Chylomicrons Promote Intestinal Absorption and Systemic Dissemination of Dietary Antigen (ovalbumin) in Mice

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Chylomicrons Promote Intestinal Absorption and Systemic Dissemination of Dietary Antigen (Ovalbumin) in Mice

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Abstract

Background: A small fraction of dietary protein survives enzymatic degradation and is absorbed in potentially antigenic form. This can trigger inflammatory responses in patients with celiac disease or food allergies, but typically induces systemic immunological tolerance (oral tolerance). At present it is not clear how dietary antigens are absorbed. Most food staples, including those with common antigens such as peanuts, eggs, and milk, contain long-chain triglycerides (LCT), which stimulate mesenteric lymph flux and postprandial transport of chylomicrons through mesenteric lymph nodes (MLN) and blood. Most dietary antigens, like ovalbumin (OVA), are emulsifiers, predicting affinity for chylomicrons. We hypothesized that chylomicron formation promotes intestinal absorption and systemic dissemination of dietary antigens.

Methodology/Principal Findings: Absorption of OVA into MLN and blood was significantly enhanced when OVA was gavaged into fasted mice together with LCT compared with medium-chain triglycerides (MCT), which do not stimulate chylomicron formation. The effect of LCT was blocked by the addition of an inhibitor of chylomicron secretion, Pluronic L-81. Adoptively transferred OVA-specific DO11.10 T-cells proliferated more extensively in peripheral lymph nodes when OVA was gavaged with LCT than with MCT or LCT plus Pluronic L-81, suggesting that dietary OVA is systematically disseminated. Most dietary OVA in plasma was associated with chylomicrons, suggesting that these particles mediate systemic antigen dissemination. Intestinal-epithelial CaCo-2 cells secreted more cell-associated, exogenous OVA when stimulated with oleic acid than with butyric acid, and the secreted OVA appeared to be associated with chylomicrons.

Conclusions/Significance: Postprandial chylomicron formation profoundly affects absorption and systemic dissemination of dietary antigens. The fat content of a meal may affect immune responses to dietary antigens by modulating antigen absorption and transport.


Introduction

Our diet contains many potentially antigenic proteins. The majority of these are enzymatically degraded, but a small fraction survives and enters the body through as yet largely unknown mechanisms. In healthy individuals, this usually leads to systemic immunological tolerance (“oral tolerance”), but in sensitized individuals, absorption can cause significant morbidity, such as with celiac disease or food allergies. Intestinal antigen absorption thus is highly important in health and disease, but knowledge of the mechanisms is limited. Mechanisms involved in sampling of gut micro-organisms, such as transcytosis of particulate matter across epithelial microfold-cells [1] or protrusion of dendritic cell extensions across the intestinal epithelium [2] are not known to be involved in the absorption of soluble dietary antigens, and paracellular leakage across the epithelium is unlikely to occur in healthy individuals due to the presence of strong tight junctions [3,4].

Recently, it was found that intestinal epithelial cells (IEC) internalize dietary antigens, such as egg-hen albumin (ovalbumin; “OVA”), at the apical surface and secrete part of the antigens from the basolateral surface in association with vesicles (exosomes) [5–7]. This would allow dendritic cells in the lamina propria to sample the antigens, by internalizing the exosomes. The physiological relevance of exosomal antigen absorption is unclear, and it is not known whether this mechanism is regulated by food intake.

IEC secrete a distinct class of large lipoprotein particles in the postprandial state, the chylomicrons [8,9]. These particles enable transport of intestinally absorbed, poorly soluble long-chain triglycerides (LCT) from the gut to other tissues. Dietary short- or medium-chain triglycerides (MCT) do not require chylomicron

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formation for their absorption and do not stimulate chylomicron secretion. Interestingly, chylomicrons are drained via intestinal lymphatics and are therefore transported through mesenteric lymph nodes (MLN), before reaching the bloodstream at the level of the left-subclavian vein. Postprandial chylomicron formation stimulates mesenteric T-cell proliferation [10,11], which has typically been attributed to mitogenic effects of fatty acids. We have recently demonstrated that chylomicron formation promotes intestinal absorption of bacterial lipopolysaccharides (LPS) [12], and that LPS is transported through the MLN. LPS transport likely occurred in association with chylomicrons, which are known to bind LPS [13,14]. Interestingly, lipoproteins with structural similarity to chylomicrons, HDL, bind many different proteins and peptides [15,16], and dietary antigens, such as OVA, peanut albumins, and milk caseins, have emulsifying properties [17–20], which suggests that protein antigens may also have affinity for chylomicrons.

We therefore hypothesized that intestinal absorption of dietary OVA is enhanced if the OVA is ingested in the context of chylomicron formation. We observed indeed that dietary LCT enhanced OVA absorption compared with MCT, and that the effect of LCT was entirely sensitive to an inhibitor of chylomicron secretion, Pluronic L-81 [21]. We also observed that plasma chylomicrons contained most of the dietary OVA, suggesting that these particles mediated systemic antigen dissemination. This was reflected by strongly enhanced antigen specific proliferation of T-cells in peripheral, non-gut draining lymph nodes. Lastly, in vitro studies with CaCo-2 cells showed that chylomicron secretion correlated with basolateral OVA secretion, suggesting that this mechanism may contribute to antigen absorption. Collectively, these data reveal a novel absorption mechanism for dietary protein antigens, which may profoundly affect immune responses to dietary antigens.

Results

Dietary LCT promote intestinal absorption of dietary OVA in a chylomicron-dependent manner

To test whether dietary LCT promote intestinal antigen absorption in a chylomicron-dependent manner, we gavaged fasted mice with \(^{125}\text{I}-\text{OVA}\) and \([\text{3H}]\)-retinol as a chylomicron marker [22–24], together with either LCT, MCT, or LCT plus the inhibitor of chylomicron secretion, Pluronic L-81 (3% by volume). Kinetics experiments were performed to confirm that Pluronic L-81 effectively blocks intestinal absorption of retinol, and they revealed that peak retinol levels were obtained near 2 h after gavage (not shown). We therefore decided to analyze blood and MLN content of OVA and retinol at 90 minutes after gavage. As shown in Figure 1, dietary LCT significantly increased intestinal absorption of OVA into blood and MLN, compared with MCT, and the effect of LCT was blocked by Pluronic L-81. Retinol absorption mirrored OVA absorption (Figure 1), suggesting that OVA was absorbed and transported in a chylomicron-dependent fashion. Notably, OVA and retinol were both increasingly transported through the MLN, through which chylomicrons are transported. Figure 2B further illustrates the potency of Pluronic L-81 in its inhibiting effect of chylomicron secretion.

Since lack of LCT might lead to delayed OVA appearance in the plasma, we also performed a kinetics experiment, in which fasted mice were gavaged as in Figure 1, except that \(^{125}\text{I}-\text{OVA}\) was replaced with 25 mg unlabeled OVA. Plasma samples were obtained from the submandibular vein at 30, 60, 120 and 240 minutes after gavage, and analyzed for the presence of OVA by Western blotting. As shown in Figure 3, OVA levels were

![Figure 1. Chylomicron formation promotes intestinal OVA absorption.](image1)

![Figure 2. Chylomicron formation promotes intestinal absorption of full-length, antigenic OVA.](image2)
significantly higher in the LCT group at all time points, compared to mice gavaged with MCT or LCT + Pl-81.

Together, these data suggest that dietary LCT promote protein antigen absorption in a chylomicron-dependent manner.

Chylomicron formation promotes systemic dissemination of dietary OVA

To determine whether the absorbed radioactivity represented antigenic OVA, and whether the absorbed antigen is systemically disseminated, we injected CFSE-labeled DO11.10 T-cells isolated from antigen-naive mice into antigen-naive BALB/C mice. After 24 h, the mice were fasted (4 h) and gavaged with OVA (25 mg) in 0.2 ml PBS or 25 mg OVA in 0.05 ml PBS+0.15 ml of either MCT, LCT, or LCT plus PI-81. Mice were then fasted for an additional 6 h. Three days later, inguinal LN cells were isolated, stained with anti-CD4 and KJ1-26 (TCR clonotypic antibody), and analyzed by flow cytometry. Histograms show representative CFSE dilution profiles of gated CD4+, KJ1-26+ T cells as a measure of cell division. The % of cells under markers M1 and M2 represent cells which have not or have undergone cell division respectively. Each panel represents a typical result of three experimental repeats. doi:10.1371/journal.pone.0008442.g003

Thus, chylomicron formation caused increased systemic dissemination of dietary OVA.

Intestinally absorbed OVA is associated with plasma chylomicrons

The increase in systemic dissemination of dietary OVA as a result of chylomicron formation prompted us to test whether chylomicrons themselves were involved in transport, and hence dissemination, of dietary OVA. We therefore tested whether chylomicrons in the plasma are enriched with dietary OVA. Such an association between chylomicrons and OVA would not be unexpected given the emulsifying properties of this antigen [17], predicting affinity for oil-in-water emulsion particles such as chylomicrons. Fasted mice were gavaged with 25 mg OVA in 0.2 ml LCT/PBS emulsion, and plasma, collected 1 h later, was fractionated by FPLC. The chylomicron fraction eluted immediately after the void volume in the first peak, as illustrated by the fact that plasma from mice injected with the lipoprotein lipase inhibitor Poloxamer P-407 1 h before the gavage, which causes plasma chylomicron accumulation, yielded a much larger first peak (Figure 4) without affecting the height of other peaks. Fractions of the eluate obtained from non-Poloxamer treated mice...
were subsequently analyzed for the presence of OVA by immunoprecipitation. Interestingly, virtually all dietary OVA eluted with the chylomicrons in the first peak (Figure 4), suggesting that chylomicrons indeed transport dietary OVA. This could have accounted for systemic antigen dissemination, as observed in Figure 3.

**CaCo-2 cells secrete cell-associated OVA during chylomicron formation**

To explore how chylomicron formation promotes antigen absorption, we studied OVA secretion by CaCo-2 cells under conditions which preclude paracellular OVA leakage. First, however, CaCo-2 cells on glass slides were incubated for 1 h with 20 μg/ml Alexa-red OVA to determine OVA uptake. Fluorescent staining could be observed in association with the cell membrane and with vesicular structures (Figure 5A). Next, we determined whether chylomicron formation promotes basolateral release of OVA from CaCo-2 cells. Fully differentiated cells, grown for three weeks on Transwell membranes, were incubated overnight with 10 mg/ml OVA on the apical side. Unbound OVA was then thoroughly washed from both surfaces with serum-free medium, and the apical chamber received 0.5 mM taurocholate and 1.6 mM oleic acid, butyric acid, or oleic acid plus 0.2% Pl-81. After 16 h, the basolateral medium was collected and analyzed for OVA. As shown in Figure 5B, cells incubated with oleic acid secreted more OVA than cells incubated with butyric acid or with oleic acid plus Pl-81. To verify whether the secreted OVA was associated with chylomicrons, we performed immunoprecipitation with protein-A coupled to Sepharose, with or without prior addition of anti-OVA antibody. Strikingly, pull-down with anti-

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**Figure 4. Plasma chylomicrons transport dietary OVA.** Fasted mice were gavaged with 0.2 ml LCT-containing emulsions also containing 25 mg OVA. Plasma was isolated 1 h later, and 55 μl were fractionated via FPLC. The grey line of the chromatogram shows the elution profile of a mouse injected i.p. with Poloxamer P-407 1 h prior to gavage to inhibit chylomicron clearance, which caused a milky plasma appearance (inset) and a greatly increased first peak. The solid line shows the elution profile of a mouse not previously injected with Poloxamer P-407. The fractions of this mouse, indicated by the vertical separators, were subjected to immunoprecipitation for detection of OVA (lower panel). The experiment was repeated three times with similar outcomes.

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**Figure 5. Uptake and secretion of OVA in association with chylomicrons by intestinal epithelial cells.** (A) CaCo-2 cells were incubated with 20 μg/ml Alexa-red OVA for 1 h at 37°C. Nuclei were stained with DAPI (blue). (B) CaCo-2 cells on Transwell filters were incubated overnight at the apical side with 0.1 mg/ml OVA, washed from both sides, then incubated apically with 1.6 mM oleic acid (OA), butyric acid (BA), or oleic acid plus Pl-81 (2 μl/ml). Basolateral medium was collected 16 h later, and OVA was detected by immunoblotting. OVA was immunoprecipitated from the basolateral medium with anti-OVA coupled to protein A-Sepharose (or protein A-Sepharose only), followed by Western blotting of unwashed precipitate for detection of Apo-B. As shown in (C), ApoB-48 co-precipitated with OVA.

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Discussion

This study provides evidence for a novel role of dietary fat in the intestinal absorption of dietary protein antigens. We observed that dietary LCT promote absorption of OVA into MLN and blood, and that the effect of LCT is sensitive to inhibition of chylomicron secretion. We also observed that dietary OVA in plasma is mainly transported in association with chylomicrons, which likely mediates enhanced systemic dissemination of the antigen as reflected by increased proliferation of T-cells in peripheral lymph nodes not draining the intestine. Finally, we observed that cultured CaCo-2 IEC acquire OVA and partially secrete OVA during chylomicron formation. Moreover, the secreted OVA was associated with OVA secretion from IEC and, importantly, that the secreted OVA was associated with chylomicrons.

OVA resulted in significant pull-down of ApoB-48 (Figure 5C) from the oleic-acid groups. Pull-down with protein-A Sepharose lacking anti-OVA IgG led to limited (a-specific) pull down of ApoB-48. Thus, it appeared as if chylomicron secretion was associated with OVA secretion from IEC and, importantly, that the secreted OVA was associated with chylomicrons.

Materials and Methods

Materials

$^{125}$I-OVA was prepared according to a slightly modified Iodine monochloride procedure [35], using grade V-OVA (Sigma-Aldrich). [3H]-retinol was purchased from American Radiolabeled Chemicals. MCT oil was purchased from Novartis, triolein (LCT), butyric acid, oleic acid and taurocholate from Sigma-Aldrich, and Pluronic L-81 (an inhibitor of chylomicron secretion [12,21,28].
was a gift from BASF chemicals. Poloxamer P-407, a potent inhibitor of lipoprotein lipase [36], was purchased from Spectrum Chemical Manufacturing Corporation. Alexa-red OVA and carboxyfluorescein-succinimidyl ester (CFSE) were obtained from Invitrogen. Protein-A-Sephrorse and horseradish peroxidase-conjugated streptavidin were purchased from Sigma-Aldrich, biotinylated anti-OVA rabbit IgG from Abcam (ab3839), rabbit anti-OVA IgG from Millipore (ab1225), goat-anti Apolipoprotein B (Apo-B) from Calbiochem (178467), the KJ1-26 antibody from eBioscience (13–5808), and anti-mouse CD4 from BD-Pharmin-gen (553049).

Cell CoC-2 human IEC were purchased from ATTC and were cultured in 1:1 DMEM/Ham’s F12, supplemented with 5% fetal calf serum and penicillin/streptomycin/amphotericin (all from Hyclone). To study OVA uptake by CaCo-2 cells, the cells were grown on glass slides and incubated with 20 μg/ml Alexa-red OVA in complete medium for 1 h, fixed with 10% paraformaldehyde in PBS, and covered with DAPI-containing Vectashield (Vectorlabs) before being sealed with a glass coverslip. Cells were observed with an epifluorescence microscope (model BX50; Olympus) equipped with a cooled charge-coupled device camera. To study basolateral secretion of antigen and of chylomicrons, the cells were seeded in 12-well plates on top of Transwell filter inserts (Corning Transwell Clear; 3 μm pore size), and allowed to differentiate for 21 days [37]. To study OVA secretion, CaCo-2 cells were incubated overnight with 10 mg/ml OVA on the apical side and then washed from both surfaces with cold phosphate-buffered saline (PBS). Chylomicron formation was induced or prevented as described elsewhere [12,37]. Briefly, cells were incubated from the apical side with 0.5 mM taurocholate and 1.6 mM oleic acid, with serum-free medium in the basolateral chamber. Control cells were incubated with butyric acid instead of oleic acid or with 2 μl Pl-81/ml. Chylomicron formation was estimated by sodium-dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) under reducing conditions (no boiling) of freeze-dried basolateral medium and immunostaining of Apo-B [12]. The ~250 kDa Apo-B isoform. OVA was detected by immunoblotting after SDS-PAGE (reduced and boiled samples).

Mouse studies

Male BALB/C mice, 6 weeks old (Jackson Laboratory) were held in a room with a 12 h light 12 h dark cycle and were used at 8 weeks of age. The syngenic DO11.10 mice were maintained as a breeding colony. This strain produces CD4 T-cells in which the majority expresses a transgenic T-cell receptor recognizing OVA peptide 323–339 presented on MHC-II [30]. The transgenic T-cell receptor binds the KJ1-26 antibody. Adoptive transfer experiments were carried out as described elsewhere [38,39]. Briefly, splenocytes and lymph node cells from DO11.10 mice were labeled with CFSE and a cell suspension containing 2.5×10^6 CFSE-labeled DO11.10 CD4 T-cells was then injected into the tail vein of BALB/C recipients. Proliferation of KJ1-26+CD4+ T-cells in inguinal lymph nodes of the recipients was estimated by measuring CFSE staining intensity [40]. All animals were handled in strict accordance with good animal practice as defined by the relevant national and/or local animal welfare bodies, and all animal work was approved by the Institutional Animal Care and Use Committee of the University of Kentucky.

Isolation of plasma chylomicrons and detection of associated OVA

Mouse plasma (55 μl) was fractionated by Fast Performance Liquid Chromatography (FPLC) using a BioRad BioLogic Duo-flow system equipped with a Superose 6 column. The eluate was continuously monitored for absorption at 340 nm, and fractions were collected immediately after the void volume. The largest plasma aggregates, chylomicrons and some very low density lipoproteins (VLDL), elute in the first peak. To verify this, some plasma was analyzed from mice injected with the lipoprotein lipase inhibitor Poloxamer P-407 (1000 mg/kg) [36] 1 h before the gavage, which delays clearance of all triglyceride-rich lipoproteins, including chylomicrons. Low- and high density lipoproteins (LDL/HDL) elute later, whereas free proteins elute last. OVA in all fractions was detected by immunoprecipitation using protein-A Sepharose and rabbit anti-OVA IgG, followed by SDS-PAGE and immunoblotting with biotinylated anti-OVA IgG and streptavidin-HRP.

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Author Contributions

Conceived and designed the experiments: JW EE. Performed the experiments: YW SG MW EE. Analyzed the data: YW SG MW. Wrote the paper: EE.

References


