention can be obtained using chemical synthesis, recombinant cloning methods, PCR, or any combination thereof.

Methods of chemical polynucleotide synthesis are well known in the art and need not be described in detail herein. One of skill in the art can use the sequence data provided herein to obtain a desired polynucleotide by employing a DNA synthesizer or ordering from a commercial service.

Alternatively, 1A7 polynucleotide sequences can be obtained from a 1A7 antibody producing cell line, 1A7 cloning vector, or 1A7 expression vector. RNA or DNA encoding the desired sequence may be isolated, amplified, and processed by standard recombinant techniques. Such techniques include digestion with restriction nuclease, and amplification by polymerase chain reaction (PCR), or a suitable combination thereof. PCR technology is described in U.S. Pat. Nos. 4,683,195, 4,800,159, 4,754,065 and 4,683,292, as well as PCR. The Polymerase Chain Reaction. Mullis et al. eds, Birkauesser Press, Boston (1994). A polynucleotide comprising a desired sequence can be inserted into a suitable vector, and the vector in turn can be introduced into a suitable host cell for replication and amplification. Polynucleotides may be inserted into host cells by any means known in the art. Cells
are transformed by introducing an exogenous polynucleotide by direct uptake, endocytosis, transfection, mating or electroporation. Once introduced, the exogenous polynucleotide can be maintained within the cell as a non-integrated vector (such as a plasmid) or integrated into the host cell genome. Amplified DNA can be isolated from the host cell by standard methods or, e.g., Sambrook et al. (1989) RNA may also be obtained from transformed host cell, it may be obtained by using an RNA-dependent DNA polymerase.

If used as a vaccine, plasmids containing 1A7 polynucleotides are preferably prepared by the method of Horn et al. (1995) Human Gene Therapy 6:565-573, which produces a pharmaceutical grade plasmid DNA suitable for administration.

Cloning and Expression vectors Comprising a 1A7 Polynucleotide

The present invention further includes a variety of vectors comprising a 1A7 polynucleotide. These vectors can be used for expression of recombinant polypeptides as well as a source of 1A7 polynucleotides. Cloning vectors can be used to obtain replicate copies of the 1A7 polynucleotides they contain, or as a means of storing the polynucleotides in a depository for future recovery. Expression vectors (and host cells containing these expression vectors) can be used to obtain polypeptides produced from the polynucleotides they contain. They may also be used where it is desirable to express 1A7 polypeptides in an individual and thus have intact cells capable of synthesizing the polypeptide, sum as in gene therapy. Suitable cloning and expression vectors include any known in the art, e.g., those for use in bacterial, mammalian, yeast and insect expression systems. Specific vectors and suitable host cells are known in the art and need not be described in detail herein. For example, see Gaeusa and Ramji, Vectors, John Wiley & Sons (1994).

Cloning and expression vectors typically contain a selectable marker (for example, a gene encoding a protein necessary for the survival or growth of a host cell transformed with the vector), although such a marker gene can be carried on another polynucleotide sequence co-introduced into the host cell. Only those host cells into which a selectable gene has been introduced will grow under selective conditions. Typical selection genes either: (a) confer resistance to antibiotics or other toxic substances, e.g., ampicillin, neomycin, methotrexate; (b) complement auxotrophic deficiencies; or (c) supply critical nutrients not available from complex media. The choice of the proper marker gene will depend on the host cell, and appropriate genes for different hosts are known in the art. Cloning and expression vectors also typically contain a replication system recognized by the host.

Suitable cloning vectors may be constructed according to standard techniques, or may be selected from a large number of cloning vectors available in the art. While the cloning vector selected may vary according to the host cell intended to be used, useful cloning vectors will generally have the ability to self-replicate, may possess a single target for a particular restriction endonuclease, or may carry genes for a marker that can be used in selecting clones containing the vector. Suitable examples include plasmids and bacterial viruses, e.g., pUC18, mp18, mp19, pBR322, PMB9, ColEl, pCR1, RP4, phage DNAs, and shuttle vectors such as pSA3 and pAT28. These and many other cloning vectors are available from commercial vendors such as BioRad, Stratagene, and Invitrogen.

Expression vectors generally are replicable polynucleotide constructs that contain a polynucleotide encoding a 1A7 polypeptide of interest. The polynucleotide encoding the 1A7 polypeptide is operatively linked to suitable transcriptional controlling elements, such as promoters, enhancers and terminators. For expression (i.e., translation), one or more translational controlling elements are also usually red such as ribosome binding sites, translation initiation sites, and stop codons. These controlling elements (transcriptional and translational) may be derived from the 1A7 gene, or they may be heterologous (i.e., derived from other genes or other organisms). A polynucleotide sequence encoding a signal peptide can also be included to allow a 1A7 polypeptide to cross or lodge in cell membranes or be secreted from the cell. A number of expression vectors suitable for expression in eukaryotic cells including yeast, avian, and mammalian cells are known in the art. For example of an expression vector is pEJ5A3 (Invitrogen, San Diego, Calif., in which transcription is driven by the cytomegalovirus (CMV) early promoter/enhancer. This vector also contains recognition sites for multiple restriction enzymes for insertion of the 1A7 polynucleotide of interest. Another example of an expression vector (system) is the baculovirus/insect system.

The vectors containing the polynucleotide of interest can be introduced into the host cell by any of a number of appropriate means, including electroporation, transfection employing calcium chloride, rubidium chloride, calcium phosphate, DEAE-dextran, or other substances; microprojectile bombardment; lipofection; and infection (where the vector is an infectious agent, such as vaccinia virus, which is discussed below). The choice of means of introducing vectors or 1A7 polynucleotides will often depend features of the on the host cell.

Once introduced into a suitable host cell, for example, E. coli or COS-7, expression of a 1A7 polypeptide can be determined using any of the assays described herein. For example, presence of 1A7 polypeptide can be detected by RIA or ELISA of the culture supernatant (if the 1A7 polypeptide is secreted) or cell lysates.

A particularly useful expression vector for 1A7 polynucleotides is a vaccinia virus comprised of a 1A7 polynucleotide sequence, which can also be used in vaccine preparations (Moss (1991) Science 252:1662-1667). To introduce polynucleotide sequences encoding 1A7 polypeptide, including 1A7 polypeptide fragments, into vaccinia, the polynucleotide sequence of interest is first inserted into a plasmid containing a vaccinia virus promoter with flanking sequences homologous to vaccinia DNA essential for replication. Plasmid-containing cells are then infected with vaccinia, which leads to a low level of homologous recombination between plasmid and virus, with resultant transfer of the vaccinia promoter and 1A7 polypeptide-encoding polynucleotide sequence into the vaccinia virus genome. Typically, the 1A7 polynucleotide is inserted into the viral tk (thymidine kinase) gene. Insertion into the tk site attenuates the virus more than 10,000 fold compared to wild type (Flexner et al. (1980) Vaccine 88 (Cold Spring Harbor Laboratory), 179-184). Recombinant virus is identified by the tk- phenotype. Preferably, expression of the 1A7 polynucleotide is under the control of the vaccinia early/late promoter (7.5 K), whereby the resultant 1A7 polypeptides can be expressed in infected cells throughout the life cycle of the virus. However, other promoters known in the art can be used, such as phosphohexose, or synthetic promoters. Expression of the 1A7 polypeptide occurs in cells infected with the recombinant vaccinia or individuals which are immunized with the live recombinant vaccinia virus. Any one of several strains of vaccinia can be used, including but not limited to WR, ALVAC, and NYVAC.

A vaccinia vector of this invention can contain one or more polynucleotides encoding a 1A7 polypeptide. It can also con-
tain polynucleotide sequences encoding other polypeptides that enhance, facilitate, or modulate the desired result, such as lymphokines, including, but not limited to, IL-2, IL-4 and GM-CSF. A preferred lymphokine is GM-CSF. Preferred GM-CSF constructs are those which have been deleted for the AU-rich elements from the 3′ untranslated regions and sequences in the 5′ untranslated region that are capable of forming a hairpin loop. Also embodied in this invention are vaccinia vectors encoding for recombinant I A7 variants containing I A7 polypeptides, such as scFvs, chimeras, and polymers, as described below.

Host Cells Transformed with I A7 Polynucleotides

Other embodiments of this invention are host cells transformed with I A7 polynucleotides and vectors comprising I A7 polynucleotide sequences, as described above. Both prokaryotic and eukaryotic host cells may be used. Prokaryotic hosts include bacterial cells, for example E. coli and mycobacteria. Among eukaryotic hosts are yeast, insect, avian, plant and mammalian cells. Host systems are known in the art and need not be described in detail herein. One example of a mammalian host cell is NS0, obtainable from the European Collection of Cell Cultures (England). Transfection of NS0 cells with a plasmid, for example, which is driven by a CMV promoter, followed by amplification of this plasmid in using glutamine synthetase provides a useful system for protein production. (Cockett et al. (1990) Bio/Technology 8:662-667).

The cells of this invention can be used, inter alia, as repositories of I A7 polynucleotides, or as vehicles for production of I A7 polypeptides and polynucleotides. They may also be used as vehicles for in vivo expression of I A7 polypeptides.

Uses for I A7 Polynucleotides

The polynucleotides of this invention have several uses. I A7 polynucleotides are useful, for example, in expression systems for the production of I A7 or I A7 fragments. They are also useful as hybridization probes to assay for the presence of I A7 polynucleotide or related sequences in a sample using methods well known to those in the art. Further, I A7 polynucleotides are also useful as primers to effect amplification of desired polynucleotides. The polynucleotides of this invention are also useful in pharmaceutical compositions including vaccines and for gene therapy.

I A7 polynucleotides can also be used as hybridization probes for detection of I A7 encoding sequences. Suitable samples include cells transformed ex vivo for use in gene therapy. In one illustration, DNA or RNA is extracted from a sample, and optionally run on a gel and/or digested with restriction nucleases. The processed sample polynucleotide is typically transferred to a medium suitable for washing. The sample polynucleotide is then contacted with the I A7 polynucleotide probe under conditions that permit a stable duplex to form if the sample contains a matching I A7 sequence. Any stable duplexes formed are detected by any suitable means. For example, the I A7 polynucleotide probe may be supplied in labeled form, and label remaining with the sample after washing will directly reflect the amount of stable duplex formed. In a second illustration, hybridization is performed in situ. A suitably prepared tissue sample is overlaid with a labeled I A7 probe to indicate the location of I A7 encoding sequences comprised therein.

A short I A7 polynucleotide may also be used as a primer for a PCR reaction, particularly to amplify a longer sequence comprising a region hybridizing with the primer. This may be conducted preparatively, in order to produce polynucleotide for further genetic manipulation. It may also be conducted analytically, to determine whether a I A7 encoding polynucleotide is present, for example, in a sample of diagnostic interest.

The I A7 polynucleotides of this invention can be used in expression systems to produce I A7 polypeptides, intact I A7, or recombinant forms of I A7, such as are described below. Another use of I A7 polynucleotides is in vaccines and gene therapy. The general principle is to administer the polynucleotide so that it either promotes or attenuates the expression of the polypeptide encoded therein. Thus, the present invention includes methods of inducing an immune response and methods of treatment comprising administration of an effective amount I A7 polynucleotides to an individual. In these methods, a I A7 polynucleotide encoding a I A7 polypeptide is administered to an individual, either directly or via cells transfected with the I A7 polynucleotide. Preferably, the I A7 polynucleotide is in the form of a circular plasmid, preferably in a supercoiled configuration. Preferably, the I A7 polynucleotide is replicated inside a cell. Thus, the I A7 polynucleotide is operatively linked to a suitable promoter, such as a heterologous promoter that is intrinsically active in cells of the target tissue type. Preferably, once in cell nuclei, plasmids persist as circular non-replicating episomal molecules. In vitro mutation may be carried out with plasmid constructs to provide, for example, molecules that are more immunogenic or that comprise a T cell epitope with a desirable HLA motif.

To determine whether plasmids containing I A7 polynucleotides are capable of expression in eukaryotic cells, eukaryotic cells such as COS-7, CHO, or HeLa cells can be transfected with the plasmids. Expression of I A7 polypeptide is then determined by immunoassay; for example, by Western blot. Smaller I A7 polypeptides can be detected, for example, by constructing the plasmid so that the resultant I A7 polypeptide is fused with a tag, such as a target epitope or enzyme label. Further characterization of the expressed I A7 polypeptide can be achieved by purifying the peptide and then conducting one of the functional assays described herein.

In one mode of gene therapy, the polynucleotides of this invention are used for genetically altering cells ex vivo. In this strategy, cells removed from a donor or obtained from a cell line are transfected or transduced with vectors encoding a 7 polypeptide, and then administered to a recipient. Suitable cells for transfection include peripheral blood mononuclear cells.

In another mode of gene therapy, the polynucleotides of this invention are used for genetically altering cells in vivo. The purpose may include (but is not limited to) eliciting an antibody response to I A7 or GD2, eliciting a T cell response to I A7 or GD2, and treating various types of cancer. These uses are elaborated more fully in a later section.

Polypeptide Derivatives of I A7

The present invention encompasses polypeptide fragments of I A7 containing at least a portion of a variable region of I A7. Preferred fragments are those with immunological activity of I A7.

Also preferred are fragments which comprise amino acid sequences substantially different from other immunoglobulins, and fragments comprising a CDR. In one embodiment, the invention includes a polypeptide fragment of the I A7 light chain variable region, comprising at least 5 consecutive amino acids, more preferably 15 consecutive amino acids, still more preferably 30 consecutive amino acids of SEQ ID NO:2, wherein the fragment contains one or the other (preferably both) of the lysine residues at positions 50 and 55 of the mature light chain variable region sequence. In other embodiments, the invention includes a polypeptide fragment of the
IA7 heavy chain variable region, comprising the 5 amino acids from the CDR1 or comprising at least 8, preferably 10, more preferably 12, and still more preferably 16 of the amino acids from the CDR2; or comprising at least 4, preferably 6, more preferably 8, and still more preferably 9 of the amino acids from the CDR3. In yet other embodiments, the peptide comprises a combination of two or more sequences from positions 50-55 of the IA7 light chain and the three CDRs of the heavy chain as outlined in this paragraph, optionally in combination with other IA7 variable region sequences.

The size of the IA7 polypeptide fragments may be only the minimum size required to provide a desired function. It may optionally comprise additional sequence, either native to IA7, or from a heterologous source, as desired. IA7 fragments may contain only 5 consecutive amino acids from a IA7 variable region sequence. Polypeptides comprising 7 amino acids, more preferably about 10 amino acids, more preferably about 15 amino acids, more preferably about 25 amino acids, more preferably about 50 amino acids, more preferably about 75 amino acids from the IA7 light or heavy chain variable region are also included. Even more preferred are polypeptides comprising the entire IA7 light or heavy chain variable region.

The invention includes modified IA7 polypeptides which are functionally equivalent to IA7, or have altered but measurable IA7 immunologic activity. Fragments with improved IA7 immunologic activity are preferred. Examples of modified polypeptides include polypeptides with conservative substitutions of amino acid residues, and one or more deletions or additions of amino acids which do not significantly deleteriously change the functional activity.

One example of this is IA7 polypeptides comprising one or more amino acid substitution in comparison with the prototype IA7 sequence. Substitutions can range from changing or modifying one or more amino acids to complete redesign of a region, such as the variable region. Amino acid substitutions, if present, are preferably conservative substitutions that do not deleteriously affect folding or functional properties of the peptide. Groups of functionally related amino acids within which conservative substitutions may be made are glycine / alanine; valine/isoleucine/leucine; asparagine/glutamine; aspartic acid/glutamic acid; serine/threonine/methionine; lysine/arginine; and phenylalanine/tyrosine/tryptophan. Polypeptides of this invention may be in glycosylated or unglycosylated form, may be modified post-translationally (e.g., acetylation, and phosphorylation) or may be modified synthetically (e.g., the attachment of a labeling group).

The invention also encompasses fusion proteins comprising one or more IA7 polypeptide. A IA7 fusion polypeptide can be prepared, for example, by chemical synthesis, or by creating and translating a polynucleotide in which the peptide regions are encoded in the desired relationship. Alternatively, fusion proteins may be provided in expression systems constructed by co-transfection with plasmids comprising encoding regions for different functional regions of the protein.

Useful heterologous sequences for inclusion in a fusion polypeptide include sequences that provide for secretion from a host cell, enhance immunological reactivity, or facilitate the coupling of the polypeptide to an immunoassay support or a vaccine carrier. One example is a bacteria "super antigens", such as staphylococcal enterotoxin A (SEA) (Dohlenst et al. (1994) Proc. Natl. Acad. Sci. USA 91:8945-8949). In a preferred example, a IA7 polypeptide is fused with a bioreponse modifier, particularly a cytokine, which may enhance immunogenicity. Examples of bioreponse modifiers include cytokines and lymphokines such as GM-CSF, interleukin-2 (IL-2), interleukin 4 (IL-4), and y-interferon. GM-CSF and IL-2 are especially preferred. The preferred arrangement is for the cytokine effector unit to be fused to the C-terminal of the immunoglobulin heavy chain.

IA7 polypeptide derivatives comprising both a IA7 light chain and a IA7 heavy chain may be formed as separate light and heavy chains and then assembled, or assembled in situ by an expression system for both chains. Such expression systems may be created by tranfecting with a plasmid comprising separate transcribable regions for the light and heavy chain, or by co-transfecting the same cell with plasmids for each chain. In a third method, a suitable plasmid with a heavy chain encoding region is transfected into a heavy chain loss mutant. For example, heavy chain loss mutants can be obtained by treating 2x10^9 IA7 cells with fluorescein-labeled rabbit anti-mouse IgG (H1 chain specific, DAKO Corporation, Carpinteria, Calif.) according to the supplier's instruction. The stained and unstained cell populations are analyzed in a fluorescence-activated cell sorter. The unstained cells are collected in a sterilized tube and placed in 96-well plates with 1 cell/well by limiting dilution. The culture supernatants are then assayed by ELISA using goat anti-mouse IgG (heavy chain specific) and goat anti-mouse kappa. The clones with kappa-positive and IgG-negative phenotype are subcloned at least 3 times to obtain stable IA7 k/H-10 mutants. mRNA from putative heavy chain loss mutant IA7 k/H-10 cones can be isolated and the sequence of the light chain variable region cDNA determined. Reverse PCR of the mRNA for the IA7 V_{\mu} is performed with 2 sets of 5' and 3' primers, used for cloning of IA7 k/H-10 cDNA (Example 2). A heavy chain loss mutant should yield no detectable DNA band. Transfection of these cells may then proceed using a suitable heavy chain plasmid construct.

The invention also encompasses a hybrid antibody, in which one pair of heavy and light chains is obtained from a first antibody, which the other pair of heavy and light chains is obtained from a second antibody. For purposes of this invention, one pair of light and heavy chains is from IA7. In one example, each light-heavy chain pair binds different epitopes of GD2. Such hybrids may also be formed using chimeric heavy or light chains.

Another embodiment is a humanized version of IA7. "Humanized" antibodies are antibodies in which at least part of the sequence has been altered from its initial form to render it more like human immunoglobulins. In one version, the heavy chain and light chain constant regions are replaced with human sequence. This is a fusion polypeptide comprising a IA7 variable region and a heterologous immunoglobulin constant region. In another version, the CDR regions comprise IA7 amino acid sequences, while the variable framework regions have also been converted human sequences. See, for example, EP 0329400. In a third version, variable regions are humanized by designing consensus sequences of human and mouse variable regions, and converting residues outside the CDRs that are different between the consensus sequences.

Another IA7 derivative contemplated by this invention is an antibody in which the IA7 heavy or light chain has been modified to provide additional properties. For instance, a change in amino acid sequence can result in greater immunogenicity of the resultant IA7 polypeptide. The changes range from changing of one or more amino acids to the complete redesign of a region such as a constant region domain. Changes contemplated affect complement fixation, interaction with membrane receptors, and other effector functions. A recombinant IA7 antibody may also be designed to aid the specific delivery of a substance (such as a lymphokine) to an effector cell. Also contemplated are proteins in which
various immunoglobulin domains have been placed in an order other than occurs in nature.

The invention also encompasses single chain variable region fragments ("scFv") of 1A7. Single chain variable region fragments are made by linking light and/or heavy chain variable regions by using a short linking peptide (Bird et al. 1988 Science 242: 423-426). Any peptide having sufficient flexibility and length can be used as a linker in a scFv. Usually the linker is selected to have little to no immunogenicity. An example of a linking peptide is (GGGGSG)(SEQ. ID NO:45), which bridges approximately 3.5 nm between the carboxy terminus of one variable region and the amino terminus of another variable region. Other linker sequences may also be used, and may provide additional functions, such as a means for attaching a drug or to a solid support.

All or any portion of the heavy or light chain can be used in any combination. Typically, the entire variable regions are included in the scFv. For instance, the light chain variable region can be linked to the heavy chain variable region. Alternatively, a portion of the light chain variable region can be linked to the heavy chain variable region, or portion thereof. Also contemplated are scFvs in which the heavy chain variable region is from 1A7, and the light chain variable region is from another immunoglobulin. It is also possible to construct a biphase, scFv in which one component is a 1A7 polypeptide and another component is a different polypeptide, such as a T cell epitope.

The scFv can be assembled in any order, for example, V_{H}(linker)-V_{L} or V_{L}(linker)-V_{H}. There may be a difference in the level of expression of these two configurations in particular expression systems, in which case one of these forms may be preferred. Tandem scFvs can also be made, such as (X)(linker)-(X)-(linker)-(X), in which X are 1A7 polypeptides, or combinations of 1A7 polypeptides with other polypeptides. In another embodiment, single chain 1A7 antibody polypeptides have no linker polypeptide, or just a small, inflexible linker. Possible configurations are V_{L}/V_{H} and V_{H}/V_{L}. The linkage is too short to permit interaction between V_{L} and V_{H} within the chain, and the chains form homodimers with a V_{L}/V_{H} antigen binding site at each end. Such molecules are referred to in the art as "diabodies".

Single chain variable regions may be produced either recombinantly or synthetically. For synthetic production of scFv, an automated synthesizer can be used. For recombinant production of scFv, a suitable plasmid containing polynucleotide that encodes the scFv can be introduced into a suitable host cell, such as yeast, plant, insect or mammalian cells, or prokaryotic, such as E. coli, and the expressed protein may be isolated using standard protein purification techniques.

A particularly useful system for the production of 1A7 scFv's is plasmid vector pET-22b(+)(Novagen, Madison, Wis.) in E. coli. pET-22b(+) contains a nickel ion binding domain consisting of 6 sequential histidine residues, which allows the expressed protein to be purified on a suitable affinity resin. Another example of a vector that can be used is pETDNA3 (Invitrogen, San Diego, Calif.), described above.

Conditions of expression should be such that the scFv polypeptide can assume optimal tertiary structure. Depending on the plasmid used (especially the activity of the promoter) and the host cell, it may be necessary to modulate the rate of production. For instance, use of a weaker promoter, or expression at lower temperatures, may be necessary to optimize production of properly folded scFv in prokaryotic systems; or it may be preferable to express scFv in eukaryotic cells.

Preferred single chain variable regions comprise at least 10 consecutive amino acids of SEQ. ID NO:2 and at least 10 consecutive amino acids of SEQ. ID NO:4, especially wherein the amino acids of SEQ. ID NO:2 and the amino acids of SEQ. ID NO:4 are joined by a linker polypeptide of 5 to 20 amino acids, or comprising the light chain variable region and the heavy chain variable region of monoclonal antibody 1A7.

The invention also encompasses polymeric forms of 1A7 polypeptides, containing a plurality of 1A7 polypeptides. One embodiment is a linear polymer of 1A7 polypeptides, optionally conjugated to carrier. These linear polymers can comprise multiple copies of a single 1A7 polypeptide, or combinations of different 1A7 polypeptides, and can have tandem 1A7 polypeptides, or 1A7 polypeptides separated by other amino acid sequences. Another embodiment is 1A7 multiple antigen peptides (MAPs). MAPs have a small immunologically inert core having radially branching lysine dendrites, onto which a number of 1A7 polypeptides are covalently attached. See Posnett et al. (1988). Biol. Chem. 263:1719-1725; Tam (1989) Methods Enzymol. 168:7-15. The result is a large macromolecule having a high molar ratio of 1A7 polypeptides to core. MAPs are efficient immunogens and useful antigens for immunosassays. The core for creating an 1A7 MAP can be made by standard peptide synthesis techniques, or obtained commercially (Quality Controlled Biochemicals, Inc., Hopkinton, Mass.). A typical core matrix is made up of three levels of lysine and eight amino acids.

In another embodiment, the immunogenicity of the 1A7 polypeptides is enhanced by preparing them in expression systems in which they are assembled with particle-forming proteins such as, for example, those associated with hepatitis B virus. See, e.g., U.S. Pat. No. 4,722,840. Constructs wherein the 1A7 polypeptide is linked directly to particle-forming protein coding sequences produce hybrids which are immunogenic for an anti-GD2 response. The vectors also comprise immunogenic HBV epitopes; for example, the pre-S peptide and stimulate a response against HBV. Such expression systems may be provided in eukaryotic cells, including yeast or mammalian cells.

In another embodiment, 1A7 polypeptides are conjugated to a carrier molecule. This is desirable for a 1A7 peptide that comprises a suitable epitope for eliciting anti-GD2, but is too small to be immunogenic. Any conjugation method known in the art may be used. Any carrier can be used which is not harmful to the host. Suitable carriers are typically large, slowly metabolized macromolecules such as proteins; polysaccharides (such as latex functionalized SEPHAROSE™, agarose, cellulose, cellulose beads and the like); polymeric amino acids (such as polyglutamic acid, polysine, and the like); amino acid copolymers; and inactive virus particles or attenuated bacteria, such as Salmonella. Especially useful carrier proteins are serum albumins, keyhole limpet hemocyanin (KLH), certain immunoglobulin molecules, thyroglobulin, ovalbumin, and tetanus toxoid. KLH is especially preferred.

1A7 polypeptides of the invention can be identified in a number of ways. For example, the variable regions of the light and heavy-chains can be screened by preparing a series of short polypeptides that together span the entire variable region amino acid sequence. Using a series of polypeptides of 20 or 50 amino acids in length, each 1A7 variable region may be surveyed for useful functional properties. It is also possible to carry out a computer analysis of a protein sequence to identify potentially interesting polypeptides, such as those that bear the shape of D2, or those involved in idiotype-anti-idiotype contact.
Those skilled in the art will readily appreciate that the various adaptations of 1A7 described in this section may be combined in various fashions to yield other 1A7 polypeptides with desirable properties. For instance, 1A7 polypeptides with modified residues may be comprised in a MAP. In another example, a 1A7 scfv is fused to a cytokine, such as IL-2. AU such combinations are contemplated in this invention.

Preparation of 1A7 Polypeptides

The polypeptides of this invention can be made by any suitable procedure, including protein synthesis of the 1A7 antibody, by recombinant methods or by chemical synthesis.

1A7 polypeptides, especially shorter polypeptides up to about 50 amino acids, are conveniently made by chemical synthesis, based on the sequence data and other information provided herein.

Certain 1A7 polypeptides which are fragments of the whole molecule may alternatively be prepared from enzymatic cleavage of intact 1A7. Examples of proteolytic enzymes include, but are not limited to, trypsin, chymotrypsin, pepsin, papain, V8 protease, subtilisin, plasmin, and thrombin. Intact 1A7 can be incubated with one or more proteinases simultaneously or sequentially. Alternatively, or in addition, 1A7 can be treated with disulfide reducing agents. Peptides may then be separated from each other by techniques known in the art, including but not limited to gel filtration chromatography, gel electrophoresis, and reverse-phase HPLC.

A 1A7 polypeptide can also be made by obtaining a polynucleotide encoding it according to the information provided elsewhere in this application, and introducing it into a suitable expression system. Typically, polynucleotides encoding a 1A7 polypeptide are ligated into an expression vector under control of a suitable promoter and used to genetically alter the intended host cell. Both eukaryotic and prokaryotic host systems can be used. The polypeptide is then isolated from lysed cells or from the culture medium and purified to the extent needed for its intended use. Examples of prokaryotic host cells appropriate for use with this invention include E. coli. Examples of eukaryotic host cells include avian, insect, plant, and animal cells such as COS7, HeLa, and CHO cells.

In certain applications, such as when a 1A7 polypeptide is expressed in a suitable storage medium such as a plant seed, the 1A7 polypeptide can be used without purification (Fiedler et al. (1995) Biotechnology 13:1090-1093). For most applications, it is generally preferable that the polypeptide is at least partially purified from other cellular constituents. Preferably, the polypeptide is at least about 50% pure as a weight percent of total protein. More preferably, the protein is at least about 50-75% pure. For clinical use, the polypeptide is preferably at least about 50% pure, according to the criteria given in another section.

Characterization of 1A7 Polypeptides

The 1A7 polypeptides of this invention can be characterized in several ways. For instance, a 1A7 polypeptide may be tested for its ability to bind specifically to 14G2a, for its ability to specifically inhibit the binding between 14G2a and intact 1A7, or for its ability to specifically inhibit the binding between 14G2a and GD2. Alternatively, a 1A7 polypeptide can be tested for its ability to elicit an immune response, preferably an anti-GD2 response. 1A7 polypeptides can also be tested for their ability to palliate or ameliorate GD2-associated diseases, such as GD2-associated tumors. It is understood that only one of these properties need be present in order for a polypeptide to come within this invention, although preferably more than one of these properties is present.

The ability of a 1A7 polypeptide to bind 14G2a may be tested by immunocassay. Any form of direct binding assay is suitable. In one such assay, the 14G2a or alternatively the 1A7 polypeptide is labeled. Suitable labels include radioisotopes such as 125I, enzymes such as peroxidase, fluorescent labels such as fluorescein, and chemiluminescent labels. Typically, the other binding partner is insolubilized (for example, by coating onto a microtiter plate) to facilitate washing. After combining the labeled component with the insolubilized component, the solid phase is washed and the amount of bound label is determined. Another such assay is a sandwich assay, in which the putative 1A7 polypeptide is captured by a first anti-immunoglobulin on a solid phase and developed with a second anti-immunoglobulin. Either the insolubilized or labeled anti-immunoglobulin or both is 14G2a. In either of these examples, the extent of binding of 1A7 is directly related to the amount of label bound to the solid phase.

To conduct the inhibition assays, the putative 1A7 polypeptide is titrated for its ability to decrease the binding of 1A7 to 14G2a, or 14G2a to GD2. Either of the binding pairs in the reaction to be inhibited is labeled, while the other is typically insolubilized in order to facilitate washing. GD2, if it is used, may be provided as the purified ganglioside, or as a GD2-expressing cell line, like M21/P6. The 1A7 polypeptide is typically mixed with the labeled component, and then the name is combined with the solid phase. Polypeptides with the characteristics of 1A7 will proportionately decrease the amount of label attached to the solid phase, compared to control polypeptides. This test may be more sensitive than measuring direct binding, because lower affinity interaction between 1A7 and AbI may be too weak to form a stable bond, but adequate to interfere with the binding of another ligand-receptor pair when present at sufficient concentration.

A 1A7 polypeptide or a molecule comprising such a peptide that has an Fe-like effector component may be tested for their ability to mediate immune effector reactions, particularly complement-mediated cytology (CMC) and antibody-dependent cellular cytotoxicity (ADCC). Suitable assays are described elsewhere in this disclosure.

Another way of characterizing 1A7 polypeptides, particularly those intended for use in therapy, is to test their ability to generate an immune response. Suitable techniques for characterizing the immune response are given in a later section.

Uses of Polypeptides

Polypeptide fragments, fusion proteins, and other derivatives of 1A7 have many uses. The uses generally parallel those of the intact 1A7 antibody as outlined earlier in this disclosure. Certain derivatives may have desirable effects for particular uses. For example, humanized or single-chain variable region molecules may be desirable for human administration, because they are less likely to stimulate an anti-mouse isotype response. Fusion proteins comprising a cytokine may be more immunogenic.

Preferred uses of these compounds include elicitating an antibody response to 1A7 or more preferably GD2, eliciting a T cell response to 1A7 or more preferably GD2, and treating various types of GD2-associated cancer. These uses are elaborated more fully in a later section.

Pharmaceutical Compositions and Vaccines Comprising 1A7 Antibody and Polynucleotide and Polypeptide Derivatives

The present invention encompasses pharmaceutical compositions and vaccines containing 1A7 antibody, or polynucleotide or polypeptide derivatives of 1A7 either alone or in
combination. Such pharmaceutical compositions and vaccines are useful for eliciting an immune response and treating GD2-associated diseases, either alone or in conjunction with other forms of therapy, such as chemotherapy or radiotherapy.

Pharmaceutical compositions include vaccines for direct administration to an individual. Vaccines may comprise IA7 antibodies, polynucleotides, or polypeptide derivatives, or a combination thereof.

Vaccines containing naked IA7 polynucleotides can be used for genetic immunization (see generally Tang et al. (1992) Nature 356: 152-154). Once in the cell nuclei, plasmids comprising IA7 encoding regions may persist as circular non-replicating episomes leading to dose-dependent and long-lived expression. (Spoon et al. (1995) Gene Therapy 2:173-180). Immunization using polynucleotides has been shown to generate cellular as well as humoral responses (Spoon et al.; Wang et al. (1995) Human Gene Therapy 6:407-418). Genetic immunization has many of the advantages of live or attenuated microorganisms as vehicles for eliciting an immune response, without the risk of infection. Preferably, IA7 polynucleotides are in the form of plasmid vectors containing appropriate control sequences for transcription and translation, such as promoters, enhancers, and signal sequences. One or more IA7 polynucleotides can be used within a single cloning vector, or multiple vectors can be used. A preferred IA7 encoding region for use in a polynucleotide vaccine encodes a IA7 scFv. The polynucleotide vaccine may also comprise encoding regions for other substances which will enhance the immune response. A preferred example is GM-CSF.

Another preferred embodiment of an IA7 polynucleotide suitable for use as a vaccine is a viral vector. Examples include adenovirus, adeno-associated retroviruses (AAV), and SV40. Preferably is a vaccinia vector, whereby the polypeptide encoded by the IA7 polynucleotide is expressed along with the immunogenic viral particle. Recombinant vaccinia comprising polynucleotides encoding IA7 polypeptides such as scFv may be used for direct vaccination at about 10^6 to 10^8 plaque forming units per dose. Vaccinia can be administered parenterally, by subcutaneous or intramuscular injection, for example, as well as through mucosal membranes, such as nasally, orally, or by inhalation. Alternatively, vaccinia can be administered via vaccinia-infected cells. In this technique, suitable cells, such as tumor cells, are infected with vaccinia in culture. The infected cells are then reintroduced to the individual. Methods for infecting cells with vaccinia and reintroducing these infected cells, have been described (see, e.g., Moss (1991)).

Other IA7 polynucleotide vaccines may be designed using other delivery vehicles known in the art. Another such delivery vehicle is cationic liposomes, to which DNA may be readily attached by virtue of its charge.

Vaccines can also be prepared from the IA7 antibody, polypeptide derivatives thereof, or a combination thereof. A protein can optionally be treated chemically to enhance its immunoresponse, especially if 100 amino acids or less. This may include cross-linking, for example, with glutaraldehyde; or linking to a protein carrier, such as keyhole limpet hemocyanin (KLH) or tetanus toxoid.

The preparation of pharmaceutical compositions that contain IA7 antibody, or polynucleotide or polypeptide derivative as an active ingredient is conducted in accordance with generally accepted procedures for the preparation of pharmaceutical preparations. See, for example, Remington’s Pharmaceutical Sciences 18th Edition (1990), E. W. Martin ed., Mack Publishing Co., PA. Depending on the intended use and mode of administration, it may be desirable to process the active ingredient further in the preparation of pharmaceutical compositions. Appropriate processing may include sterilizing, mixing with appropriate non-toxic and non-interfering components, dividing into dose units, and enclosing in a delivery device.

Liquid pharmaceutically administrable compositions can, for example, be prepared by dissolving or dispersing a vector embodied herein in a liquid excipient, such as water, saline, aqueous dextrose, glycerol, or ethanol. The composition may optionally also contain other medicinal agents, pharmaceutical agents, adjuvants, carriers, and auxiliary substances such as wetting or emulsifying agents, and pH buffering agents.

Protein vaccines of this invention typically comprise an adjuvant, which may be the same as or in addition to the excipient or carrier. Examples of adjuvants include but are not limited to aluminum hydroxide, alum, QS-21 (U.S. Pat. No. 5,057,540), DHEA (U.S. Pat. Nos. 5,407,684 and 5,077,284) including its precursors and modified forms (e.g., DHEA-S, the sulfonylated form of DHEA), β2 microglobulin (WO 91/16924), muramyl dipeptides, muramyl tripeptides (U.S. Pat. No. 5,171,568), monophosphoryl lipid A (U.S. Pat. No. 4,436,728; WO 92/16231) and its derivatives, as well as various forms and generations of DETOX™ and BCG (U.S. Pat. No. 4,726,947). Other suitable adjuvants are aluminum salts, squelaine mixtures (SAF-1), muramyl peptide, saponin derivatives, mycobacterium wall preparations, mycolic acid derivatives, nonionic block copolymer surfactants, Quil A, cholera toxin subunit, polyphosphazene and derivatives, and immunostimulating complexes (ISCOMs) such as those described by Takahashi et al. (1990) Nature 344:873-875. For veterinary use and for production of antibodies in animals, complete and incomplete Freund’s adjuvant can be used. The choice of an adjuvant will depend, in part, on the stability of the vaccine in the presence of the adjuvant, the route of administration, and the regulatory acceptability of the adjuvant, particularly when intended for human use. For instance, alum is approved by the United States Food and Drug Administration (FDA) for use as an adjuvant in humans.

A preferred vaccine composition comprising IA7 antibody or peptide derivative is prepared by mixing with aluminum hydroxide and incubated to about 48°C for about 30 min. Even more preferred are vaccine compositions comprising IA7 and QS-21 or RIBITMPC. QS-21 and RIBITMPC are equally preferred; and the selection between them is made on the basis of availability.

Pharmaceutical compositions of the present invention are administered by a route appropriate for the form of composition. Possible routes include intracutaneous, subcutaneous, intramuscular, intraperitoneal, intradermal, oral, intranasal, intradermal, and intrapulmonary (i.e., by aerosol). Protein vaccines of this invention for human use are typically administered by a parenteral route, most preferably subcutaneous. A series of injections is preferably given at different subcutaneous sites.

Pharmaceutical compositions for oral, intranasal, or topical administration can be supplied in solid, semi-solid or liquid forms, including tablets, capsules, powders, liquids, and suspensions. Compositions for injection can be supplied as liquid solutions or suspensions, as emulsions, or as solid forms suitable for dissolution or suspension in liquid prior to injection. For administration via the respiratory tract, a preferred composition is one that provides either a solid or liquid aerosol when used with an appropriate aerosolizer device. Although not required, pharmaceutical compositions are preferably supplied in unit dosage form suitable for administration of a precise amount. Also contemplated by this invention are slow release or sustained release forms, whereby a
relatively consistent level of the active compound are provided over an extended period. It is recognized that a number of alternative vaccine compositions, not limited to those described herein, may be efficacious in inducing an immune response. All such compositions are embodied within the present invention, providing they include a 1A7 polynucleotide or polypeptide as an active ingredient. The pharmaceutical compositions of this invention can be used in conjunction with other modes of therapy, whether established or experimental, whenever his is clinically desirable.

Testing 1A7 Compounds and compositions for the Ability to Elicit a Specific Immune Response

Compounds embodied in this invention may be assessed for their ability to elicit a specific immune response. Accordingly, test compounds are prepared as a suitable pharmaceutical composition and administered to test subjects. Initial studies are preferably done in small animals such as mice or rabbits, optionally next in non-human primates and then ultimately in humans. Immunogenicity is preferably tested in individuals without a previous anti-1A7 response. A test composition in an appropriate test dose is administered on an appropriate immunization schedule. It may be appropriate to compare different doses and schedules within the predicted range.

Samples (usually blood samples) are collected regularly during treatment, and assayed for a specific immune response. The response may include antibody, helper-inducer T cells, or cytotoxic T cells, or a combination thereof. As a screening test, the samples may be measured for an anti-1A7 response. Since the objective is to identify compositions useful in cancer therapy, the samples are preferably measured for an anti-GD2 response, as manifest in direct or inhibition type experiments.

This section outlines some non-limiting examples of assays that are suitable to monitor the immune response.

Presence of anti-1A7 (Ab3) and anti-GD2 (Ab1) activity in a humoral response is preferably determined after first pre-incubating sera with autologous immunoglobulin or adsorbing on a suitable affinity resin to remove antibody activity against isotypic and allotypic determinants. The adsorbed serum is then tested for Ab3 or Ab1 activity, for example, using ELISA or RIA. For instance, different dilutions of pre-reacted sera are reacted with 1A7 (or 1A7 polypeptide) coated on microtiter plates. An unrelated Ab2 serves as a control. After washing, the Ab3-1A7 complex is labeled using, for example, 125I-labeled 1A7 in a homogeneous sandwich assay. Results from this assay are compared to those obtained before administration of the 1A7 polypeptide (Example 1). Alternatively, binding to GD2 positive cells, such as M21/P6 cells, can be tested using immune flow cytomtery. In a third example, the specificity of Ab3 is determined by Western blot GD2 is separated by SDS-PAGE and blotted to a nitrocellulose filter. The filter is then incubated with sera containing Ab3, and the reaction developed by a suitably labeled anti-immunoglobulin. If the Ab3 binds to GD2, a band at the appropriate molecular weight should appear.

If desired, the specificity of the Ab3 can be further characterized. For example, competition assays can be performed to determine whether Ab3 share Ab1 idiotopes. Accordingly, competition experiments are conducted in which Ab3 is tested for inhibition of binding between 1A7 and 14G2a. Inhibition indicates that Ab3 and 14G2a contain at least some similar binding determinants. Competition of Ab3 with the binding of 14G2a to GD2 may also be measured.

Another way of characterizing a composition of this invention is testing its ability to elicit an antibody that is cytotoxic. For determination of complement mediated cytotoxicity (CMC), M21/P6 target cells expressing GD2 are labeled with 51Cr. Labeling may be accomplished by incubating about 10⁷ cells with approximately 200 μCi Na₂CrO₄ for 60 minutes at 37°C, followed by washing. The assay is conducted by adding and incubating serum suspected of containing antibody. Guinea pig serum pre-adsorbed with M21/P6 cells (or other source of complement) is then added. After a suitable incubation period at 37°C, extent of 51Cr release is then measured and compared with that of unopsonized control cells. Release of 51Cr correlates with CMC activity (Herlyn et al. (1981) Int J Cancer 27:769).

Another way of characterizing a composition of this invention is by testing its ability to elicit an anti-GD2 antibody that participates in an ADCC response (Cheresh et al. (1986) Cancer Research 46:5112-5118). In this assay, cultured human M21/P6 cells (which express GD2 in their surface) are labeled with 51Cr and are used as target cells. Normal human peripheral blood mononuclear cells (PBMC) are used as effector cells. The Ab3 containing serum from immunized subjects is supplied to mediate the ADCC reaction. Preferably, the ADCC assay is conducted in the presence of heat-inactivated serum with an effector to target cell ratio of 100:1 for 4 hours, although other suitable conditions may be used. The amount of 51Cr released is then measured.

The 1A7 antibodies, polynucleotides and polypeptides of this invention can also be characterized by their ability to elicit a cellular immune response. As used herein, a “cellular immune response” is a response that involves T cells, and can be observed in vitro or in vivo. Subjects are immunized with the 1A7 composition, and cells are recovered for assaying the response. Where the subject is a small animal, cells are typically obtained from the spleen. For larger animals including humans, cells are typically obtained from peripheral blood. A suitable cell population is recovered from the sample by standard separation techniques, typically involving centrifugation over an appropriate medium such as FICOLL/HYPAQUE™.

One way of detecting a cellular immune response is by assaying for T cell proliferative activity. In this test, cellular immune response is measured by proliferation of peripheral blood mononuclear cells (PBMCs) incubated with 1A7 polypeptide. PBMCs are isolated after a requisite number of administrations of 1A7 polypeptide, and are incubated with varying concentrations of 1A7 polypeptide. A non-specific mitogen such as PHA serves as a positive control; incubation with an unrelated anti-idiotypic antibody serves as a negative control. Preferably, the stimulator cells are autologous with the responder cells, particularly in terms of histocompatibility Class II antigens. After incubation of the PBMCs for an appropriate period (typically 5 days), [3H]thymidine incorporation is measured. If desired, determination of which subset of T cells are proliferating can be performed using flow cytometry. Optionally, splenic T cells can be pre-depleted of either CD4⁺ or CD8⁺ cells before the proliferation assay by incubation with monoclonal antibody RL.172 (anti-CD4⁺) or monoclonal antibody 168 (anti-CD8⁺) and complement.

Another way of detecting a cellular immune response is to test for T cell cytotoxicity (CTL) activity. In this test, an enriched T cell population are used as effectors in a standard 51Cr release assay (Kantor et al. (1992) J. Nutl Cancer Inst. 84:1084-1091). An example of a 51Cr release assay is the following. GD2-positive tumor cells (typically 1x10⁵ cells) are radiolabeled as target cells with about 200 μCi of Na₂CrO₄ (Amersham Corp., Arlington Heights, Ill.) for 60 min-
utes at 37°C, followed by thorough washing to remove unincorporated isotopes. T cells and targets (1×10^6/well), both resuspended in culture medium, are then be combined at various effector-to-target ratios in 96-well, U-bottom plates (Costar Corp.). The plates are centrifuged at 1000g for 5 minutes to initiate cell contact, and are incubated for 4 or 16 hours at 37°C, with 5% CO₂. After incubation, supernatants are collected using a Supernatant Collection System (Skatron, Inc., Sterling, VA) and radioactivity will be quantitated in a gamma counter (Beckman Instruments). Spontaneous release of ^51Cr is determined by incubation of targets in the absence of effectors, while maximum or total release of ^51Cr will be determined by incubation of targets in 0.1% TRITON X-100. Percentage of specific release of ^51Cr is calculated in relation to spontaneous and maximal release.

Another way of what a 1A7 polypeptide is testing its ability to ameliorate, delay the progression or reduce the extent of GD2-associated disease, as outlined in the following section.

Use of Pharmaceutical Compositions for Eliciting an Immune Response and Treating Disease

Compositions embodied in this invention such as those outlined in the previous section may be used for administration to individuals. They may be administered for experimental purposes, or to obtain a source of anti-GD2.

Compositions of this invention are particularly suitable for administration to human individuals with a GD2-associated disease. A GD2 associated disease is one in which expression of the GD2 ganglioside is altered at the affected tissue site, usually an elevation in cell-surface expression. Relevant diseases are those in which an active immune response against GD2 would confer a clinical benefit. Especially relevant are GD2-associated cancers; particularly melanoma, neuroblastoma, glioma, sarcoma, and small cell lung cancer.

The compositions of this invention may be administered to an individual with one of several objectives in mind. For example, the various compositions of this invention may be used to elicit an immune response. This includes an anti-1A7 specific response, and more preferably an anti-GD2 response. The desired response may be a specific antibody response; a specific T helper-inducer response, or a specific cytotoxic T cell response. An ADCC response or a cytotoxic T cell response is especially preferred in the context of cancer therapy, since these arms of the immune system are believed to be important effector elements in immune surveillance. The presence of an antibody response may provide a convenient means for routine clinical monitoring. Thus, a response that involves several components of the immune response in combination is especially preferred.

Assays for measuring and characterizing antibody response, ADCC, antibody-mediated cytolytic activity, T cell proliferative activity, and cytotoxic T cell activity are all described elsewhere in this disclosure.

Also included in this invention are methods for tainting GD2-associated disease, such as a tumor expressing GD2. The method comprises administering an amount of a pharmaceutical composition effective to achieve the desired effect by it palliation of an existing tumor mass or prevention of recurrence.

Dose

For treatment of a GD2-associated disease in vivo, the amount of a pharmaceutical composition administered is an amount effective in producing the desired effect. An effective amount may be provided in one or a series of administrations.

For intact 1A7, a mouse requires approximately 100 μg of KLH-coupled 1A7 emulsified in CFA and IFA in each of at least three and typically at least four administrations.

Monkeys require approximately 2 mg. The range of intact 1A7 that can be appropriately administered to humans is from about 10 μg to 20 mg, preferably 200 μg to 15 mg, more preferably 500 μg to 10 mg, still more preferably 1 mg to 4 mg, and even more preferably 2 mg. Smaller peptides and fusion proteins may be more potent on a per-weight basis, and the preferred dose may be lower than with the intact molecule. Appropriate doses can easily be determined by comparing various doses of intact 1A7 and derivatives thereof in animal models, and scaling appropriately for human use.

The amount of 1A7 polynucleotide to be administered will depend upon several factors, such as the route of administration, the condition of the individual, and the desired objective. Typically, if administered directly, the amount per administration is about 10 μg to 1 mg, preferably 25 μg to 500 μg, more preferably 30 μg to 250 μg, even more preferably 50 to 100 μg.

Administrations are typically conducted on a weekly or biweekly basis until a desired, measurable parameter is detected, such as elicitation of an immune response. Administration can then be continued on a less frequent basis, such as biweekly or monthly, as appropriate.

The various compounds of this invention can be used alone, or in conjunction with other active agents that promote the desired objective, or provide a desirable adjunct therapy.

The exact dose and timing for the administration of any composition of this invention depends on the individual to be treated, the capacity of the individual’s immune system to synthesize antibodies, the route of administration, the degree of protection desired, and the immunological and clinical response to previous doses. The immunological response may be assessed by assays given in the previous section. Choosing an appropriate amount to administer in accordance with the guidelines suggested are the responsibility of the administering physician.

Appropriate Subjects for Therapy, and Desirable Effects

Suitable subjects include those who are suspected of being at risk of a pathological effect of any GD2-associated condition are suitable for treatment with the pharmaceutical compositions of this invention. Those with a history of a GD2-associated cancer are especially suitable.

The clinical studies described in the Example section are designed to exclude subjects who have not been treated previously with mouse immunoglobulin. The concept is that a proportion of subjects who have been so treated may have circulating anti-mouse immunoglobulin (HAMA). This selection criteria has been implemented to facilitate initial testing. However, the presence of HAMA is not believed to be an impediment to therapy, since the purpose of the 1A7 is to elicit a response, not remain in the circulation. Patients are more typically chosen for therapy irrespective of their history of previous treatment with mouse immunoglobulin. Of course, for certain engineered compounds like humanized 1A7 and scFv, most mouse isotype determinants have been deleted, and the presence of HAMA in a potential recipient is even less of a consideration.

Suitable human subjects for therapy comprise two groups, which may be distinguished by clinical criteria. Patients with “advanced disease” or “high tumor burden” are those who bear a clinically measurable tumor. A clinically measurable tumor is one that can be detected on the basis of tumor mass (e.g., by palpation, CAT scan, or X-ray, positive biochemical or histopathological markers on their own are insufficient to identify this population). A pharmaceutical composition embodied in this invention is administered to these patients to elicit an anti-GD2 response, with the objec-
tive of palliating their condition. Ideally, reduction in tumor mass occurs as a result, but any clinical improvement constitutes a benefit. Clinical improvement includes decreased risk or rate of progression or reduction in pathological consequences of the tumor.

A second group of suitable subjects is known in the art as the “adjuvant group.” These are individuals who have had a history of a GD2-associated cancer, but have been responsive to another mode of therapy. The prior therapy may have included (but is not restricted to) surgical resection, radiotherapy, and traditional chemotherapy. As a result, these individuals have no clinically measurable tumor. However, they are suspected of being at risk for progression of the disease, either near the original tumor site, or by metastasis.

This group may be further subdivided into high-risk and low-risk individuals. The subdivision is made on the basis of features observed before or after the initial treatment. These features are known in the clinical arts, and are suitably defined for each different GD2-associated cancer. Features typical of high risk subgroups are those in which the tumor has invaded neighboring tissues, or who show involvement of lymph nodes.

A pharmaceutical composition embodied in this invention is administered to patients in the adjuvant group, or in either of these subgroups, in order to elicit an anti-GD2 response. Ideally, the composition delays recurrence of the cancer, or even better, reduces the risk of recurrence (i.e., improves the cure rate). Such parameters may be determined in comparison with other patient populations and other modes of therapy.

Of course, cross-overs between these two patient groups are possible, and the pharmaceutical compositions of this invention may be administered at any time that is appropriate. For example, 1A7 therapy may be conducted before or during traditional therapy of a patient with high tumor burden, and continued after the tumor becomes clinically undetectable. 1A7 therapy may be continued in a patient who initially fell in the adjuvant group, but is showing signs of recurrence. The attending physician has the discretion to determine how or when the compositions of this invention are to be used.

It is recognized in the art that the immunological status of each of the aforementioned category differs one from another by several criteria. For example, patients with active disease are generally immunosuppressed, either due to tumor-related pathology or to recent radiotherapy or chemotherapy. Their immune system may be under a barrage of tumor-associated antigen from the tumor site. On the other hand, patients who are in remission may have stronger active suppression against autoantigens. Accordingly, the ability of an anti-idiotype based vaccine to elicit an anti-tumor response, or improve the clinical condition, must be determined separately for each patient category.

Other Clinical Indications

Various compounds and compositions of this invention have other clinical indications, of which this section provides only a survey.

One indication is the treatment of cells ex vivo. This may be desirable for experimental purposes, or for treatment of an individual with a GD2-associated disease. In one example, the 1A7 antibody, or a polynucleotide or polypeptide derivative are administered to a culture of cells, such as peripheral blood cells obtained from a donor, or a suitable cell line. This may be done, for example, with the objective of stimulating T cell activity. About 0.5 to 2 μg/mL of 1A7 is an effective dose for this purpose. If desired, the stimulated cells may then be administered to a recipient, in an effort to convey passive immunity. In a second example, donor cells are genetically altered with an expression vector of this invention, to provide for ongoing secretion of 1A7 antibody after administration of the cells to the recipient.

The invention also encompasses compositions and methods using 1A7 antibodies and polypeptide derivatives to remove a label (particularly a radiolabel) from an individual who has received a labeled anti-GD2 antibody (such as 14G2a) in the course of radioscintigraphy or radiotherapy. Effective imaging using radiolabeled antibodies is hampered due to excess circulating radiolabeled antibody, which often takes several days to clear. Accordingly, 1A7 antibody or a polypeptide derivative is administered to the individual at a specified time after administration of the labeled anti-GD2. The intention is for the 1A7 polypeptide to complex with any GD2 at sites other than the tumor, such as in the circulation and interstitial spaces, and thereby promote its clearance. As a result, the level of label in unaffected tissues is reduced, and the image of the tumor (in comparison to neighboring tissues) is enhanced. Similarly, when radiomolecules are given to subjects for irradiation of a tumor site, it is desirable to reduce collateral exposure of unaffected tissue. This invention thus includes methods of treatment in which a radiolabeled anti-GD2 antibody is administered in a therapeutic dose, and followed by a molar excess of 1A7.

In either of these applications, an amount of 1A7 polypeptide is chosen that is in sufficient molar excess over the labeled anti-GD2 to locate and bind any anti-GD2 that is not localized at the tumor site. The timing of administration and amount of 1A7 polypeptide will depend upon the nature of the radiolabeled antibody, the type of radioisotope used and the condition of the individual. Preferably, the molar ratio of 1A7 polypeptide to the anti-GD2 antibody is at least about 5:1, more preferably about 25:1 to 200:1. Preferably, 1A7 polypeptide is administered 5 to 24 hours after the individual has received the anti-GD2 antibody.

The invention also includes methods of detecting the presence of an anti-GD2 antibody bound to a tumor cell comprising the steps of treating an individual with 1A7 for a sufficient time to allow binding to the anti-GD2 antibody, and detecting the presence of any complex formed. The invention is for the 1A7 to detect anti-GD2 that has pre-attached to the tumor cell; or alternatively, to promote the binding of anti-GD2 to the tumor cell by forming a polyclonal anti-GD2/1A7 immune complex. In the former instance, the anti-GD2 is provided with a detectable label or a means by which a label can be attached. In the latter instance, either the anti-GD2 or the 1A7 is provided with a label. Suitable labels include radiolabels such as 111In, 131I and 99mTc. The anti-GD2 and 1A7 are administered (usually sequentially) into the subject and allowed to accumulate at the tumor site. The tumor is then detected or visualized using standard techniques of radioscintigraphy.

Diagnostic Kits

The present invention encompasses kits containing 1A7 antibodies, polynucleotides, or polypeptides. Diagnostic procedures using the 1A7 polynucleotides or polypeptides of this invention can be performed by diagnostic laboratories, experimental laboratories, practitioners, or private individuals. Kits embodied by this invention include those that allow someone to conduct an assay for anti-GD2 or anti-1A7 activity, or for an 1A7 encoding sequence. An alteration in one of these components resulting, for example, from the presence of a GD2-associated disease or treatment directed towards it is typically compared with that in a sample from a healthy individual. The clinical sample is optionally pre-treated for enrichment of the target being tested for. The user then applies
a reagent contained in the kit in order to detect the changed level or alteration in the diagnostic component.

Each kit necessarily comprises the reagent which renders the procedure specific: a reagent 1A7 antibody or polypeptide, used for detecting anti-1A7 or anti-GD2 in the sample; or a reagent 1A7 encoding polynucleotide, used for detecting a 1A7 encoding polynucleotide in the sample. Optionally, the reagent may be conjugated with a label to permit detection of any complex formed with the target in the sample. In another option, a second reagent is provided that is capable of combining with the first reagent after it has found its target. For example, labeled anti-mouse IgG may be provided as a secondary reagent for use with intact 1A7. Labeled avidin may be provided as a secondary reagent when the primary reagent has been conjugated with biotin.

The kits may be employed on a variety of biological samples, including both liquid samples, cell suspensions and tissue samples. Suitable assays using 1A7 antibodies, polypeptides, and polynucleotides that can be supplied in kit form include those described elsewhere in this disclosure. Each reagent is supplied in a solid form or liquid buffer that is suitable for inventory storage, and later for exchange or addition into the reaction medium when the test is performed. Suitable packaging is provided. The kit may optionally provide additional components that are useful in the procedure. These optional components include buffers, capture reagents, developing reagents, labels, reacting surfaces, means for detection, control samples, instructions, and interpretive information.

Deposit
The foregoing disclosure provides, inter alia, detailed methods for preparing monoclonal antibody 1A7, along with 1A7 encoding polynucleotides, 1A7 polypeptide fragments, and other derivatives.

A practitioner of ordinary skill in the art may practice embodiments of this invention by referring to the sequence data for 1A7, which is provided herein. Alternatively, a practitioner may practice the invention by first purifying the 1A7 antibody, or a 1A7 encoding polynucleotide from a 1A7 antibody producing cell. A hybridoma cell line producing 1A7 antibody has been deposited with the American Type Culture Collection (ATCC) under terms of the Budapest Treaty, and has been given Accession No. HB-11786.

The following examples are provided to illustrate but not limit the present invention.

STATEMENT OF DEPOSIT
A deposit of the hybridoma producing the 1A7 monoclonal antibody was made prior to the filing date of the above-identified patent application under the terms of the Budapest Treaty with the American Type Culture Collection, Parklawn Drive, Rockville, MD, USA, Accession No. HB-11786.

During the pendency of this application, access to the deposit will:

(a) be forwarded to one determined by the Commissioner to be entitled thereto;

(b) all restrictions imposed by the deposit on the availability to the public of the deposited material will be irrevocably removed upon the granting of the patent;

(c) the deposit will be maintained for a period of at least thirty years or at least five years after the most recent request for the furnishing of a sample of the deposited material; and

(d) the deposit will be replaced should it become necessary due to inviability, contamination or loss of capability to function in the manner described in the specification.

EXAMPLES

Example 1

Generation and Characterization of 1A7 Anti-Idiotypic Antibody

The monoclonal anti-idiotype antibody producing hybridoma cell line 1A7 was created and identified according to the following description. Aspects of both the immunization procedure and the screening procedure were important to obtain an antibody with the desired specificity and functionality. 1A7 was one of a number of Ab2 that were initially produced, and was identified as the candidate with the most desirable features.

1A7 was obtained by using the 14G2a mouse monoclonal antibody as immunogen for an anti-idiotype response. 14G2a binds to a unique epitope of GD2 that is not present on other members of the ganglioside family. Since the responding animal was also a mouse, the Ab2 generated were expected to be directed against idiotypic features of 14G2a. However, only a fraction of those would be directed against the 14G2 paratope, an even smaller proportion would be immunogenic and able of eliciting an Ab3, and a still smaller proportion would elicit Ab3 that cross-reacted with the tumor-associated antigen.

To render 14G2a sufficiently immunogenic in an autologous species, it was conjugated to the carrier KLH, and emulsified in Freund's adjuvant. It was administered repetitively into the recipient animals on an unusual schedule with only 2 weeks between doses. Five mice were immunized according to this schedule. Substantial responses arose in about 3 mice only after the fourth immunization. Responding animals were boosted with a fifth dose of 14G2a intravenously, spleen cells were isolated, and hybridomas were prepared separately from each animal. Cloning was performed according to standard techniques.

The screening procedure comprised four important steps: (1) Positive selection for antibody binding to 14G2a; (2) Negative selection against antibody recognizing isotypic or allotypic determinants; (3) Positive selection for an ability to inhibit the binding of 14G2a to GD2; and (4) Positive selection for an ability to induce a humoral immune response against the original tumor-associated antigen (GD2) in both mice and rabbits. The rest of this section provides an overview of the screening procedure, which is given in more detail in the sections that follow.

Initial screening was conducted by immunoassay to identify the clones that reacted with 14G2a, but not with other target monoclonal antibodies sharing the same allotypic or isotypic determinants. A critical assay was a sandwich RIA in which 14G2a is attached to a solid phase, overlaid with culture supernatant, and developed with radioiodinated 14G2a. This assay requires the antibody in the hybridoma supernatant to be functionally bivalent, and be able to span between the capture 14G2a and the developing 14G2a. Several clones that were idiotype specific and gave a strong signal in this assay were selected for further study.

Subsequent screening was conducted by competition assays, in which the Ab2 was required to block the binding of 14G2a to GD2. This established that Ab2 recognized the paratope of 14G2a. GD2 was provided in the form of M21/P6 cells, a human melanoma cell line expressing GD2 at the cell surface. The nature of the assay requires the Ab2 to block the interaction between 14G2a and the tumor antigen in its particular manner of presentation on tumor cells. At a minimum, candidate Ab2 which had passed the earlier screening tests
were required to inhibit the binding of 14G2a to the cells by at least 75%. There were about three Ab2 that substantially exceeded the minimum, with 1A7 providing about the highest level of inhibition.

The ultimate screening test was a determination of whether the candidate Ab2 were capable of eliciting an Ab3 of the desired specificity when injected into a recipient. Sufficient quantities of Ab2 were prepared from mouse ascites, and tested in mice and rabbits. Sera from the test animals were first assayed for the presence of Ab3 in a sandwich immunoassay using the same labeled Ab2 used for immunization. Sera testing positively were then assayed for ability of the Ab3 to react against the tumor-associated antigen, namely GD2. Preparation of GD2 was used to coat microtiter plates, overlaid with the test serum in serial dilutions, and the Ab3 that bound was detected using labeled anti-immunoglobulin. The titer of the Ab3 binding to GD2 defined the "quality" of Ab2, as a reflection of its capacity as an inducer of anti-GD2.

Monoclonal antibody 1A7 emerged as the anti-idiotypic with the highest quality, and is the original basis for various compounds, compositions, and procedures embodied in this invention. The cell line producing 1A7 was recloned twice by limiting dilution, to ensure the stability of the line.

Materials

Antibody: The hybridoma cell line producing monoclonal antibody 14G2a was obtained from the Scripps Research Institute. 14G2a has been subtyped as an IgG2a. The specificity of 14G2a was confirmed by immunoperoxidase staining and flow microfluorimetry analysis using cells expressing GD2. Other monoclonal and myeloma mouse immunoglobulins were used as controls in various experiments herein described.

Ascites of 14G2a hybridomas and other cell lines were prepared by injection individual Pristane-primed mice i.p. with 2-10×10^7 viable cells. The IgG fraction was isolated from ascites by 45% saturated ammonium sulfate precipitation and subsequent chromatography on Protein A SEPHAROSE™ CL-4B (Iy et al. (1978) Immunochemistry 15:429). The purity of the isolated IgG was checked by immuno-diffusion, immunoelectrophoresis, and high pressure liquid chromatography (HPLC) fractionation.

Coupling of antibody with KLH: 14G2a was coupled to keyhole limpet hemocyanin (KLH) according to a method described by Maloney et al. (1985; Hybridomas 4:191), Antibody stock solution (1 mg/ml) was mixed with KLH (1 mg/ml) in PBS in the presence of freshly diluted glutaraldehyde solution (final concentration 0.05%). The mixture was rotated end-over-end for 1 h at room temperature, and then dialyzed exhaustively against PBS at 4°C.

Immunization of syngeneic BALB/c mice: BALB/c females were immunized four times over a period of 2 months. The first injection was given i.p. using 100 μg of 14G2a, emulsified in complete Freund’s adjuvant. The next two injections were given with 100 μg of 14G2a coupled to KLH in incomplete Freund’s adjuvant, either s.c. or i.p. Mice were bled from time to time, and sera were checked for anti-Id activity by ELISA in a binding assay by using F(ab')2 fragments of 14G2a and normal pooled BALB/c mouse serum IgG as control. Three days before the fusion, the mice were boosted i.v. with 14G2a in PBS.

Production of Anti-Idiotypic Hybridomas

The fusion partner used to produce the hybridomas line was the mouse non-secretory myeloma cell line P3-653, anecdotally related to P3X63Ag8.653, available from the ATCC as No. CRL-1580. Established human cell lines were cultured in RPMI 1640 supplemented with 5% fetal calf serum as described elsewhere (Scon et al. (1984) J. Immunol. 132:2089).

Hybridomas were produced essentially following the method of Oi and Herzenberg ([1980] Selected Methods of Cellular Immunology, Mishell & Shiigi eds., Freeman Publs., at 351-372). Spleen cells from immunized mice were mixed with P3-653 cells at a ratio of 1:1 to 10:1, in the presence of 50% polyethylene glycol (PEG, mw ~4500). Fused cells were then washed and cultured. Hybridlys were selected using hypoxanthine-aminopterin-thymidin media.

Initial Selection of Anti-Idiotypic Antibody (Ab2) Secreting Hybridoma Clones:

Initial screening of the hybridoma clones was performed by RIA. Purified 14G2a was radiiodinated by the Chloramine T method (Hunter (1970) Proc. Soc. Exp. Biol. Med. 133:989). The assay was conducted by coating microtiter plate wells with 14G2a antibody (or control) at 500 ng/well. After incubating overnight at 4°C, the plates were blocked with 1% bovine serum albumin (BSA) in PBS. 100 μl of hybridoma culture supernatants or 20X concentrate was incubated in the well for 4 h at room temperature. After washing with PBS, the plates were further incubated for 4 h at room temperature or overnight at 4°C with labeled 14G2a, washed, and counted. This RIA is a stringent test for antibody specificity, since it requires that the antibody be able to span between two 14G2a molecules.

A ELISA was conducted in a similar fashion, using subclass-specific anti-immunoglobulin as both the plate coat and detecting reagent. Generally, antibody of certain IgG subclasses is desired because it is stable, easily purified by protein A chromatography, and may have useful effector functions.

A number of monoclonal Ab2 secreting cell lines emerged from these screening assays with the desired properties. Amongst them was monoclonal antibody 1A7.

Confirmation that Ab2 are Specific for 14G2a Idotype: Idiotype specificity of Ab2 was confirmed by direct binding to Ab1. Various purified Ab2 were labeled with 125I, and tested for binding to plates coated with a panel of monoclonal anti-TAA Ab1. 1A7 bound almost exclusively to 14G2a: there was virtually no cross-reactivity with any of the other Ab1 tested.

Specificity for the 14G2a idotype was further established in competition experiments. ~25,000 cpm of various labeled Ab2 was mixed with different members of a panel of unlabeled competitors comprising Ab2, Ab1, and other mouse immunoglobulins. The Ab2 was then tested for binding to 14G2a-coated plates. For the best Ab2, greater than 90% inhibition was obtained using either Ab2 or 14G2a as competitor. Virtually no inhibition was obtained, up to a concentration of 5 μg, using control immunoglobulins as potential competitors.

Screening for Anti-Idiotypic Directed Against the 14G2a Paratope

To determine whether the Ab2 were directed against the paratope of 14G2a, the Ab2 were used to compete for the binding of radiolabeled 14G2a to GD2. This was performed conducted using M21/P6 cells, a human cancer cell line expressing GD2 as a membrane constituent.

To conduct the assay, M21/P6 cells were grown as confluent monolayer in 96-well tissue culture plates. Various dilutions of the test Ab2 (either culture supernantant or purified antibody) were mixed with the labeled 14G2a, and then added to the cultured cells. Percent inhibition of the assay was calculated according to the formula:
where $R_T$ is the average cpm of the experimental well with inhibitors; $R_c$ is the average background cpm; and $R_{MAX}$ is the average maximum binding without any inhibitors.

Three Ab2, including 1A7, inhibited the binding of labeled 14G2a to the GD2 expressing cells at amounts as low as about 25 ng. Purified control antibody demonstrated no inhibition.

Antibody-producing clones testing positively in the screening tests described so far were used to prepare mouse ascites as a source of Ab3. The Ab2 were purified by chromatography using Protein A and Protein G affinity resins by standard techniques.

Screening for Anti-Idiotypes Capable of Eliciting a Tumor-Specific Immune Response

Since a central purpose of these experiments was to find an anti-idiotype capable of eliciting an anti-GD2 immune response, the next screening step was to test its immunogenicity in animal models. The Ab2 would have to be not only immunogenic, but capable of raising Ab3 that cross-reacted back to the tumor antigen GD2.

Accordingly, the monoclonal antibody that gave the strongest result in the competition experiments with the GD2-expressing cells was brought forward for testing in this part of the study. The other two antibodies showing specific inhibition were held in reserve in case 1A7 failed to demonstrate the desired properties in this test.

<table>
<thead>
<tr>
<th>TABLE 1</th>
<th>Results of Immunizing BALB/c Mice with 1A7-KLH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Assay</td>
<td>Serum Dilution</td>
</tr>
<tr>
<td>Sandwich RIA (CPM)</td>
<td>1:50</td>
</tr>
<tr>
<td>% Inhibition of Ab1-Ab2 Binding by Ab3 Sera</td>
<td>1:50</td>
</tr>
<tr>
<td>% Inhibition of Ab1 Binding to M21/P6 Melanoma Cells</td>
<td>1:50</td>
</tr>
<tr>
<td>Direct Binding to GD2 by ELISA (OD405 nm)</td>
<td>1:10</td>
</tr>
<tr>
<td>PBS-BSA Control</td>
<td>0.08</td>
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</tbody>
</table>

There was no reactivity with GD2 negative cell lines or unrelated gangliosides such as GD3 and GM3. Results are expressed as mean value of triplicate determinations (S.D.<10%).

Next, allogeneic C57BL/6 mice (6–8 weeks old) were immunized with three different formulations of anti-id 1A7 vaccine: (i) 1A7-KLH and Freund’s Adjuvant; (ii) 1A7-KLH+QS-21 (10 μg per mouse); (iii) 1A7+QS-21 (10 μg per mouse). Results shown in Table 2.

<table>
<thead>
<tr>
<th>TABLE 2</th>
<th>Humoral Immune Response</th>
</tr>
</thead>
<tbody>
<tr>
<td>Immunized with</td>
<td>Sandwich RIA of Ab3 Sera (1:50 dilution) (CPM)</td>
</tr>
<tr>
<td>1A7-KLH + Freund’s</td>
<td></td>
</tr>
<tr>
<td>Mouse #1</td>
<td>4,729</td>
</tr>
<tr>
<td>Mouse #2</td>
<td>6,067</td>
</tr>
<tr>
<td>Mouse #3</td>
<td>9,391</td>
</tr>
<tr>
<td>PBS-BSA Control</td>
<td>650</td>
</tr>
<tr>
<td>1A7-KLH + QS-21</td>
<td></td>
</tr>
<tr>
<td>Mouse #1</td>
<td>5,506</td>
</tr>
<tr>
<td>Mouse #2</td>
<td>5,831</td>
</tr>
<tr>
<td>Mouse #3</td>
<td>7,315</td>
</tr>
<tr>
<td>PBS-BSA Control</td>
<td>549</td>
</tr>
</tbody>
</table>

Results are expressed as mean value of the triplicate determinations (S.D.<10%). There was no reactivity with GD2 negative cell lines or unrelated gangliosides, such as GD3 and GM3.

Comparison between the three vaccine preparations suggests that in C57BL/6 allogeneic mice 1A7-KLH and Freund’s adjuvant induced almost identical humoral immune responses as 1A7-KLH and QS-21. The production of total Ab3 response was less in 1A7+QS-21 immunized mice; however, the binding of Ab1 to melanoma cells was inhibited much more strongly and the production of anti-GD2 antibodies (Ab1') was comparable to the other two groups. Thus, there was no additional advantage of coupling of KLH to 1A7. KLH apparently induced strong anti-isotypic and anti-

Accordingly, 1A7 was prepared as a vaccine composition and injected into test animals. First, syngeneic BALB/c mice (6–8 weeks old) were immunized with 50 μg of 1A7 coupled to the carrier KLH (1:1 ratio) in the presence of equal volume (0.1 ml) of Freund’s Complete or Incomplete adjuvant on days 0, 14, 28, and 42, subcutaneously. Blood samples were drawn from each mouse 10 days after the 4th immunization and analyzed for total Ab3 response (anti-isoall/idiotype) by sandwich RIA and anti-anti-idiotype response by the inhibition of Ab1 (14G2a) binding to Ab2 (1A7 on the plate) by Ab3 sera. In addition, serum was checked for inhibition of 125I-14G2a binding to GD2 positive melanoma cells (M21/P6). Also, direct binding of sera to purified GD2, coated onto
allotypic responses in C57BL/6 mice. Immune mice sera at 1:100 dilution reacted strongly with M21/P6 melanoma cells and EL4 lymphoma cells, but not with the unrelated colon carcinoma LS174-T cells by FACS analysis.

The T cell responses of the spleen cells were also compared from the differently immunized groups of mice to various stimuli by T-cell proliferation assay. Representative data from all three groups of C57BL/6 mice and BALB/c mice are presented in Table 3.

<table>
<thead>
<tr>
<th>Stimulant</th>
<th>1A7-KLH + Freund's (BALB/c)</th>
<th>1A7-KLH + Freund's (C57BL/6)</th>
<th>1A7-KLH + Q5-21 (C57BL/6)</th>
<th>1A7+Q5-21 (C57BL/6)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anti-id 1A7 (2 μg)</td>
<td>10,919 (35.1)</td>
<td>4,428 (44.1)</td>
<td>8,388 (63.5)</td>
<td>5,692 (63.5)</td>
</tr>
<tr>
<td>Anti-id 3H1, Control (2 μg)</td>
<td>8,451 (10.1)</td>
<td>1,648 (1.4)</td>
<td>5,296 (40.0)</td>
<td>3,235 (24.4)</td>
</tr>
<tr>
<td>M21/P6 irradiated Melanoma Cells (1 x 10⁶)</td>
<td>29,671 (35.5)</td>
<td>9,648 (9.5)</td>
<td>17,753 (13.5)</td>
<td>27,739 (21.0)</td>
</tr>
<tr>
<td>EL4 Murine Lymphoma</td>
<td>n.d.</td>
<td>12,037 (11.8)</td>
<td>17,528 (13.3)</td>
<td>12,999 (9.8)</td>
</tr>
<tr>
<td>Irradiated Cells (1 x 10⁶)</td>
<td>2,973 (3.5)</td>
<td>2,074 (2.0)</td>
<td>3,944 (3.0)</td>
<td>2,340 (1.7)</td>
</tr>
<tr>
<td>LS174-T Control Colon Carcinoma Irradiated Cells (1 x 10⁶)</td>
<td>514 (0.6)</td>
<td>2,121 (2.1)</td>
<td>2,932 (2.2)</td>
<td>2,520 (1.9)</td>
</tr>
<tr>
<td>GI2 (1 μg)</td>
<td>290 (0.3)</td>
<td>1,346 (1.3)</td>
<td>1,180 (0.9)</td>
<td>1,285 (0.9)</td>
</tr>
<tr>
<td>GI55 (1 μg)</td>
<td>834 (1.0)</td>
<td>1,015 (1.0)</td>
<td>1,313 (1.0)</td>
<td>1,320 (1.0)</td>
</tr>
</tbody>
</table>

The data are expressed as mean CPM of triplicate wells (S.D.<10%). S.I. = Stimulation Index. S.I. >3.0 was arbitrarily considered as positive.

The results indicated that in C57BL/6 mice, immunization with all three regimens induced 1A7 specific, M21/P6 melanoma cell specific and EMA cells specific proliferative responses, some reactivity against control 3H1 and no reaction against control cell line LS174-T cells or ganglioside GD2 or GD3. These data support the postulate that for T cell activation, GD2 needs to be associated with cell surface oligopeptides. There was also no significant difference in Stimulation Index obtained with any of these three regimens. 1A7+Q5-21 was as good as 1A7-KLH+Q5-21 or 1A7-KLH+Freund’s in C57BL/6 mice.

In order to assess the ability of these vaccines to induce T-cell mediated DTH response, C57BL/6 mice immunized with 1A7-KLH+Freund’s, 1A7-KLH+Q5-21 and 1A7+Q5-21 were given intradermal foot pad injection of irradiated M21/P6 cells or irradiated LS174-T (control) cells. In another experiment, mice received intradermal foot pad injection of purified GD2 or purified GD3. Mice were observed for development of DTH response at the inoculation site at 24 hours and 48 hours. There were significant DTH responses directed at GD2-positive M21/P6 cells but not GD2-negative LS174-T cells in all three groups of immunized mice (data not shown).

There was, however, no DTH reactivity directed at GD2 or GD3 in any of the groups of immunized mice.

Rabbits were selected next for the immunization studies with 1A7-KLH+Q5-21 and 1A7+Q5-21. The relative composition and tissue distribution of gangliosides in rabbits is similar to that of humans. Each group of three rabbits were immunized with either 500 μg of 1A7 mixed with 50 μg of Q5-21, or 500 μg of 1A7-KLH+50 μg of Q5-21. The injections were given intramuscularly on days 0, 14, 28 and 42. Blood samples were collected 10 days after the fourth immunization, and analyzed for Ab3 (Ab1') responses and T-cell proliferation. Table 4 shows the humoral immune responseinduced.
Results are expressed as the mean value of triplicate determinations (S.D.<10%). There was no reactivity with GD2-negative cell lines or the gangliosides GD3 or GM3. The O.D. value obtained with PBS-BSA control was 0.08.

KLH-coupled 1A7 plus QS-21 induced higher levels of anti-isotypic and anti-allotypic responses in all three rabbits. Ab3 and GD2-positive cell binding inhibition reactions were better in all three 1A7+KLH+QS-21 immunized rabbits. Two out of 3 rabbits in each group raised anti-GD2 antibodies, and the response was better in 1A7+QS-21 immunized group as compared to 1A7-KLH+QS-21 group.

Thus, 1A7+QS-21 was capable of raising desired tumor responses in both mice and rabbits. There was no additional advantage of coupling 1A7 to KLH. The isotype of the anti-GD2 antibodies in the rabbit sera was mostly of IgG type with trace amount of IgM. The Ab1 antibody in rabbit sera also reacted with melanoma cells but not with GD2-negative carcinoma cells by FACS analysis.

The Ab3 sera were cytotoxic to M21/P6 and EL4 cells by in vitro ADCC assay, conducted as described elsewhere in this disclosure. Briefly, target cells were labeled with 51Cr, washed thrice with DMEM without FCS and suspended in growth medium. 10⁴ target cells in 25 μL were added to microtitre plate wells and incubated with different dilutions of sera from immunized animals and effector cells. The immune sera induced 30-40% specific lysis of the target cells.

PBL were isolated from immunized rabbits and used to study the cellular immune response in a PBL-transformation assay. Results are shown in Table 5.

<table>
<thead>
<tr>
<th>Stimulant</th>
<th>CPM</th>
<th>Stimulation Index</th>
<th>CPM</th>
<th>Stimulation Index</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anti-id 1A7 (2 μg)</td>
<td>21,329</td>
<td>4.42</td>
<td>9,219</td>
<td>4.92</td>
</tr>
<tr>
<td>Anti-id 3H1</td>
<td>11,590</td>
<td>2.39</td>
<td>5,691</td>
<td>2.85</td>
</tr>
<tr>
<td>Control (2 μg)</td>
<td>71,596</td>
<td>14.86</td>
<td>28,845</td>
<td>14.48</td>
</tr>
<tr>
<td>M21/P6</td>
<td>5,196</td>
<td>1.07</td>
<td>3,131</td>
<td>1.57</td>
</tr>
<tr>
<td>Melanoma Cells</td>
<td>44,619</td>
<td>9.26</td>
<td>28,040</td>
<td>14.08</td>
</tr>
<tr>
<td>Irradiated EL4</td>
<td>5,196</td>
<td>1.07</td>
<td>3,131</td>
<td>1.57</td>
</tr>
<tr>
<td>Irradiated L5174-T Col</td>
<td>5,196</td>
<td>1.07</td>
<td>3,131</td>
<td>1.57</td>
</tr>
<tr>
<td>Carcinoma Cells</td>
<td>11,345</td>
<td>2.35</td>
<td>5,988</td>
<td>3.00</td>
</tr>
<tr>
<td>CD2 (1 μg)</td>
<td>7,329</td>
<td>1.52</td>
<td>4,678</td>
<td>2.34</td>
</tr>
<tr>
<td>CD3 (1 μg)</td>
<td>4,816</td>
<td>1.0</td>
<td>3,191</td>
<td>1.50</td>
</tr>
</tbody>
</table>

Results are expressed as the mean value of triplicate determinations (S.D.<10%). Stimulation Index >3.0 was considered as positive.

The results demonstrate that immunization of rabbits with both 1A7-KLH+QS-21 and 1A7-S-21 induced T cell proliferation in PBL against anti-id 1A7, irradiated GD2-positive M21/P6 cells and EL4 cells but not against GD2-negative L5174-T cells or against GD2 and GD3. There was insignificant stimulation against normal isotype-matched control Ab2 (S1,5,3,0). Stimulation Index against various stimuli was almost identical in both groups of immunized rabbits.

Rabbits immunized with 1A7 were subjected to skin testing to confirm that the cellular response induced by administration of 1A7 can mediate a hypersensitivity response. The rabbits were shaved on the back and challenged with an intradermal inoculum of purified gangliosides. Slight erythema and induration was observed as a result of challenge with either GD2 or GD3. In a separate experiment, immunized rabbits were challenged with an intradermal inoculum of 1x10⁶ cells inactivated by irradiation at 12,000 rads. In a rabbit immunized with 1A7+QS-21 and challenged with M21/P6 cells (a GD2-expressing line), the induration was 13x12 mm at 24 h and 18x13 mm at 48 h. In a rabbit immunized with 1A7-KLH+QS-21 and challenged with M21/P6 cells, the induration was 18x16 mm at 24 h and 24x16 mm at 48 h. In contrast, when challenged with L5174-T cells (a GD2-negative line), the induration was 4x4 mm and 5x3 mm respectively at 24 h, and negligible at 48 h.

Example 2

Obtaining the 1A7 Heavy and Light Chain Sequences

The polynucleotide sequence was obtained for the 1A7 antibody by isolating messenger RNA from the 1A7 producing cell line. For each sequence determination, total RNA was isolated from 1x10⁶ 1A7 hybridoma cells. Messenger RNA was prepared by passage through two cycles of chromatography of oligothymidylic-cellulose columns. The yield of mRNA was about 10 μg. First strand cDNA was synthesized using SUPERSCRIPT™ Preamplification kit (GIBCO/BRL).

To sequence the heavy chain variable region, PCRs were conducted on the cDNA using a reverse primer corresponding to amino acids 126 to 119 of the murine γ, constant region:

(SEQ. ID NO: 46)

and various mixtures of forward primers, corresponding to the N-terminal leader sequences of murine variable region subgroups. The forward primer that gave a positive reaction was:

(SEQ. ID NO: 47)

and various mixtures of forward primers, corresponding to amino acids 20 to 12.

The amplified fragment of cDNA was subcloned into pT7 plasmid and NOVABLU™ competent cells were transformed using a protocol provided by the supplier (Novagen). Recombinant colonies were picked up by color selection and plasmid DNA was prepared by miniprep procedure. The DNA sequence of the double stranded plasmid was determined using a Sequenase Version 2.0 kit (USB, Cleveland, Ohio). The sequence of the DNA insert in the plasmid was determined from both orientations using primers specific for the plasmid: namely T7 promoter (TAATACGACTCACTATAGGG) (SEQ. ID NO:48) and U-19 (GTTTTCCTCAGT-CACGACGT) (SEQ. ID NO:49). At least 8 clones were picked for sequence determination.

The sequence of the 1A7 light chain variable region was determined in a similar fashion. The forward and reverse primers giving a positive result in the PCR were:
corresponding to amino acids -19 to -10 of the leader sequence, and 122 to 116 of the mouse κ chain constant region.

In order to minimize the error rates in PCR amplification, plasm DNA polymerase (Stratagene, San Diego) was used for amplification in all subsequent experiments. Mutant frequency with this thermostable DNA polymerase is 1/10 compared to Taq DNA polymerase.

Confirmation that the isolated cDNA correspond to the IAT7 heavy and light chains is obtained by amino acid sequencing of the N-terminal of the isolated antibody. Fifty μg of purified IAT7 antibody is diluted with sample loading buffer (50 mM Tris-HCl, pH 6.8, 1% SDS, 1% glycerol, 0.1% β-mercaptoethanol) and heated to 100°C for 3 minutes. The denatured protein is loaded onto a 7.5% polyacrylamide gel (BioRad MiniProtein II Dual Slab Cell) containing SDS and subjected to electrophoresis at 200 V for 1 hour. Proteins in the gels are transferred to polyvinylidene difluoride (PVDF) membranes by the procedure described by Towbin et al. ((1979) Proc. Natl. Acad. Sci. USA. 78: 4350-4354) at 150 mA overnight. The transfer buffer contains 25 mM Tds, 192 mM glycine, 20% (v/N) methanol. The membranes are stained by quick dipping in 0.1% Coomassie Brilliant Blue in 50% methanol-50% acetic acid, followed by washing in a solution containing 40% methanol plus 10% acetic acid. After drying the membrane at room temperature, the stained heavy and light chain bands are excised with a clean razor blade. The proteins on the membrane slices are subjected to N-terminal microsequencing by automated Edman degradation using an Applied Biosystem Model 477A protein sequencer employing pulsed-liquid chemistry and on-line phenyl-ethanolysis amino acid identification. Each protein is subjected to 10-15 degradative cycles and the converted cleavage products from each cycle were analyzed by reverse-phase HPLC.

The nucleic acid sequence and the corresponding translation for the heavy and light chain variable regions of monoclonal antibody IAT7 is shown in FIGS. 1 and 2.

The heavy and light chain polynucleotide and amino acid sequences were compared using the BLAST algorithm at the National Center for Biotechnology. Information with sequences available from the PDB, SwissProt, PIR, SPUdate, GenPept, and GNPupdate databases. The comparison was performed on Dec. 16, 1995.

FIG. 17 shows the ten most closely matched polynucleotide sequences to the IAT7 light chain variable region encoding sequence. FIG. 18 shows the ten most closely matched polynucleotide sequences to the IAT7 heavy chain variable region encoding sequence.

FIG. 3 is a comparative depiction of the IAT7 light and heavy chain amino acid sequence with the 15 closest sequences found in the BLAST search. Panel (A) shows the light chain comparison. Panel (B) shows the heavy chain comparison. The database identifiers for the matched sequences are shown in Table 6:

<table>
<thead>
<tr>
<th>Matched immunoglobulin amino acid sequences</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Light Chain Variable Region</strong></td>
</tr>
<tr>
<td>1 ggi: M34588/MU: SIGKABR_.1 Mouse Ig kappa-chain mRNA V-reg...</td>
</tr>
<tr>
<td>2 ggi: L1834/MU: SIG438B_.1 Mouse rearranged immunoglobulin li...</td>
</tr>
<tr>
<td>3 ggi: Z22305/MID: IGKVAH_1 immunoglobulin variable region [Mu...</td>
</tr>
<tr>
<td>4 ggi: M32857/MU: SIGKSCP_.1 Mouse Ig rearranged kappa-chain mRNA...</td>
</tr>
<tr>
<td>5 ggi: M34588/MU: SIGKABR_.1 Mouse Ig kappa-chain mRNA V-reg...</td>
</tr>
<tr>
<td>6 ggi: JO4348/MUSIKGCAW_1 Mouse Ig kappa-chain (PAC1) mRNA V...</td>
</tr>
<tr>
<td>7 ggi: M31271/MUSIKGCSM_1 IgM gene product [Mus musculus]</td>
</tr>
<tr>
<td>8 ggi: M32858/MUSIKGCSQ_.1 Mouse Ig rearranged kappa-chain mRNA...</td>
</tr>
<tr>
<td>9 ggi: U2642/MUSIKGCAW_1 antis-PC Ig kappa chain [Mus musculus]</td>
</tr>
<tr>
<td>10 ggi: X6870/MMGMMMA_1 IgM gene product [Mus musculus]</td>
</tr>
<tr>
<td>11 ggi: M36723/MUSIKGDKA_.2 immunoglobulin kappa-chain VK-1 [M...</td>
</tr>
<tr>
<td>12 prf: B39276/B39276 Ig light chain precursor V-D-J reg...</td>
</tr>
<tr>
<td>13 ggi: M34370/MUSIKGJS_.1 immunoglobulin kappa chain [Mus mu...</td>
</tr>
<tr>
<td>14 prf: A31807/A31807 Ig kappa chain V region (PAC1) - mu...</td>
</tr>
<tr>
<td>15 ggi: U29267/MUSIKGCAW_1 IgL rearranged kappa chain V-reg...</td>
</tr>
</tbody>
</table>

| **Heavy Chain Variable Region**             |
| 1 ggi: M36221/MUSIHABE_.1 immunoglobulin heavy chain V-region |
| 2 ggi: M30118/MUSIHABE_.1 immunoglobulin heavy chain [Mus mu... |
| 3 ggi: P18189/HV34_MOUSEB Ig HEAVY CHAIN PRECURSOR V-REGION... |
| 4 ggi: M36217/MUSIHADK_.1 Ig gene product [Mus musculus] |
| 5 ggi: M36217/MUSIHADK_.1 immunoglobulin heavy chain V-region... |
| 6 ggi: M36228/MUSIHAEI_.1 immunoglobulin heavy chain V-region... |
| 7 ggi: M34626/MUSIHACK_.1 Mouse Ig rearranged heavy chain [N... |
| 8 ggi: A05515/A05515_1 Vector pS2HPEO DNA sequence, fun... |
| 9 prf: 1FDFLI IgG1 Fab Fragment (Anti-Lyn:Wy-A5.1) |
| 10 ggi: L34544/MUSACLA_.1 antibody [Mus musculus] |
| 11 ggi: A03097/A03097_1 antibody D1.3 V region (VDJ) [Hom... |
| 12 prf: S8563/S8563 Ig heavy chain V region (ASW1) |
| 13 prf: A32456/A32456 Ig heavy chain precursor V region... |
| 14 ggi: A05904/A05904_1 pSW1 protein (unidentified) >pp:A0... |
| 15 ggi: L34544/MUSACLA_.1 Mus musculus (clone pCT.leva1) ant... |
Amongst the 50 database sequences matched most closely to that of the 1A7 light chain variable region, none was identical. 1A7 differed from the five closest sequences by 2 substitution differences at residues 50 and 55, which are contained in the second complementary determining region (CDR2). The two differences at these positions were non-conservative substitutions, and persisted in comparisons with other light chain sequences.

Amongst the 50 database sequences matched most closely to that of the 1A7 heavy chain variable region, none was identical. The following summarizes the main points deduced from the comparison.

The closest match was with a heavy chain fragment beginning at residue 9 (designation gpIM36221|MUSIGHAEB_1). There were 6 substitutions between residues 1 and 97 (before the VDJ junction), 6 substitution differences after residue 97, and 1A7 was shorter about the VDJ junction by 2 residues.

1A7 differed in length from all sequences but one, due to insertions or deletions of 1 to 8 residues about the VDJ junction. For the sequence of equal length (designation piriS11106/S11106), there were 18 substitution differences between residues 1 and 97, and 8 substitutions after residue 97.

All other comparisons showed at least 14 substitution differences between residues 1 and 97.

All other comparisons showed at least 4 substitution differences after residue 97.

All other comparisons showed a total of at least 22 insertions, deletions and substitution differences.

Differences appeared throughout the variable region.

Amino acid consensus sequences of the 15 most closely matched Vc and VH regions were designed, and compared with the 1A7 sequences. This is shown in FIG. 3(C). Other than splicing differences about the VDJ junction, there appear to be about 16 differences between 1A7 and the prototype sequences. Two of these differences are present in the light chain; 14 are present in the heavy chain. Seven occur in the CDRs, while nine occur in the variable region framework. The point differences likely have arisen from somatic mutation of germline variable region sequences.

Example 3

1A7 as an Immunogen in Non-Human Primates

As a model more closely related to humans, the effect of anti-id 1A7 on the induction of GD2-specific humoral responses was investigated in cynomolgus monkeys (Macaca fascicularis). The normal tissue distribution of GD2 in cynomolgus monkeys is very similar to that in human. As such, this primate model is ideal to gauge toxicities induced by these agents and to establish useful dosage for initial clinical trials.

Cynomolgus monkeys (two per group, 2-4 kg weight) received four intramuscular injections of purified Ab2 (2 mg) mixed with 100 µg of QS 21 as adjuvant. Control monkeys were immunized with unrelated Ab2, 3H11 mixed with QS 21 in the similar way. All injections were given at 2-week intervals. Monkeys were bled 10 days after each immunization.

The sera were analyzed for Ab3 responses by sandwich RIA and inhibition of Ab2 binding to Ab1. For these assays, the sera were pretreated with normal mouse immunoglobulin (500 µg/ml) to block anti-isotypic and anti-idiotype reactivities. 250 ng of 1A7 (Ab2) was coated into 96-well plate. After blocking, 50 µl of different concentrations of PRO#685 or PRO#778 (Ab3) was added and incubated 2 h at room temp with shaking. After washing, 90,000 cpm of 125I-1A7 was added to each well and incubated 1.5 h at room temp. The plate was then washed and bound radioactivity was measured.

Results are shown in FIG. 4. Ab3 sera from monkeys (PRO 685 and PRO 778) immunized with 1A7 bound specifically to the immunizing Ab2 (1A7) with minimal reactivity with unrelated Ab2 (3H11).

FIG. 5 shows that monkey Ab3 sera also inhibited the binding of radiolabeled Ab2 to Ab1 500 ng of 1A7 (Ab2) or 14G2a (Ab1) was coated into 96-well plate. After blocking, 50 µl of different concentrations of PRO#685 (Ab3) along with 50 µl of 125I-14G2a or 125I-1A7 (90,000 cpm) where added to each well. After 1.5 h incubation, plates were washed and bound radioactivity was counted. There was no inhibition with preimmune sera or sera obtained from monkeys (PRO 541 and PRO 667) immunized with the unrelated Ab2 3H11. These results indicate that monkey Ab3 sera share idiotypes with the Ab1.

To measure anti-GD2 reactivity in the serum of immunized monkeys, purified GD2 (250 ng/well) was absorbed into 96-well plates. After blocking wells with 1% BSA in PBS, test serum and Ab1 were diluted in same buffer and added to wells and incubated overnight at room temp. After washing, the bound antibodies were detected using alkaline phosphatase labeled anti-mouse or anti-human Ig reagents as second antibodies. Sera from both monkeys tested positively.

To determine whether 1A7 immunized monkey sera bound specifically to GD2 positive melanoma cells, the binding of monkey Ab3 sera to the melanoma cell line M21/P6 was tested M21/P6 cells (2x10^5) were incubated with different concentrations of PRO#685, PRO#778 (Ab3) or 14G2a (Ab1), and the amount of bound antibody was determined.

FIG. 6 shows the sera collected after the fourth immunization, reacted with melanoma cells but not with the antigen-negative MCF-7 breast cancer cell line. The Ab3 sera also bound specifically to purified GD2 coated onto microtiter plates by ELISA. Control sera from preimmune monkeys or monkeys immunized with unrelated Ab2 (3H11) did not show appreciable binding to GD2. In parallel experiments, the same Ab3s from monkey PRO 685 were compared on a plate coated with CEA (an unrelated tumor-specific antigen) and were negative.

The Ab3 antibodies were then purified from sera by absorption and elution from the affinity column made of antibody 1A7 coupled to SEPHAROSE® 4B. The eluted antibody was then passed over an immunoadsorbent column consisting of normal mouse Ig coupled to SEPHAROSE® 4B to remove anti-isotypic and anti-idiotype reactivities. Antibody that passed through was concentrated and used as purified Ab3. The purified Ab3 from monkey sera were then compared with the reactivity of purified Ab1 (14G2a) in different assays. 2.6 mg of purified Ab3 were recovered from 10 ml of sera (i.e. about 260 µg of Ab3 per ml of serum) from monkey # PRO 685 and a little less from monkey # PRO 778.

Binding of Ab3 to M21/P6 cells was analyzed by flow cytometry. Target cells M21/P6 or control cells MOLT-4 (5x10^5 in PBS supplemented with 0.2% BSA) were incubated with different dilutions of Ab3 and Ab1 for 1 h at 4° C.
After washing with PBS, the staining was done with FITC labeled second antibody and analyzed on a FACScan flow cytometer.

Results are shown in Fig. 7. In Panel A, tumor cells (M21/ P6) were reacted with preimmune sera and Ab1 sera (1:100 dilutions) from monkeys immunized with 1A7 mixed with QS-21. The reaction was developed with goat anti-human F(ab')2_2 IgG-FITC labeled antibody. In Panel B, MOLT-4 cells that do not express GD2 were reacted with preimmune and immunized monkey Ab3 sera raised against 1A7 plus QS-21. In Panel C, tumor cells (M21/P6) were reacted with PBS-BSA control and purified Ab1 (14G2a). The reaction was developed with goat anti-mouse-F(ab')2 IgG-FITC labeled antibody. The results show that Ab3 from immune but not preimmune sera was specific for GD2-bearing M21/P6 cells.

Fig. 8 shows results from an experiment in which Ab3 was shown to bind directly to the GD2 target antigen in a specific fashion. 250 ng of different gangliosides were coated into 96-well plate. After blocking, 50 ng of different concentrations of PRO/685 (Ab3) and 14G2a (Ab1) was added and incubated 4 h at room temperature. Bound antibody was detected using alkaline phosphatase conjugated secondary antibody.

Fig. 9 shows the corresponding inhibition experiment. Different gangliosides (250 ng) were coated into 96-well plates as before. Different concentrations of Ab3 and Ab1 along with 90,000 cpm of 125I-14G2a were added. The plate was incubated 2 h at room temperature with shaking, washed and counted. Percent inhibition was calculated and plotted against concentration of Ab1 and Ab3.

Reactivity of immunized sera and purified Ab3 for anti-GD2 antibodies against various gangliosides was also measured by immunoblotting (Fig. 10). Purified gangliosides (2 μg each of GM3, GM2, GM1, GD3, GD2 and GT1b) were spotted on strips of PVDF cellulose membrane at 1 cm intervals. After blocking with 3% BSA in PBS, the strips were incubated with PRO/685 (Ab3) or PRO/778 (Ab3) or 14G2a (Ab1) or an unrelated monkey Ab3 which was raised against an unrelated Ab2, 1ID10 and PBS control; each antibody was used at 10 μg/ml, 5 ml of total solution. The incubation was done for 4 h at room temperature with shaking. After washing the strips were incubated with alkaline- phosphatase labeled second antibody (1:1000 dilutions) for 2 h at room temperature.

The results clearly demonstrate that 1A7-QS21 immunized monkey Ab3 antibody binds to the same antigen GD2 as Ab1.

The ability of the induced Ab3 to mediate antibody-dependent cellular cytolysis was confirmed in a standard ADCC assay. M21/P6 cells were labeled with 51Cr, and incubated in microtiter plate wells with dilutions of immune monkey sera. Normal human PBMC isolated by FICOLL/HYPAQUE® were then added as effector cells at an effector:target ratio of 50:1. The plates were centrifuged at 400 xg for 2 min, and incubated for 4 h at 37°C in a humidified atmosphere containing 5% CO2. After incubation, the plates were centrifuged at 400 xg for 5 min, and specific lysis was calculated from the cpm released into the supernatant. The sera of two immunized monkeys (PRO 778 and PRO 685) each mediated specific lysis of 40-50% of target cells at a dilution of 1:10. In contrast, preimmune serum or the serum from a monkey immunized with an unrelated tetradisototy mediated specific lysis of only 4-8% of the labeled target cells.

The presence of a cellular immune response in the immunized monkeys was demonstrated in an in vitro proliferation assay. Mononuclear cells were isolated from the peripheral blood of immunized monkeys by FICOLL/HYPAQUE® gradient centrifugation. 5 x 106 cells per well were incubated with different concentrations of stimulant for 5 d at 37°C, and then-pulsed with 1 μCi/well [3H]-thymidine for 20 h. Stimulation index was calculated by dividing the sample cpm by the average cpm obtained in medium control wells. The SD was <10% for each determination, done in triplicate.

Result of the proliferation assay are shown in Fig. 11. Legend: Close hatching, 2 μg antibody 1A7; Open bars, 2 μg of an unrelated murine antidiotypic; Filled bars, radiation-killed M21/P6 cells (a GD2-expressing cancer line); Shaded bars, radiation-killed LS174-T cells (a GD2-negative cancer line); Light hatching, medium control. The reactivity indicated by the open bars likely represents a cellular response to non-idiotype antibody components (i.e., an anti-immunoglobulin reaction). The additional reactivity observed when stimulated with 1A7 is consistent with a cellular response to the idiotype components of 1A7. The ability of a GD2-expressing cell line but not a GD2-negative cell line to induce proliferation suggests that the cellular response comprises anti-GD2 activity.

The induction of Ab3 responses in monkeys did not cause any apparent side effects in animals despite the presence of GD2 in some normal tissues. Only mild local swelling and irritation were observed at the injection site as a result of multiple immunizations. The monkeys were routinely checked by physical examinations and weight measurements. They did not show any signs of abnormalities or neurological problems. Immunohistochemistry analysis of autopsy specimens obtained after 7 months indicated that there was no toxicity induced by the anti-Id 1A7 plus QS-21 vaccine treatment.

**Example 4**

Clinical use of 1A7 Antibody Vaccines in Patients with High Tumor Burden

9.7 g of purified 1A7 antibody have been prepared for clinical use. The antibody was purified by TSD BioServices under GMP-conditions. The regulatory testing on the antibody preparation have been completed according to FDA guidelines. Approval from the FDA has been obtained (BB-IND #6183) for use of 1A7 plus QS-21 in advanced melanoma patients.

The objectives of this study are to determine the effects of a first generation vaccine on various components of the immune response (both humoral and cellular), to determine the optimum immunomodulatory dose and toxicity of anti-Id antibody, and to determine whether there is a the benefit to clinical condition.

Eligible patients are those having metastatic melanoma that is confirmed as bearing the GD2 antigen. Patients must have a life expectancy greater than six months, adequate nutrition, non-pregnant, Southwest Oncology Group performance score 0, with all previous anti-cancer therapy for at least four weeks, no prior mAb therapy, no ongoing use of nonsteroidal anti-inflammatory agents or cimetidine or other H2 receptor antagonists, adequate blood count and the ability to sign an informed consent.

All patients in the study are immunized with either 1 mg, 2 mg, 4 mg or 8 mg of 1A7 mixed with 100 μg of QS-21 adjuvant. Patients are randomized to one of the four dose levels. The total number of patients is between about 12 and 32. Injections are given biweekly for four total doses, or until an immune response is observed. Therapy continues with monthly injections until tumor progression is found. Patients are monitored carefully for anaphylaxis, serum sickness, and other potential side effects.
Periodic blood samples are obtained to determine the effect on hematopoietic cells as well as renal and hepatic function. All patients entered into the study undergo leukapheresis prior to the first immunization (pre-therapy). In addition, blood samples are obtained prior to each injection of IA7 to determine serum levels of Ab3 and Ab1 antibodies and cytotoxic T cell responses. The specificity of the humoral responses is confirmed by immune flow cytometry, radioimmunoassay, and dot blot analysis. Antibody responses to the murine antibody are tested by sandwich RIA. Sera are also tested for the ability to inhibit the binding of anti-GD2 mAb to GD2 antigen. The immunological profile of patients is further assessed by testing the proliferative response of patient’s lymphocytes to anti-id antibody, purified GD2 antigen, and irradiated tumor cells and the cytotoxicity of patient’s lymphocytes for GD2-positive HLA-matched cell lines or autologous tumor cells (where possible).

The development of humoral immunity induced by immunization with Ab2 is assessed by testing sera obtained from patients at different time points (pre-therapy, after 3 and 4 immunizations, and then periodically before injection of therapy continues). The sera is initially tested for total human anti-murine antibody (HAMA) response (anti-isotype and anti-allotype) by sandwich RIA. Briefly, microtiter plates are coated with IA7 and incubated with different dilutions of patients’ sera. After washings, the antigen-antibody reaction is developed using 125I-labeled IA7 in a homogeneous sandwich RIA. Since IA7 is injected as intact IgG in this study, patients are expected to mount HAMA responses.

Sera from immunized patients with positive responses are then tested for the presence of anti-idiotypic antibodies (Ab3). Sera are diluted with buffer coating normal murine immunoglobulin to block human antibodies against isotypic and allotypic determinants, and then checked for the presence of anti-IA7 by RIA. Unrelated Ab2 is used as a control. After washing, the antigen-antibody reaction is developed using 125I-labeled anti-id reagent in a homogeneous sandwich RIA as above. Pre-treatment non-immune sera and sera from normal donors is used as control in these assays.

If antibody is detected against IA7, the sera are examined for its ability to inhibit the binding of 125I-labeled IA7 to 14G2a bound to microtiter plates, or inhibit the binding of radiolabeled 14G2a to IA7 on the plate. An unrelated Ab1-Ab2 system is used as a control. This demonstrates whether Ab3 in patients’ sera share idiotopes with 14G2a (Ab1). This inhibition assay of Ab1-Ab2 binding by Ab3 sera also demonstrates whether Ab3 is a true anti-idiotypic.

To assess humoral immune responses directed against native target antigens, patients’ Ab3 sera is tested for reactivity with cell lines known to express GD2 in a RIA, and also by FACS analysis, using anti-human IgG and IgM tracer reagents. In addition, the sera is checked for reactivity against a solubilized purified preparation of GD2 antigen coated onto microtiter plates. The antigen-antibody reaction is detected by using 125I-labeled anti-human Ig reagents. Pre-immune sera is used as a control. Unrelated antigen is also used in the assay. Isotype of human Ab3 sera binding to GD2 antigen is determined by ELISA using anti-human isotype specific reagents.

To demonstrate that Ab3 and Ab1 bind to the same antigenic determinant, inhibition of 14G2a binding to an Ag positive tumor cell line or GD2 antigen by Ab3 sera is determined in an RIA. If Ab3 in patients’ sera bind specifically to tumor cells, the ability of Ab3 to lyse these cell in conjunction with ADCC effector cells or complement can be demonstrated in standard ADCC or complement-mediated cytotoxicity (CMC) assays.

Quantitation of the Response:

Host Ab3 (anti-IA7 antibody) and Ab1’ (anti-tumor antibody) reactivity are a key measurement in this example. The expression of anti-idiotype antibody (Ab3) in the patients’ sera is quantitated by RIA inhibition studies as follows. Briefly, microtiter plates are coated with 14G2a (Ab1) and reacted with a fixed amount of 125I-labeled Ab2. A standard inhibition curve is generated using purified 14G2a as inhibitors. Next, patients’ sera depleted of anti-allotypic activity at different dilutions are checked for an ability to inhibit the Ab1-Ab2 reaction, and the amount of Ab1-like activity in the sera is estimated from the standard inhibition curve. The induction and duration of Ab3 response are compared among different dose levels. If there is no statistical difference between Ab3 responses or duration at a number of doses, the titer of specific anti-tumor response (Ab1’) in the sera by ELISA assay using purified GD2 antigen coated plates is compared among different dose levels.

If the serum of a particular patient tests positive for anti-IA7 (Ab3) but negative for anti-GD2 (Ab1’), it may be because the anti-GD2 is bound to the patient’s tumor cells. The production of anti-GD2 is optically demonstrated by stimulating the patients’ PBMC in culture with IA7 and then measuring anti-GD2 activity in the supernatant.

According to network hypothesis, patients immunized with IA7 may eventually also induce an Ab4 response (anti-anti-idiotype) which may mimic the specificity of the anti-idiotypic IA7. To study this possibility, Ab3 positive patients’ sera (depleted of anti-isotype and anti-allotypic antibodies) are optionally measured for anti-idiotypic activity by reacting with 14G2a (Ab1) by ELISA or RIA. Positive and negative controls are included as described for the Ab3 assay. Sera for this assay are obtained three months after the last therapy.

Cell-Mediated Immunity

Whether a specific T cell response to the tumor associated glycolipid GD2 is generated in the IA7 treated melanoma patients is tested by the following criteria: (1) if a T cell response is present which targets GD2 on the tumor cells, and (2) whether this response increases with repeated immunizations. Analysis proceeds in 2 phases. The first is to determine whether T cells from all PBMC samples received can be expanded following in vitro immunizations against the IA7. If this occurs, the next step is to determine whether these T cells, can lyse or release cytokines against autologous GD2 bearing melanoma tumor cells or allogeneic GD2 expressing melanoma cells sharing a single HLA antigen in common with the autologous CTL.

Patients entered into the study are leukapheresed prior to the first immunization and FICOLL-HYPAQUE® separated peripheral blood mononuclear cells (PBMC) are prepared and cryopreserved for future studies. These PBMC (1) provide antigen presenting feeder cells for subsequent studies (2) serve as baseline for T cell responses and (3) are used to generate an autologous EBV cell line which is a transfection target for immunological studies. In addition, following each immunization, 60 ml of peripheral blood is drawn, FICOLL-HYPAQUE® separated and cryopreserved for the determination of T cell responses. The T cell responses to be studied are generation of specific cytotoxic T cells, cytokine producing T cells, and proliferation of the T cell cultures in response to the antigens. When available, cutaneous tumor biopsies are obtained from the patients to provide a source of tumor infiltrating lymphocytes (TIL). Similar studies are run using TIL to determine if tumor biopsies become a source of GD2 specific cells. Also, tumor biopsies provide a source of tumor
cells to serve as critical autologous targets for cytotoxicity assays, cytokine production, and proliferation assays.

In-Vitro Functional activity of T cells is measured as follows: FICOLL/HYPAQUE-separated PBMC (1-3x10^6) are incubated in the presence of medium alone, IL-2 (10 Cetus units/ml), or 0.1 to 100 μg/ml anti-id 1A7 antibody. The cell culture medium includes isovolos medium supplemented with 10% human AB serum, gentamycin, sodium pyruvate, non-essential amino acids, L-glutamine and 10 Cetus units/ml recombinant IL-2. Every 7 days the cultures are stimulated with irradiated autologous PBL pre-sensitized with the appropriate antigen used at day 0. The method of in vitro sensitization is similar to that recently described by Rivolitti et al., (1995) J. Immunol. 154:2225-226. Beginning day 21 and on a weekly basis, proliferating cells are assessed for cell surface phenotype and cytotoxic and cytokine producing potential. Initially, all T cells are tested for their ability to recognize and lyse autologous and allogeneic EBV cells transfected with the cDNA containing the sequence for the 1A7 anti-id molecule. Cultures lysing 1A7 transfected autologous EBV cells>10% are further tested against the NK sensitive line K562, the LAK sensitive line Daudi, autologous tumor if available and other HLA matched and mismatched GD2 bearing melanoma tumor cells. Preferably, a panel of over 20 well characterized melanoma tumor cell lines each expressing both class I and class II antigens is used. In addition, GM-CSF assays are run to determine if there is specific release of cytokines in addition to or in place of specific cytotoxicity. Proliferation of the cultures to the agents is determined by increases in cell numbers following in vitro stimulations.

The possible outcome of these studies following up to 6 rounds of in vitro stimulation is a kinetic increase in T cell cytotoxicity against both 1A7 transfected EBV cells, and autologous tumor and/or HLA matched allogeneic melanoma tumor cell lines. This indicates a successful immunization of the patients against their own tumor cells using the anti-id 1A7 molecule. Studies are then done to determine if the antigen recognized is GD2 on the tumor cells and identify the possible mechanisms of recognition.

Obectives of this study include: (1) determination of an optimal dose to elicit an immune response against GD2 in the various arms of the immune system; a T cell response being particularly desirable; (2) ideally, remission or palliation of the cancer.

Results
Five patients have been participating in the study over a sufficient period to provide confirmation of an immunological response. Each patient was immunized with 1 mg, 2 mg, 4 mg, or 8 mg of antibody 1A7 in QS-21 on a biweekly schedule. The first 2-4 doses were given intramuscularly, and periodic serum samples were collected to determine the presence of human anti-mouse (HAMA) activity and anti-1A7 activity. Titers were low, and it was decided to continue the course of immunization subcutaneously. All patients seroconverted positive with respect to both HAMA and anti-1A7, as determined by immunoassay. The response comprised specific Ab3 activity, as demonstrated by the ability of each serum to inhibit the binding of radiolabeled 1A7 to solid-phase linked 14G2a (Ab1).

To investigate the nature of the response further, anti-1A7 antibody was affinity purified from the sera of 4 of the patients. First, each sample was passed over a column of 14G2a antibody, eluted with a glycine buffer (pH 2.5), and exchanged into PBS. Next, HAMA activity that was not Ab3 was depleted by negative selection on a mouse immunoglobulin adsorbant. The amount of specific anti-1A7 (Ab3) obtained was as follows: Patient 1 (administered 1 mg 1A7 per dose), yield 0.67 mg Ab3 from 10 ml serum. Patient 2 (administered 2 mg 1A7 per dose), yield 1.32 mg Ab3 from 10 ml serum. Patient 3 (administered 4 mg 1A7 per dose), yield 1.71 mg Ab3 from 10 ml serum. Patient 4 (administered 4 mg 1A7 per dose), yield 0.73 mg Ab3 from 10 ml serum. This indicates that a substantial amount of Ab3 is produced as a result of administering 1A7 at any of the doses tested, and apparently is in molar excess of antigen in the circulation.

The affinity and specificity of the response to GD2 was further confirmed by using the affinity purified Ab3 in several of the assay systems described earlier. Results are shown in FIGS. 12 and 13.

FIG. 12 shows the results of ELISA conducted on Ab3 affinity purified from three different patients. In the upper panel an assay plate has been coated with ganglioside GD2 (hatched bars) or GD3 (solid bars), overlaid with purified Ab3, and then developed with alkaline phosphatase labeled anti-immunoglobulin. The results show that each patient’s response comprises the production of anti-GD2 antibody (Ab1). In the lower panel, the plate was coated with GD2, overlaid with purified Ab3, and then developed with isotype-specific anti-immunoglobulin reagents. The anti-GD2 response is apparently a mature response comprising more IgG (hatched bars) than IgM (solid bars).

FIG. 13 shows the results of inhibition titration experiments conducted using purified Ab3 from three different patients. In the upper panel, an assay plate was coated with ganglioside GD2, and varying amounts of purified Ab3 were tested for the ability to inhibit the binding of radiolabeled 14G2a (Ab1). Diamonds: Patient 1; Squares: Patient 2; Triangles, Patient 3. The half-titration point was comparable to that of unlabeled 14G2a (Circles). In the lower panel, varying amounts of purified Ab3 were tested for their ability to inhibit the binding of radiolabeled 14G2a to the GD2-expressing murine lymphoma cell line EL4. The results indicate that the Ab3 induced by administration of 1A7 competes for binding to GD2 both in plate-binding assays and when presented on cancer cells.

Additional patients are enrolled in the study to characterize the immune response elicited by administration of antibody 1A7, and to follow any effect on the melanoma.

At the completion of the study, a second study is designed using the optimum immunomodulatory dose. The total number of patients in the second study is chosen for statistical reasons and is between about 16 to 25.

Example 5

Clinical use of 1A7 Antibody Vaccines in the Adjuvant Setting

The objectives of this study comprise ascertaining the effects of 1A7 in patients who have been treated for a GD2-associated cancer and have no clinical manifestations of the disease. Ideally, 1A7 given at an optimal dose lessens the risk or rate of recurrence.

Eligible patients are those with GD2-positive small cell lung cancer. All of the patients must have entered a complete clinical remission following standard chemotherapy, and be within 6 weeks of completion of chemotherapy and radiation therapy. Patients must have a life expectancy greater than six months, adequate nutrition, non-pregnant. Southwest Oncology Group performance score 0, no history of monoclonal antibody therapy, no ongoing use of nonsteroidal anti-inflamm-
Various dose levels of 1A7 are combined with a suitable adjuvant. One candidate is QS-21. The QS-21 molecule consists of a triterpene glycoside with the general structure of a quilliac acid 3,28-O-bis glycoside. It consists of two structural isomers designated V-1 and V-2 at a typical ratio of about 2:1. Both isomers have adjuvant activity. QS-21 has been shown to promote a response to both T-dependent antigens and unconjugated T-independent antigens. QS-21 also augments the induction of Class I MHC-restricted cytotoxic T lymphocytes as well as antigen specific T-cell proliferation when used in subunit antigen vaccines. 100 µg QS-21 has been established as the optimal amount of adjuvant per dose in a trial conducted at the Memorial Sloan-Kettering Cancer Center, NY. The other candidate is RIBITMPC, which is obtained from Ribi Immunochem Research Inc., Hamilton Mont., and used according to manufacturer’s directions.

Patients receive one intramuscular injection of 1A7 every two weeks for a total of four injections. Patients are immunized with either 1 mg, 2 mg, or 4 mg of 1A7 per injection mixed with 100 µg QS-21. Therapy continues on a monthly basis for about 24 months, then every 3 months until tumor progression is detected. The total number of patients is between 9 and 24.

Blood samples are obtained monthly prior to each treatment. Serum levels of Ab3 (anti-1A7), Ab1’ (anti-Gd2) and human anti-mouse antibody (HAMA) are measured by standard immunoassay. The specificity of these responses is confirmed by indirect immunoprecipitation of radiolabeled cells and SDS-PAGE. Sera is also tested to inhibit the binding of labeled 1A7 to M21/P6 cells or purified Gd2.

To determine if the vaccine is inducing T-cell immunity, the huffy coat of one unit of blood is centrifuged on FICOLL-HYPAQUE®. The peripheral blood mononuclear cells (PBMC) are removed, washed, and the lymphocyte precursor frequency is determined. Immunostaining for CD3, CD28, and CD45R markers is used to measure and sort cytotoxic T cells from suppressor T cells, using three-color flow cytometry. The proliferative response to 1A7 antibody and purified tumor antigen is determined. Cytotoxicity assays are conducted using HLA-matched colon cancer cell lines or autologous tumor cells. Suppressor cell function of CD8+ CD28+ CD45R+ cells is measured as the suppression of β cell immunoglobulin secretion.

The clinical features of each recipient are monitored regularly, and compared with the control population to determine the efficacy of treatment. The dose and frequency of 1A7 treatment is adjusted if necessary as more patients enter the study. A dose of 1A7 is identified that significantly increases the risk or rate of progression compared with controls.

**Example 6**

**Construction of a Recombinant Vaccinia Vector Encoding a 1A7 Polypeptide Fragment**

Recombinant vaccinia virus (rVV) can provide a powerful agent for overcoming immune tolerance. Vaccinia virus is highly immunogenic and stimulates both humoral and cell-mediated immune responses. Vaccinia is also capable of representing TAA along with the cellular MHC complex.

This example describes plasmid construction and production of recombinant vaccinia viruses. The scheme for construction of a general vaccinia vector (rVV) is shown in FIG. 14. The complete sequence of TK gene of the wild type WR strain of vaccinia virus (GenBank, accession number J02425) was obtained from the National Center for Biotechnology Information (NCBI) by the BLAST program. Aitschul et al. (1990) J. Mol. Biol. 215:403-410. From the sequence data, forward and reverse PCR primers 5'-GAATGGAAAGGGCCCAAC (SEQ. ID NO:52) and 5'-GATGACGATCATATTAC (SEQ. ID NO:53) were synthesized, corresponding to nucleotides 22-39 and 727-708 respectively of the TK sequence (Hruby et al. (1983) Proc. Natl. Acad. Sci. USA 80:3411-3415. An Apa 1 site (underlined) was introduced into the forward primer and a Nsi I site (underlined) in the reverse primer for insertion into the plasmid pGEM-7Zf+ (Promega). DNA from the wild type WR strain of vaccinia was isolated and TK gene was amplified by PCR. A DNA fragment of expected size (about 700 bp) was obtained by PCR. This DNA was separated by electrophoresis in low melting point agarose and purified by digestion with Gelase (Epigenetix Tech.). The TK DNA fragment was ligated to the pGEM-7zf+ (+) after digestion with Apa I plus Nsi I. The resulting plasmid (pGEM-1K) was amplified by standard transformation techniques. Insertion was verified by restriction mapping.

Promoter 7.5 K was amplified from wild type vaccinia virus by PCR using the forward primer 5'-GTTATCGATG-TGCGAATAGCC (SEQ. ID NO:54) and the reverse primer 5'-TGTCGTGCAGATGACTGTCTT (SEQ. ID NO:55), corresponding to nucleotides 69-88 and 335-312 of the 7.5 K promoter sequence. Cochran et al. (1985) J. Virol. 54:30-37. A Cla I site (forward) and a Pst I site (reverse) were included in the primers. The amplified DNA fragment was digested with Pst I. A polynucleotide adaptor was synthesized with the smaller oligonucleotide being phosphorylated at the 5'-end by polynucleotide kinase. The hemi-phosphorylated adaptor was ligated to Pst I digested PCR amplified 7.5 K promoter DNA fragment. The product was digested with Cla I/EcoRI digested pGEM-TK.

A cDNA insert encoding a 1A7 polypeptide is inserted between the Neo I and Xma I (Sma I) sites of pVV. This plasmid also contains the leader sequence of the Vp5 at the 5' end of the svp CDNA. If desired, a vaccinia control plasmid can be constructed containing cDNA for E. Coli β-galactosidase.

**Construction of rVV**

Recombinant vaccinia virus (rVV) can be constructed by homologous recombination of vaccinia plasmids and wild-type WR strain of vaccinia virus according to the procedure of Mackett et al. (DNA Cloning, Vol. II, D. M. Glover, ed., IRL Press 1985) using CV-1 cells. Recombinant viral clones expressing β-galactosidase are selected by growth on TK-143B cells in the presence of 5-bromo-4-chloro-3-indolyl-β-D-galactosidase (X-Gal). Blue recombinant viruses are picked by Pasteur pipettes and plaque purified. As a second step in clone selection, Southern blot of extracted DNA is performed, using 1A7 cDNA as the probe. Further selection of rVV is made by assay of culture supernatant of the virally infected CV-1 or any other eukaryotic cells by ELISA. If cell-associated 1A7 polypeptide is in the rVV (i.e., if the leader sequence is deleted), cell lysate is assayed. Western blotting with 14G2a as probe is also performed. Biological activity of the 1A7 polypeptide synthesized by the vaccinia virus is determined by cell binding inhibition assay, as described above. RVV clones containing 1A7 polynucleotides are selected by staining with 0.1% neutral red and plaque purified as above. Viral clones are grown into a high-titer lysate using standard techniques (Mackett et al. (1982) Proc. Natl. Acad. Sci. USA 79:7415-7419). Typically a clone producing the highest amount of 1A7-polypeptide is selected for further studies.
Assay of 1A7 Polypeptides (Foreign Proteins) Expressed By Recombinant Vaccinia Virus: CV-1 cells are propagated in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% fetal calf serum and 100 units of penicillin and 100 μg of streptomycin per ml in 25-cm² flasks or Swell Cluster flasks. Cells are inoculated with rrv at a MOI of 30. The virus is allowed to adsorb for 2 hours at 37°C in a tissue culture incubator, following which the inoculum is replaced with the culture medium and the incubation was continued. Supernatant is removed after incubation for indicated time and the 1A7 polypeptide secreted is assayed. As a control, supernatant from mock infected cells is used. Assay of 1A7 polypeptides can be performed by testing for binding to 14G2a as described elsewhere in this disclosure. β-D-galactopyranoside produced by rrv-lacZ is assayed according to Miller (Experiments in Molecular Genetics, Cold Spring Harbor Press 1972) with p-nitro-β-D-galactopyranoside as the substrate. Culture supernatant from virus infected cells is treated with β-proprionate to inactivate the virus before assay (Corcoran et al. (1988) J. Parasit 74:763). Incorporation of 3H-thymidine by NFS60 cells is used as a measure of cell proliferation (Jaffe et al. (1993) Cancer Res. 53:2221-2226). Radioactivity due to 3H-thymidine incorporation in the presence of supernatant from mock infected CV-1 cells is subtracted as background. As positive control and for standard of biological activity, intact 1A7 is used. Alternatively, standard solutions of GM-CSF can be used as described in Qin and Chatterjee (1996) Gene Therapy.

Testing vaccinia 1A7 vaccines: For administration of vaccinia, a virus titer of 10⁶ to 10⁸ pfu is injected in a mouse. Injections can be subcutaneous, intramuscular, intradermal or intraperitoneal. Immunizations are performed weekly. Mice are bled 7 days after every immunization for determination of Ab3 (including Ab1). Testing for development of T cell immunity is performed 10 days after the booster immunization. Mice can also be tested by tumor challenge, in which survival after injection with tumor cells is monitored.

For administration of vaccinia via virally infected tumor cells, autologous tumor cells are maintained in Eagles medium containing 10% vol/vol fetal calf serum, 2 mM glutamine and gentamicin. A monolayer of confluent cells in a 75-cm² flask is inoculated with (3x10⁵) plaque forming units (pfu) of rrv. After 2 hours at 37°C, the inoculum is replaced by DMEM and the incubation was continued for another 24 hours. After examination under microscope, cells are collected by scraping and washed two times with PBS and resuspended in PBS at a desired concentration. (10⁶ to 10⁷/200 μl). Female C57BL/6 mice, 6-8 weeks old are purchased from Hurlan Bioproducts for Science Inc., (IN). Animals are injected subcutaneously with the cellular vaccine in the rear left flank and two weeks later tumor cells are injected at the rear right flank for challenge. Survival of mice following tumor challenge and the presence of tumor is monitored daily. If the tumor is measurable, tumors are measured weekly by a caliper in two dimensions and the volumes are calculated using the formula (width x length)/2. Tumors which are palpable but too small for measuring the volume accurately are recorded as zero volume, but tumor incidence is recorded as positive. Tumor volumes are averaged only for those that actually develop tumors over the observation period of 120 days. Zero values are included for those mice that eventually develop tumors but were tumor-free at a given time point.

Statistical evaluation is done using SigmaStat software (Jandel, Inc. San Rafael, Calif., USA), A P value of <0.05 is considered to indicate statistical significance.

Example 7

Expression and Characterization of a 1A7 Single Chain Variable Region Molecule

Based on the sequence data listed elsewhere in this application, a CDNA construct is prepared by standard recombinant cloning techniques for single chain 1A7 derivatives in which V₃H and V₃L are joined through a linker sequence. The strategy is as follows. The construct is incorporated into the pET-22b(+) plasmid vector (Novagen, Madison, Wis.) and expressed in E. coli. Sequence analysis is performed to confirm the plasmid construct, which contains the carboxy-end of V₃H linked to the framework of V₃L and not the leader region. pET-22b(+) contains a nickel ion binding domain consisting of 6 sequential histidine residues (His₆). The His₆ domain has a high affinity for nickel, which is used for the purification of the recombinant 1A7 scFv.

A cell binding competition assay is performed to investigate whether the 1A7 scFv retains the antigen mimicry shown by intact 1A7. GD2-positive M21/P6 cells (1x10⁶ cells/well in 50 μl volume) are placed in a 96-well plate. The cells are incubated for 2 hours at room temperature with [125I] 14G2a (Ab1), 100,000 cpm, in the absence and presence of increasing concentrations of 1A7 or the 1A7 scFv fragment. Percent inhibition is calculated according to standard formula.

The strategy has been implemented to prepare genetic constructs encoding scFv of the form V₃H-(GGGGS)₉-V₃L, and V₁L-(GGGGS)₉-V₃H. T-vector derived plasmids containing the encoding sequence for the 1A7 heavy chain variable region (pTL12A7H1) and for the 1A7 light chain variable region (pTL12A7L1) were obtained as described in Example 2. The plasmids were linearized by digestion with Avai. Using the linearized plasmids as template, PCR amplification was conducted using a special set of primers designed to incorporate the following features:

1. A unique restriction nucleic site for subsequent ligation of the construct into a vector plasmid. An EcoRV site was used.
2. A Kozak’s consensus sequence for ribosome binding operably linked at the 5’ end to a start codon and the encoding region for the signal peptide of the first variable region;
3. A (GGGGS)₉ encoding sequence linked directly at the 3’ end of the coding sequence of the first variable region (Framework 4);
4. A (GGGGS)₉ encoding sequence linked directly at the 5’ end of the coding sequence of the second variable region without the signal peptide (Framework 1);
5. A stop codon linked directly at the 5’ end of the coding sequence of the second variable region (Framework 4).
6. A second unique restriction nucleic site link. Xbal was used. For the construct V₁L-(GGGGS)₉-V₃L, V₁H was the first variable region and V₃L was the second; whereas for V₃H-(GGGGS)₉-V₃L, V₃H was the first variable region and V₃L was the second.

Using these primers in the initial PCR incorporates the translation and linker sequences during amplification. A region of overlap is present through the linker, which then permits the heavy and light chain sequences to be amplified together in a second round of PCR using the outermost primers.
The primers were synthesized by standard techniques by the Macromolecular Structure Analysis Facility at the University of Kentucky. The sequences are shown in Table 7. The abbreviation Lk stands for the linker (GGGGS)₃ (SEQ. ID NO:45).

**TABLE 7**

<table>
<thead>
<tr>
<th>Designation Construct</th>
<th>Sequence Used to amp- Orient-ify (5'→3')</th>
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<tbody>
<tr>
<td>P1-VH</td>
<td>V₄₋₅(Lk) - V₅</td>
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<tr>
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<tr>
<td></td>
<td>Reverse: TTGCTGCTATGCAAAATCCGATGGCCGCCG</td>
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<tr>
<td>P3-VLL</td>
<td>V₄₋₅</td>
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<tr>
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<tr>
<td></td>
<td>Reverse: CCTCCCTTGGAGGAGACGTGACCAAA</td>
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The V₄₋₅(Lk)-V₅ construct was assembled in the following way: A PCR amplification was conducted on the linearized pT1A7VH plasmid using P1-VH and P3-VLL, and on the linearized pT1A7VL plasmid using P2-VHL and 3′-VL. The amplified products were freed of primers, annealed, and another round of PCR amplification was done using the primers P1-VH and 3′-VL. The product was a DNA fragment comprising an encoding sequence for a single-chain molecular containing in order: the V₄₋₅ signal sequence, V₅, the linker, and V₅.

FIG. 15 lists the nucleotide sequence (SEQ. ID NO:65) for the V₄₋₅(Lk)-V₅ obtained from the second round of PCR amplification. Restriction sites EcoRV and XbaI are shown by underlining. The ATG start codon is italicized and preceded by a l. The linker sequence is the long underlined section near the middle of the sequence. The termination codon TAA is italicized and followed by a l. The Kozak’s consensus sequence ACCATGG (SEQ. ID NO:64) precedes the start codon shown below the nucleotide sequence is the expected translation product (SEQ. ID NO:66) which is 265 amino acids in length.

The V₄₋₅(Lk)-V₅ construct was assembled in exactly the same fashion, except that the second set of primers listed in Table 7 were used in the PCR amplification reactions.

Each of the constructs was cloned into a pcDNA3-derived vector plasmid. pcDNA is a ~5445 base-pair circular DNA containing in order the CMV promoter, a polycloning site, and the bovine growth hormone polyadenylation signal (BGHpA), and was obtained from Invitrogen. The pcDNA3 plasmid was first modified to remove unneeded sequences to improve its suitability for administration to human subjects. First, it was digested with PvuII and self-ligated. This resulted in removal of 2165 base pairs comprising the neomycin resistance gene. The product was digested with NruI and PvuII, and blunted by digestion with 3′-5′-exonuclease. This removed the ampicillin resistance gene, resulting in a linear DNA designated pcDNA 3.2. Kanamycin resistance gene was cut out from pUC4K with PstI, and blunted by 3′-5′-exonuclease digestion to make a DNA fragment designated pUC4K1.2. pUC4K1.2 and pcDNA3.2 were then ligated, resulting in the formation of a "humanized" plasmid with a kanamycin resistance gene, designated pHeiDNA3. The scFv encoding sequences were cloned into pHeiDNA3 by digesting both the plasmid and the scFv DNA fragment with EcoRV and XbaI, followed by ligation.

FIG. 16 is a map of the construct comprising an expressible gene for the V₄₋₅(Lk)-V₅ scFv. The encoding region is under control of the CMV promoter, and linked downstream to the bGHPA polyadenylation sequence.

Ability of the plasmids to express the encoded protein in a suitable conformation is tested by transfecting CHO cells and MC38 cells. TRANSFECTAMINETM or electroporation are alternatively used to effect the transfections. Both the culture supernatant and a cell lysate are tested for the presence of the gene product. A sandwich ELISA is conducted using antibody 14G2a(1b1) as both plate coat and 125I-labeled developing reagent. A parallel assay using an irrelevant antibody as developing reagent is used as negative control. Positive results are confirmed by conducting a Western blot using 125I-14G2a. The expected size of the scFv gene product is ~44 kDa.

The scFv can be isolated by affinity chromatography using a 14G2a adsorbent, eluted with a pH ~2.5 glycine buffer. Further investigation of the specificity of the scFv is conducted using the affinity-purified protein. GI22-positive M21/P6 cells (1×10⁷ cells/well in 50 uL volume) are placed in a 96-well plate. The cells are incubated for 2 hours at room temperature with 125I-14G2a (Ab1), 100,000 cpm, in the absence and presence of increasing concentrations of 1A7 or the 1A7 scFv fragment Percent inhibition is calculated according to standard formulae.

Plasmids encoding scFv with the correct specificity are assembled into candidate 1A7 vaccines using methods known in the art for assembling polynucleotide vaccines. One such candidate is a naked DNA vaccine. It is prepared by growing the plasmid to sufficient quantities, purifying it from the host cell, and mixing it with a pharmaceutically compatible buffer. Another candidate comprises cationic liposomes to facilitate transfection. Materials and methods for assembling cationic liposomes are known in the art: see U.S. Pat. Nos. 5,264,618; 5,334,761; and 5,459,127.
Candidate 1A7 polynucleotide vaccines are tested for immunogenicity as follows. Two groups of 10-15 female C57BL/6 mice (6-8 weeks old) are immunized intramuscularly with doses of 50-100 μg purified plasmid. Various routes of administration are compared, such as intramuscular, intradermal, subcutaneous and interperitoneal.

Mice are bled 7 days after each immunization for determination of anti-1A7 and anti-GD2 activity as described elsewhere in this disclosure. Three mice are sacrificed from each group for isolation of spleens for the T cell proliferation assay 10 days after a booster immunization. To determine whether effects are specific, the following negative controls are used: (a) plasmid without the 1A7-seFv polynucleotide insert; (b) plasmid with the seFv polynucleotide insert in the opposite orientation with respect to the promoter, and (c) plasmid containing a polynucleotide encoding an scFv of an unrelated anti-idiotypic antibody.

Example 8

Development of an Animal Model for Testing Vaccines Comprising a 1A7 Polynucleotide or Polypeptide Derivative

A number of different fragments, constructs, plasmids, and fusion proteins are contemplated in this invention as a second generation vaccine for GD2-associated tumors. Animals have been established in the examples given so far as suitable for testing whether a candidate vaccine can elicit an immune response. Ultimately, the object is to use the vaccine in tumor therapy, and it is desirable to have an animal model that can be used to screen candidate vaccines for this endpoint.

Accordingly, a (57BL/6) mouse EL4 lymphoma model was developed.

Cheung et al. (1993, Int. J. Cancer 54:499-505) reported that murine lymphoma EL4 cells express GD2 at high density. MAb 14G2a was tested for binding to EL4 cells. Essentially 100% of the EL4 cells (a gift from Dr. Suzanne Rosenberg, Univ. of Maryland) bound at high fluorescence intensity with 14G2a by FACS analysis. The cell binding inhibition curve showed that 1A7 can effectively inhibit the binding of 125-labeled 14G2a to EL4 cells. Immunization of C57BL/6 mice with anti-Id 1A7 plus Q8-21 induced anti-GD2 antibodies which bind to EL4 cells and kill EL4 cells in vitro ADCC assay. Also, spleen cells from immunized C57BL/6 mice showed in vitro cell proliferation in presence of irradiated EL4 cells. Thus, murine EL4 lymphoma in syngeneic immunocompetent C57BL/6 mice can serve as a suitable animal model for evaluating the efficacy of anti-Id A7-based vaccines.

100,000 EL4 cells are injected subcutaneously (s.c.) into C57BL/6 mice. This is lethal to the host, and the median survival time is approximately four weeks. Since median survival time is dependent on the inoculum of tumor cells, a dose-related survival time is generated using 10^3, 10^4, 10^5, and 10^6 tumor cells/mouse, injected s.c., with 10 mice per group. A dose is determined that develops into a slow growing or rapidly growing tumor. The subcutaneous tumor model permits us to more closely monitor tumor growth and host immune response without slicing the mouse. EL4 cells also can be injected i.p. which grow very rapidly with intra-abdominal metastasis. The intraperitoneal model can be adapted to study the immune response to tumors which develop systemically.

Five different forms of vaccines are explored and compared for efficacy. The vaccines are:

(i) 1A7-QS21 (as our standard control)
(ii) Irradiated EL4 cells (Positive Control)
(iii) GD2-KLH plus QS-21 (Antigen Vaccine)
(iv) 1A7-plasmid
(v) 1A7-Vaccinia Construct

The serum levels of anti-anti-Id (Ab3) and anti-GD2 antibodies are measured as described elsewhere in this disclosure. Typically, blood samples are obtained before vaccination and ten days after each immunization and assayed for anti-GD2 antibodies. The time course is determined over which the immune response develops, the intensity of the immune response, the effect of multiple injections of vaccine (booster), duration of the humoral response and variability of the humoral response between animals. Comparing the anti-GD2 titers with survival of tumor challenge establishes whether there is any correlation between the level of humoral response and tumor protection. The specificity of the immune response is examined by ELISA, RIA, Dot Blot analysis and FACS analysis. The serum Ab3 is also studied by in vitro ADCC or CMC assays. The target cells are EL4. The isotypes of anti-GD2 antibodies in the serum of mice are determined by ELISA using isotype specific reagents.

The cellular immune response is assayed using a T cell proliferation assay and a cytotoxic T lymphocyte (CTL) assay. The proliferation assay is used to assess the proliferative response of cells from vaccinated mice to various stimuli. Spleen cells are harvested from animals immunized with an experimental vaccine or control vaccine and placed into vitro cultures. The splenocytes are then stimulated with either media alone, phytohemagglutinin (PHA), irradiated EMA cells, purified GD2, anti-Id 1A7 or an irrelevant Ab2. Cell proliferation is measured after 5 days of culture and then stimulation for 18 hours by [3H]-thymidine incorporation. Stimulation of the spleen cells with PHA shows the maximum proliferation of T cells, and culturing with media alone measures baseline proliferation. Splenocyte cultures are of mixed cell type, including antigen presenting cells. Pulsing cells with the above stimulants results in antigen processing by these cells and a reactive T lymphocyte proliferation. By comparing the proliferative response of the differently vaccinated mice, it is possible to determine if there is an advantage of any particular vaccine form over others.

In order to determine which subset of T-cells are being induced, the proliferating cells are phenotyped by flow cytometry. Alternatively, splenic T-cells are depleted of either CD4+ or CD8+ cells before proliferation assay by incubation with monoclonal antibody RL172 (anti-CD4+) or mAb168 (anti-CD8+) and complement.

One of the effector mechanism thought to be important for tumor protection is antigen specific CTL killing. EL4 or GD2 specific CTL activity will be assayed to determine if the vaccines induce this type of cellular response. Splenocytes are harvested from vaccinated mice and co-cultured with irradiated EL4 tumor cells and IL-2 for five days to expand Ag specific CTL. The splenocytes are then mixed with 51Cr-labeled target cells at various effector to target ratios, and then assayed for released radioactivity. The target cells are EL4 cells and MC38 cells. MC38 is a murine-colonic tumor cell line which was derived from the same mouse strain and thus shares the same H-2 haplotype but has different surface antigen makers. Both of these tumor cell lines take up 51Cr and have had spontaneous release. Vaccination of mice should stimulate the production of CTL which specifically lyse EL4 cells and not MC38 cells.

By comparing the CTL activity in differently vaccinated mice, the particular type of vaccine stimulates different levels of CTL response is determined. Also determined is whether
different adjuvants or cytokines are needed to influence this type of immune response generated.

**Example 9**

Evaluating the Efficacy of Derivative Vaccines

Efficacy of the different vaccinations is measured by the degree of tumor protection caused by vaccines using the animal model outlined in the previous example. Tumor protection is compared with other measurements of the immune response to determine if there is any correlation between tumor protection and level of humoral or cellular response.

To evaluate the ability of vaccinations to prevent tumor growth, vaccinated mice are challenged with a lethal number of tumor cells. The immunization schedule is determined by evaluating the humoral and cellular immune response as described above. It is likely that the schedule and number of immunizations necessary to induce a significant immune response varies between the different types of vaccines. Fifteen mice are used in each vaccination group including one group of mice immunized with an irrelevant Ab2. Blood samples are obtained after 7 days of each immunization to monitor the humoral response. Once immunized, 10 of the mice are challenged with a previously established lethal number of EL4 cells injected s.c. The survival of the different vaccination groups is compared using Kaplan-Meier analysis. The other five mice in each vaccination group are used for CTL assays. These studies give information about the immune status at the time of tumor challenge and protection from tumor growth. Different types of vaccines are also compared (anti-id protein, cells, GI2-KLH or DNA) for their ability to stimulate a protective immune response.

The level of tumor protection is evaluated by the number of mice that survive a tumor challenge and the dose of tumor cells immunized mice can survive. The level of tumor protection is measured for each of the vaccines which stimulate tumor protecting immune responses.

Specificity of tumor protection is evaluated using an unrelated tumor cell such as MC38 which grows subcutaneously in C57BL/6 mice. Groups of mice are vaccinated with different vaccines and then challenged with either EMA or MC38 cells. The mice are observed daily and survival comparison between two groups of mice is made using Kaplan-Meier analysis.

To evaluate the effectiveness of vaccine therapy in the treatment of mice with established tumors, C57BL/6 mice are injected with appropriate number of EL4 cells subcutaneously and then 24 hours later immunized with different vaccines or unrelated Ab2 vaccine. The mice are reimmunized every week and followed for tumor growth and survival.

Experiments are conducted to determine the immune effector arm, involved in protective immunity against syngeneic GI2 antigen bearing tumors. Adoptive transfer of immune Ab3 serum (containing Ab1') or immune T-lymphocyte subsets (CD4+ or CD8+) or NK cells is done into naive recipient C57BL/6 mice previously treated with sublethal dose of radiation to allow for cell expansion. The recipient mice and previously vaccinated immunized positive control mice are then injected with a lethal number of tumor cells and followed for tumor growth and survival.

**Example 10**

Clinical Use of Derivative Vaccines

Promising results using plasmids in the animal tumor model of the previous example lead to development of plasmid vectors for human use. The pcDNA3 vector is modified according to Conry et al. for incorporation of the appropriate Ab2β fragment. This modification involves deletion of the neomycin gene and replacing it with Tn903 kanamycin resistance gene. Ampicillin resistance gene and viral sequences corresponding to nucleotide 5004–208 are also deleted. The plasmid is prepared on a large scale under Good Manufacturing Practice (GMP) conditions. Purified plasmid is stored at −70°C in aliquots of 5 mg/ml.

Vaccinia constructs are made in Wyeth-calf adapted strain of vaccinia. This strain of vaccinia has been used for smallpox vaccination and has been shown to be safe in cancer patients. Clones of rVV are picked and plaque purified. Rvv are grown under GMP conditions using certified strains of eukaryotic cells. Virus is concentrated by sucrose gradient centrifugation.

An initial study is done in six advanced melanoma patients. Depending on results, anti-id based DNA vaccines is combined with the standard control A17 plus QS-21 vaccines for augmenting tumor specific immunity in vaccinated patients.

**ADDITIONAL REFERENCES**

Gangliosides as Tumor Antigens


Plasmid and Vaccinia Vaccines


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(D) TOPOLOGY: linear

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Ser Val Ser Ile Tyr Tyr Gly Arg Ser Asp Lys
1 5 10

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:
Xaa Xaa Xaa Xaa Xaa Xaa
1 5

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:
Xaa Xaa Xaa Xaa
(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 12:
Arg Asp Tyr Arg

(2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 5 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 13:
Gly Tyr Tyr Asp Xaa

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 5 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 14:
Xaa Xaa Xaa Xaa Xaa

(2) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 112 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 15:
Amp Val Leu Met Thr Gln Thr Pro Leu Ser Leu Pro Val Ser Leu Gly
1  5  10  15
Amp Gln Ala Ser Ile Ser Cys Arg Ser Ser Gln Ser Ile Val His Ser
20  25  30
Aam Gly Asn Thr Tyr Leu Glu Trp Tyr Leu Gln Lye Gly Gln Ser
35  40  45
Pro Lys Leu Leu Ile Tyr Phe Val Ser Asn Arg Phe Ser Gly Val Pro
50  55  60
Amp Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile
65  70  75  80
Ser Arg Val Glu Ala Glu Asp Leu Gly Val Tyr Cys Phe Gln Gly
85  90  95
Ser His Val Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lye
100 105 110
(2) INFORMATION FOR SEQ ID NO: 16:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 119 amino acids
(B) TYPE: amino acid
(C) STRANDINESS: single
(D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

Gln Val Gln Leu Lys Glu Ser Gly Pro Gly Leu Val Ala Pro Ser Gln
1      5      10      15
Ser Leu Ser Ile Thr Cys Thr Val Ser Gly Phe Ser Leu Thr Ser Tyr
20     25     30
Gly Val His Trp Val Arg Gln Pro Pro gly Lys Gly Leu Glu Trp Leu
35     40     45
Gly Val Ile Trp Gly Asp Gly Ser Thr Asn Tyr Asn Ser Ala Leu Lys
50     55     60
Ser Arg Leu Ser Ser Lys Asp Ser Ser Lys Ser Gln Val Phe Leu
65     70     75     80
Lys Met Asn Ser Leu Gln Thr Asp Thr Ala Arg Tyr Tyr Cys Ala
85     90     95
Arg Glu Xaa Xaa Xaa Xaa Tyr Tyr Ala Met Asp Tyr Trp Gly Gln Gly
100    105    110
Thr Ser Val Thr Val Ser Ser
115

(2) INFORMATION FOR SEQ ID NO: 17:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 336 base pairs
(B) TYPE: nucleic acid
(C) STRANDINESS: single
(D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

GATGTTTTCA TGACCCCAA TCACACTTCCT CTGCTGTTCTCA GCTTGGAGAG TCAGCCTCCC  60
ATCTCTGAGA GATGATGCTA CGACATTGTA CAGATGAA TGAACACCTA TTTAGATGGG 120
TACCTGAGA AACCAGCCA GTCTCCGAAG CTCTGGATCT ACAAGTCTTC CAACCCATT  180
TCTGAGGCTC CAGACAGCTT CATGCGAGT GATCGGGAGA CAGATTTCAC ACTCAAGATC 240
AGCGAGAATG AGCGGGAGAGG TCTGAGGTTTT TATTACTGCT TTCAAGGCTC ACGUGTCCCG 300
TGAGATCAGC GTGGAGGSCAC CAAGCTGGAA ATCAAA  336

(2) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 336 base pairs
(B) TYPE: nucleic acid
(C) STRANDINESS: single
(D) TOPOLOGY: linear

(ii) MOLECULAR TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

GATGTTTTCA TGACCCCAA TCACACTTCCT CTGCTGTTCTCA GCTTGGAGAG TCAGCCTCCC  60
ATCTCTGAGA GATGATGCTA CGACATTGTA CAGATGAA TGAACACCTA TTTAGATGGG 120
TACCTGAGA AACCAGCCA GTCTCCGAAG CTCTGGATCT ACAAGTCTTC CAACCCATT  180
TCTGAGGCTC CAGACAGCTT CATGCGAGT GATCGGGAGA CAGATTTCAC ACTCAAGATC 240
AGCGAGAATG AGCGGGAGAGG TCTGAGGTTTT TATTACTGCT TTCAAGGCTC ACGUGTCCCG 300
TGAGATCAGC GTGGAGGSCAC CAAGCTGGAA ATCAAA  336
TCTGCGGTCC CAGCACGACT CAGTGGGCTG GATTCAGGGA CAGATTCAC ACTCAAGATC 240
AGCACGAGTG AGCCTGAGGA TCTGGGAGTT TATTTACGCT TCTCAAGGTC ACATGTTCCG 300
TGACAGCGTC GTCGGAGGAC CAGGCTGCAA ATCACA 336

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 336 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) SEQUENCE DESCRIPTION: SEQ ID NO: 19:
GATGTTTCT GACCCCAAC TCCACCTCC CTCCCACTCA GTCCTGGACA TCAAGCCTCC 60
ATCTCTCTCA GACTCTGCTA GAGCATTGTA CATAGTAATG GAAACACCTA TTTAGAATGG 120
TACCTGCGAA AACGAGCACA GTCTCCAAAG CTCTCTATCT ACTAAAGTTT CAAAGCAGTT 180
TCTGGGGTCC CAGGCGGCT GATGGGCTG GATGCAGGGA CAGATTCAC ACTCAAGATC 240
AGCACGAGTG AGCCTGAGGA TCTGGGAGTT TATTTACGCT TCTCAAGGTC ACATGTTCCG 300
TGACAGCGTC GTCGGAGGAC CAGGCTGCAA ATCACA 336

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 333 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) SEQUENCE DESCRIPTION: SEQ ID NO: 20:
GATGTTTCTA TGACCCCAAC TCCACCTCC CTCCCACTCA GTCCTGGACA TCAAGCCTCC 60
ATCTCTCTCA GACTCTGCTA GAGCATTGTA CATAGTAATG GAAACACCTA TTTAGAATGG 120
TACCTGCGAA AACGAGCACA GTCTCCAAAG CTCTCTATCT ACTAAAGTTT CAAAGCAGTT 180
TCTGGGGTCC CAGGCGGCT GATGGGCTG GATGCAGGGA CAGATTCAC ACTCAAGATC 240
AGCACGAGTG AGCCTGAGGA TCTGGGAGTT TATTTACGCT TCTCAAGGTC ACATGTTCCG 300
TGACAGCGTC GTCGGAGGAC CAGGCTGCAA ATC 333

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 336 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) SEQUENCE DESCRIPTION: SEQ ID NO: 21:
GATGTTTCTA TGACCCCAAC TCCACCTCC CTCCCACTCA GTCCTGGACA TCAAGCCTCC 60
ATCTCTCTCA GACTCTGCTA GAGCATTGTA CATAGTAATG GAAACACCTA TTTAGAATGG 120
TACCTGCGAA AACGAGCACA GTCTCCAAAG CTCTCTATCT ACTAAAGTTT CAAAGCAGTT 180
TCTGGGGTCC CAGGCGGCT GATGGGCTG GATGCAGGGA CAGATTCAC ACTCAAGATC 240
AGCACGAGTG AGCCTGAGGA TCTGGGAGTT TATTTACGCT TCTCAAGGTC ACATGTTCCG 300
(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 336 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

CGGACGTTTC GTGGAGGCAC CAAGCTGAAA ATCAA

336

(2) INFORMATION FOR SEQ ID NO: 23:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 336 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

GATTTTTGA TGACCAAAAC TCCACTCTCC CTGCCGCTCA GTCTGGAGA TCAAGCCCTCC 60
ACTCTTGCA GATCTAGTCGA GAGCATGTA CATAGTAAG GAAACACCTA TTTAGATGG 120
TACTGGCAGA AACCCGAGCA GTCTCCAAAG CTCCGATCT ACAAGTTTGC CAAGCTGAAA 180
TGGACGTTTC GTGGAGGCAC CAAGCTGAAA ATCAA

336

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 333 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

GATTTTTGA TGACCAAAAC TCCACTCTCC CTGCCGCTCA GTCTGGAGA TCAAGCCCTCC 60
ACTCTTGCA GATCTAGTCGA GAGCATGTA CATAGTAAG GAAACACCTA TTTAGATGG 120
TACTGGCAGA AACCCGAGCA GTCTCCAAAG CTCCGATCT ACAAGTTTGC CAAGCTGAAA 180
TGGACGTTTC GTGGAGGCAC CAAGCTGAAA ATC

333
(2) INFORMATION FOR SEQ ID NO: 25:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 336 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) SEQUENCE DESCRIPTION: SEQ ID NO: 25:

GATGTTTGYA TGACCCAAAC TCCACTCTCC CTGCTGGTCGA GCTTGGAGA TCAAGCCTCC 60
ATCTCTGGCA GATCATGCA GAGCATTGA CATAAGTAGT GAAACACCTA TTTAGATGG 120
TACCTGCGA AAACAGCCCA GTCTCCAAAG GCTCTGATCT ACAAAGTTTC CAACCGATT 180
TCTGCGGTCC CAGACAGATG CAGTGCGATG GAGTACGGAA CAAGATTTCC ACTCAAGATC 240
AGCAGATGGG AGCTGCGAGA TCTGGGAGTT TTACTCTGT TTTCAAGGCT TACGTTCCG 300
TGAGATGTCG GTGAGGCAC CAGGCTGGA AAATCAA 336

(2) INFORMATION FOR SEQ ID NO: 26:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 336 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) SEQUENCE DESCRIPTION: SEQ ID NO: 26:

GATGTTTGYA TGACCCAAAC TCCACTCTCC CTGCTGGTCGA GCTTGGAGA TCAAGCCTCC 60
ATCTCTGGCA GATCATGCA GAGCATTGA CATAAGTAGT GAAACACCTA TTTAGATGG 120
TACCTGCGA AAACAGCCCA GTCTCCAAAG GCTCTGATCT ACAAAGTTTC CAACCGATT 180
TCTGCGGTCC CAGACAGATG CAGTGCGATG GAGTACGGAA CAAGATTTCC ACTCAAGATC 240
AGCAGATGGG AGCTGCGAGA TCTGGGAGTT TTACTCTGT TTTCAAGGCT TACGTTCCG 300
TGAGATGTCG GTGAGGCAC CAGGCTGGA AAATCAA 336

(2) INFORMATION FOR SEQ ID NO: 27:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 291 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(x) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

CAAGCTGCAC TGCAAGAACT CAGACCTGCC CGTGCGCGGC CTCACAGAG CCTGCTAAC 60
AAGAGCCTG TGCTCTTGCG TCTTATCCAC AAGTAGGTT TAAGCTGCG TGCCGAGCT 120
CCAGAAAGG GTCTGAGGTC GTGCTGAGTA ATATGCGGTG ACCGAAACAC AAATATTCA 180
TCAGCCTCA TATTACGACG AAGCATAACT CCAAGAGCCA AGTTTCTTA 240
AAACTGAGC GTCTGCGAAC TGATGACAGA GCCACGACT ACGTGCGCAA A 291

(2) INFORMATION FOR SEQ ID NO: 28:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:

GCTAAGGGACT AGTGGGTC TCA AGGAA CTTCA GTCCACGT CT CCTCA  

(2) INFORMATION FOR SEQ ID NO: 29:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 292 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

CAGGTGCAGC TGAAAGGAC AGGACCTGGC CTGTGGCAGC CCTCAGAAC GCTGCCATC  

ACATGCACCG TCTCAAGGGT CTCTAATACC ACGTATGGTG TACACTCTGTG TC GCCAGCTT  

60

CCAGAAAGG GCTCTAGAGT GTGGTAGTT ATATGGAGTG ATGGAAGC GAAACTATAAT  

120

TCAGCTCTCA AAATCCAAGT GAGCATCAGG AAGAACAT CT AAGAGCGCA AATTTCTCTA  

180

AAAATGAAC GTA GCCAACAGT TGAGTACACA GCATACGAT ACTGTCAGG AC  

240

292

(2) INFORMATION FOR SEQ ID NO: 30:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 57 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:

GCTGACATCT ATCTATAGA CTCTGCGGT CAGGAACCT AGTCCACGG CT CCTCA  

57

(2) INFORMATION FOR SEQ ID NO: 31:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 336 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 31:

CAGGTGCAGC TGAAAGGAC AGGACCTGGC CTGTGGCAGC CCTCAGAAC GCTGCCATC  

ACATGCACCG TCTCAAGGGT CTCTAATACC ACGTATGGTG TACACTCTGTG TC GCCAGCTT  

60

CCAGAAAGG GCTCTAGAGT GTGGTAGTT ATATGGAGTG ATGGAAGC GAAACTATAAT  

120

TCAGCTCTCA AAATCCAAGT GAGCATCAGG AAGAACAT CT AAGAGCGCA AATTTCTCTA  

180

AAAATGAAC GTA GCCAACAGT TGAGTACACA GCATACGAT ACTGTCAGG AC  

240

AAATGCAACA TGTCCCAACT TGATGACACA GCCATGTACT ACTGTCAGG AC  

300

TACAGGCC AAGGCACAC TCTCCAGGTC TCCCTCA  

336

(2) INFORMATION FOR SEQ ID NO: 32:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 304 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 32:

CAGTGCCAGC TCAAGGACTG AAGACCTGCG CTGTGGCGC CCTCTAGAGG CCGTGCCATC 60
ACTGCACTG TCTCTGGAAT TTTCTAAACC AGCTATGTTG TACACTGGGT TGCCAGGCT 120
CCGCGAGGG GCCTGGAGGTC CTTGGAAATG G ATATGGGT GTGGAAAGAC CATATATAAT 180
TGCCCTCTCA TGCCCTGACT GGGCAATGAC ACAAGGACACT CAACAGGACCA AGTTTTCTTA 240
AAAATGAACA GTCTCAGAAC TGATGCACACA GCCAGTTACT AGCTGCCCAG AGGAGCATAC 300
TACG 304

(2) INFORMATION FOR SEQ ID NO: 33:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 49 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 33:

CTACTATGCT ATGAGTACT GGGGTCAGGG AACCTCTGTC ACCGTCCTC 49

(2) INFORMATION FOR SEQ ID NO: 34:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 292 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 34:

CAGTGCCAGC TCAAGGACTG AAGACCTGCG CTGTGGCGC CCTCTAGAGG CCGTGCCATC 60
ACTGCACTG TCTCTGGAAT TTTCTAAACC AGCTATGTTG TACACTGGGT TGCCAGGCT 120
CCGCGAGGG GCCTGGAGGTC CTTGGAAATG G ATATGGGT GTGGAAAGAC CATATATAAT 180
TGCCCTCTCA TGCCCTGACT GGGCAATGAC ACAAGGACACT CAACAGGACCA AGTTTTCTTA 240
AAAATGAACA GTCTCAGAAC TGATGCACACA GCCAGTTACT AGCTGCCCAG AC 292

(2) INFORMATION FOR SEQ ID NO: 35:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 50 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 35:

ACATGCTAT GAAGTACTG GCTCAAGGAA CCTCAGTCAC NGTCCTCTCA 50

(2) INFORMATION FOR SEQ ID NO: 36:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 351 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 36:
CAGCTGCAGC TGAAGAGCTC AGACACCTTC GCCTGTGGGC CACCACACAG CCGTGCCATC 60
ACATGGACCT TCCTCTGGCTT TCATTTATCC AGATATAGTG TACACTGGGT TGCCGCGCTT 120
CAGGAAAGAG GTCTGGAGTG GCTGGAGATG ATATGAGCG GTGGGAACAC AGATATATA 180
TCAGCTCTCA AATCCGACACT GAGACATCAG AAGAAGAAT CCAAGAGCCA ATGTCCCTTA 240
AAATGAACA GTCTCAGAAG TGATGACACA GCACATGACT AGCTGGAAG AGAAGGGTAC 300
TAGGATCTG TATGACTGTA GGGTCAAGA GAACCTCGAG TCACCGTCTC C 351

(2) INFORMATION FOR SEQ ID NO: 37:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 291 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 37:
CAGCTGCAGC TGAAGAGCTC AGACACCTTC GCCTGTGGGC CACCACACAG CCGTGCCATC 60
ACTGGACCT TCCTCTGGCTT TCATTTATCC AGATATAGTG TACACTGGGT TGCCGCGCTT 120
CAGGAAAGAG GTCTGGAGTG GCTGGAGATG ATATGAGCG GTGGGAACAC AGATATATA 180
TCAGCTCTCA AATCCGACACT GAGACATCAG AAGAAGAAT CCAAGAGCCA ATGTCCCTTA 240
AAATGAACA GTCTCAGAAG TGATGACACA GCACATGACT AGCTGGAAG AGAAGGGTAC 291

(2) INFORMATION FOR SEQ ID NO: 38:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 48 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 38:
TACTACCTCA TGCACTACGGGGTCAAGAAGA ACCTCAAGTCA CCCTCTCC 48

(2) INFORMATION FOR SEQ ID NO: 39:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 292 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 39:
CAGCTGCAGC TGAAGAGCTC AGACACCTTC GCCTGTGGGC CACCACACAG CCGTGCCATC 60
ACTGGACCT TCCTCTGGCTT TCATTTATCC AGATATAGTG TACACTGGGT TGCCGCGCTT 120
CAGGAAAGAG GTCTGGAGTG GCTGGAGATG ATATGAGCG GTGGGAACAC AGATATATA 180
TCAGCTCTCA AATCCGACACT GAGACATCAG AAGAAGAAT CCAAGAGCCA ATGTCCCTTA 240
AAATGAACA GTCTCAGAAG TGATGACACA GCACATGACT AGCTGGAAG AGAAGGGTAC 292

(2) INFORMATION FOR SEQ ID NO: 40:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 51 base pairs
(B) TYPE: nucleic acid
(C) STRANDINESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 40:
TACTATGCTA TGGAATATTG GGGTCAAGGA ACCTCAAGTCA CGGTCTCTC A 51

(2) INFORMATION FOR SEQ ID NO: 41:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 288 base pairs
(B) TYPE: nucleic acid
(C) STRANDINESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 41:
CAGGTCGACG TGAAAGGAGC AAAGACCTGCG CTGGTGGCGC CTCACAGAG CCTGTTTAC 60
ACCTAGAGC TCTCGGATT TCCTATTAC ACCTAGTTG TATCATGTTG TCGCATCGT 120
CCAGGAAAGG GCTCTGAGTG GCTGGAGTA ATATGGGTG GTGAAGACAC NAATTATAAT 180
TCGCTCCCTA TCTCCAGACT GAGCCACGC AAGAGCAACT CCAAGAAGCA AGTTTTCTTA 240
AAAAAGAACA GTCTCTAACG TGGTAACCA GCCATCTACT ACTGTTCC 288

(2) INFORMATION FOR SEQ ID NO: 42:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 57 base pairs
(B) TYPE: nucleic acid
(C) STRANDINESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 42:
ACGCGGNNMT TACTATGCTA TGGAATACTG GGGTCAGGA ACCTCAAGTCA CGGTCTC 57

(2) INFORMATION FOR SEQ ID NO: 43:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 293 base pairs
(B) TYPE: nucleic acid
(C) STRANDINESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 43:
CAGGTCGACG TGAAAGGAGC AAAGACCTGCG CTGGTGGCGC CTCACAGAG CCTGTTTAC 60
ACCTAGAGC TCTCGGATT TCCTATTAC ACCTAGTTG TATCATGTTG TCGCATCGT 120
CCAGGAAAGG GCTCTGAGTG GCTGGAGTA ATATGGGTG GTGAAGACAC NAATTATAAT 180
TCGCTCCCTA TCTCCAGACT GAGCCACGC AAGAGCAACT CCAAGAAGCA AGTTTTCTTA 240
AAAAAGAACA GTCTCTAACG TGGTAACCA GCCATCTACT ACTGTTCC ATT 293

(2) INFORMATION FOR SEQ ID NO: 44:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 53 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 44:

ACGACTATGC TGTGGACTAC TGGGGTCAG GAACTCAGT CACCGTCTCC TCA 53

(2) INFORMATION FOR SEQ ID NO: 45:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 15 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 45:

1 5 10 15

(2) INFORMATION FOR SEQ ID NO: 46:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 35 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 30
(D) OTHER INFORMATION: /note= "H represents the Nucleotide Inosine(I)"

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 46:

CCCAGGCTC CAGGCGCCAR KOGATAACGN GRTCG 35

(2) INFORMATION FOR SEQ ID NO: 47:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 47:

ACTAGTCUC ATGCTGTC TRGNGCTGCT CYTCTG 36

(2) INFORMATION FOR SEQ ID NO: 48:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 48:

TAATACGACT CACTATAGG 20

(2) INFORMATION FOR SEQ ID NO: 49:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 49:

GTTTCCCAG TCGAAGCT

19

(2) INFORMATION FOR SEQ ID NO: 50:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 39 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 50:

ACTAGTCGAC AGAAGTTC GCTTGGCTG TGGGCTGCT

39

(2) INFORMATION FOR SEQ ID NO: 51:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 51:

CCCAAGCTTA TCTGATGCT GCGAGATGGA

30

(2) INFORMATION FOR SEQ ID NO: 52:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 52:

CAGATGGAAG GCCTCGAC

18

(2) INFORMATION FOR SEQ ID NO: 53:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 53:

GATCGAAGCA TATCATTACC

20

(2) INFORMATION FOR SEQ ID NO: 54:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 54:

GTTATCGATG TCGAATACCC

20

(2) INFORMATION FOR SEQ ID NO: 55:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 55:

TTGCTCCAGA TTGAGTACTG TTCT

(2) INFORMATION FOR SEQ ID NO: 56:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 33 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 56:

GCCCATATCA CCATGAGCTT CTGGGGCTG CTC

(2) INFORMATION FOR SEQ ID NO: 57:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 81 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 57:

TTGCTCATC AAAAAATCGG ATCCGCGCC ACCGAGCCG CCACCGCCCG AGCCACCTCC

CCCTGAGGG ACCGATGACTG A

(2) INFORMATION FOR SEQ ID NO: 58:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 81 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 58:

TCACTCCGG TCTCCCGAG GCGAGTGGGC TCGGGGCTG GCGCTCAGG TCGGCCGAGA

TCCGATGTT TCAGACCCA A

(2) INFORMATION FOR SEQ ID NO: 59:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 34 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 59:

CATCTCTAA GTTATTGATT TCGAGTGGGG TGGCC

(2) INFORMATION FOR SEQ ID NO: 60:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 33 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 60:

GCCCATATCA CCATGAGCTT GCCGTTAGG CTG

(2) INFORMATION FOR SEQ ID NO: 61:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 87 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 61:

TGACTCTCTC ACCCTGACTG GGATCCGGC GCCACCCAGG CCAGCCACCG CCGAGCAACC 60
TCCCCCTTGG AATTCGAGCT TGCTGCC 87

(2) INFORMATION FOR SEQ ID NO: 62:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 87 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 62:

GCCACACGG TGAAATACCA AAGGGGAGGT GGCCTAAGGCT GTGGCCGCTC GGGTGCGGC 60
GGATCCAGG TGCCGCTGAA GGCTGCTA 87

(2) INFORMATION FOR SEQ ID NO: 63:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 34 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 63:

CATCTCTAGA TTAATAGGAG ACGGTGAGCT AGGT 34

(2) INFORMATION FOR SEQ ID NO: 64:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 7 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 64:

ACCATGG 7

(2) INFORMATION FOR SEQ ID NO: 65:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 814 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 13..801

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 65:

GCCGATATCA CC ATG GCT GTC TTG GGG CTG CTC TTC TGC CTG GTG ACA 48
Met Ala Val Leu Gly Leu Leu Phe Cys Leu Val Thr 135 140 145

TTC CCA AGC TGT GTC TTC CAG GTG CAG GTG AAG GAG TCA GGA CCT 96
Phe Pro Ser Cys Val Leu Ser Gln Val Gln Val Lys Glu Ser Gly Pro 150 155 160

TTC CTG GTG CCC CCC TCA AGC CTG TTC ATC ACA TGC ACT GTC TCA 144
Phe Leu Val Pro Pro Ser Gln Ser Leu Ser Ile Thr Cys Thr Val Ser 165 170 175
GGG TTC TCA TTA ACC ACC TAT GGT GTA AGC TGG ATT CGC CAG CCT CCA 
180 185 190
GAG AAG GGT CTG GAG TGG TGC GGA AAT TGG GGT GAC GGG ACC ACA 
195 200 205
Gly Lys Glu Leu Glu Trp Leu Gln Gly Ala Ile Trp Gly Aep Gly Thr Thr 
210 215 220 225
AAT TAT CAT GCT CTC ATA TCC AGA CTG AGC ATT GCC AAG GAT AAC 
230 235 240
Aam Tyr His Ser Ala Leu Ser Arg Leu Ser Ile Ser Lys Aep Asn 
245 250 255
TCC AAG AGC CAA GGT TTC TTA AAA CTG AAC AGT CTG CAA ACT GAT GAC 
336
Ser Lys Ser Glu Val Phe Leu Lys Leu Aam Ser Leu Gln Thr Asp Aep 
340 345 350
ACG GCC AGG TAC TAC TGG GCC AAA CTG GGT AAC TAG CAT GCT CTG GAC 
384
Thr Ala Thr Thr Tyr Cys Ala Lys AAA CTG GGT AAC TAC CAT GCT CTG GAC 
390 395
TAC TGG GGT CAA GGA ACC TCA GTC ACC GTC TCC TCA GGG GGA GGT GCC 
432
Tyr Trp Gly Glu Gly Thr Ser Val Thr Val Ser Ser Gly Gly Gly Gly 
440 445 245
TGG GCC GGC TGG GCC GGC GTC GCC GCA GTC GAT GGT TGG ATG ACC 
450
Ser Gly Gly Gly Ser Gly Ser Gly Ser Asp Val Leu Met Thr 
460 465
CAA ACT CCA CTG TCC CTG GTC CCT GAT GAG GTA AAC GCC TCC ATC 
528
Gln Thr Pro Leu Ser Leu Pro Val Ser Leu Gly Asp Glu Ala Ser Ile 
535 540 545
TCT TCC AGA TCT AGT CAT AGC ATT GTA CAT AGT AAT GTA AAC ACC TAT 
576
Ser Cys Arg Ser Ser Gln Ser Ile Val His Ser Asn Gly Gly Thr Tyr 
580 585
TTA GAA TGG TAC CTA GAG AAA CCA GCC CAG TCT CCA AAC CTC CTG ATC 
624
Leu Glu Trp Tyr Leu Gln Lys Pro Gly Glu Ser Pro Asp Val Leu Ile 
630 635
TAC TTT GTT TCC AAC CGA TTT TCT GGG GTC CCA GAC AGG TCC AGT GGC 
672
Tyr Phe Val Ser Asn Arg Phe Ser Gln Val Pro Asp Arg Phe Ser Gln 
680 685
AGT GGA TCA GGG ACA GAT TCC ACA ATC AGG AGA TGG GAG GCT 
720
Ser Gly Ser Gly Thr Asp Phe Thr Leu Lys Ile Ser Arg Val Glu Ala 
730 735
740
GAG GAT CTG GGA GTT TAT TAC TCC TGC TTT CAA GGT TCA CAT GTT CCG TGG 
768
Glu Asp Leu Val Tyr Tyr Cys Phe Glu Gly Ser His Val Pro Trp 
780 785
AGC TTC GGT GGA GGC ACC AAG CTG GAA ATC AAA TAATCTAGG ATG 
814
Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys 
820 825 830
(2) INFORMATION FOR SEQ ID NO: 66:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 263 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 66:
Met Ala Val Leu Gly Leu Leu Phe Cys Leu Val Thr Phe Pro Ser Cys 
1 5 10 15
Val Leu Ser Glu Val Glu Val Val Lys Glu Ser Gly Pro Phe Leu Val Pro 
20 25 30
Pro Ser Glu Ser Leu Ser Ile Thr Cys Thr Val Ser Gly Phe Ser Leu 
35 40 45
Thr Thr Tyr Gly Val Ser Thr Ile Arg Gin Pro Pro Gly Lys Gly Leu 
50 55 60
What is claimed as the invention is:

1. A monoclonal antibody 1A7 produced from hybridoma cell line deposited under ATCC Accession No. HB-11786.

2. An antibody producing cell, being progeny of hybridoma cells, deposited under ATCC Accession No. HB-11786 said progeny producing a monoclonal antibody which is the same as the monoclonal antibody produced from hybridoma cell line deposited under ATCC Accession No. HB-11786.

3. A purified antibody produced by a cell according to claim 2.

4. A vaccine comprising monoclonal antibody 1A7 of claim 1 and a pharmaceutically acceptable excipient for the treatment of melanoma.

5. The vaccine of claim 4, comprising an adjuvant.

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