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Aspirin Treatment of Mice Infected with *Trypanosoma cruzi* and Implications for the Pathogenesis of Chagas Disease

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**Abstract**

Chagas disease, caused by infection with *Trypanosoma cruzi*, is an important cause of cardiovascular disease. It is increasingly clear that parasite-derived prostaglandins potently modulate host response and disease progression. Here, we report that treatment of experimental *T. cruzi* infection (Brazil strain) beginning 5 days post infection (dpi) with aspirin (ASA) increased mortality (2-fold) and parasitemia (12-fold). However, there were no differences regarding histopathology or cardiac structure or function. Delayed treatment with ASA (20 mg/kg) beginning 60 dpi did not increase parasitemia or mortality but improved ejection fraction. ASA treatment diminished the profile of parasite- and host-derived circulating prostaglandins in infected mice. To distinguish the effects of ASA on the parasite and host bio-synthetic pathways we infected cyclooxygenase-1 (COX-1) null mice with the Brazil-strain of *T. cruzi*. Infected COX-1 null mice displayed a reduction in circulating levels of thromboxane (TX)A₂ and prostaglandin (PG)F₂α. Parasitemia was increased in COX-1 null mice compared with parasitemia and mortality in ASA-treated infected mice indicating the effects of ASA on mortality potentially had little to do with inhibition of prostaglandin metabolism. Expression of SOCS-2 was enhanced, and TRAF6 and TNFα reduced, in the spleens of infected ASA-treated mice. Ablation of the initial innate response to infection may cause the increased mortality in ASA-treated mice as the host likely succumbs more quickly without the initiation of the “cytokine storm” during acute infection. We conclude that ASA, through both COX inhibition and other “off-target” effects, modulates the progression of acute and chronic Chagas disease. Thus, eicosanoids present during acute infection may act as immunomodulators aiding the transition to and maintenance of the chronic phase of the disease. A deeper understanding of the mechanism of ASA action may provide clues to the differences between host response in the acute and chronic *T. cruzi* infection.

**Introduction**

In Latin America millions of people are at risk of infection with the parasite *Trypanosoma cruzi*, the cause of Chagas disease. The cardiac manifestations are the most prominent symptoms of disease. Acute myocarditis is accompanied by an intense inflammatory response including upregulation of inflammatory mediators such as cytokines, chemokines, nitric oxide and endothelin-1 [1–6]. As the acute infection wanes individuals may remain asymptomatic; however, 10 to 30% of infected individuals ultimately develop chronic cardiomyopathy [7]. Manifestations during this stage of the disease include congestive heart failure, conduction abnormalities and thrombo-embolic events [7,8]. The etiology of the chronic cardiomyopathy is primarily the result of parasite persistence but may also result from microvascular spasm with focal ischemia and autoimmune mechanisms [4,5,9–11]. Our group has investigated the etiology of vascular spasm in the setting of *T. cruzi* infection. In this regard, we suggested as early as 1990 that the eicosanoid, thromboxane (TX)A₂ contributed to *T. cruzi*-associated vasospasm and platelet aggregation [12].
Eicosanoids are a family of lipid mediators that participate in a wide range of biological activities including vascular tone, inflammation, ischemia and tissue homeostasis [13]. In mammals, the biosynthetic pathways for these important biological mediators are well described. Arachidonic acid (AA) derived from membrane phospholipids on the inner leaflet of the plasma membrane by phospholipase A2 is hydrolyzed by the prostaglandin (PGH) synthase/cyclooxygenase (COX) enzymes to PGH2 [14]. PGH2 is the central substrate for subsequent eicosanoid synthesis which is mediated by species-specific synthases to generate PGs and TXA2 [15]. Enzymes in the COX family are structurally and enzymatically similar but have different pathological roles. COX-1 is constitutive and mediates gastric mucus production, platelet activation and vascular tone while COX-2 is inducible and functions in inflammation, cancer and tissue damage [13,14]. The relevance of these enzymes, and the bioactive lipids they produce, are not well understood in parasitic disease.

Phospholipase A1 (PLA1) the enzyme that initiates the AA metabolic pathway by cleaving the Sn-1 acyl chain was reported in T. brucei [16,17]. PGF2α synthases have been identified in Leishmania, T. cruzi (Old Yellow Enzyme) and T. brucei [18,19]. PGF2α is the predominant eicosanoid species produced in Leishmania and T. brucei, along with smaller quantities of PGE2 and PGD2 [16,18,20]. Importantly, T. cruzi preferentially synthesizes TXA2 [21]. Eicosanoids released by T. cruzi may contribute to parasite differentiation, phagocytosis [22] and host survival [23] as acting as immunomodulators to aid transition and maintenance of the chronic phase of the disease. Moreover, recent studies have demonstrated that trypanosomes are capable of AA metabolism complicating the interpretation of the potential significance and source of these bioactive lipids [18–20].

Our recent data [21] indicated that host- and parasite-derived prostaglandins potentially contribute to the pathogenesis of Chagas disease. Given the increasing importance of eicosanoids in T. cruzi infection, it is not unexpected that there should be interest in non-steroidal anti-inflammatory agents (NSAIDS) in the pathogenesis and clinical management of this infection. However, administration of NSAIDS may enhance mortality in patients [23,24] and in experimental T. cruzi infection [25]. Moreover, characterization of COX inhibition on disease exacerbation in T. cruzi has not been fully addressed. We sought to determine what effect NSAID use would have on the development of acute and chronic Chagas disease. To examine the consequences of COX inhibition we administered aspirin (ASA) to T. cruzi infected mice either early in the course of disease, 5 days post infection (dpi) or late in infection (60 dpi). ASA exhibits irreversible inhibition of COX isoforms and is widely used to treat the symptoms of Chagas disease making it the most clinically applicable choice for these studies. COX inhibition early in the disease increased parasitemia and mortality. Administration of ASA during the chronic phase had no effect on mortality or parasitemia but improved ejection fraction. ASA ablated the increased release of PGF2α and TXA2 in response to T. cruzi infection; however, infection of COX-1 null mice only mimicked the effects of ASA on parasitemia, primarily through decreased TXA2 release. The enhanced mortality in response to ASA was likely due to “off-target” effects of ASA. ASA treatment of T. cruzi infected mice suppressed TNF-α release through increased expression of suppressor of cytokine signalling-2 (SOCS-2) and reduced TNF-α receptor-associated factor (TRAF6) expression in the spleen. Thus, the effects of ASA in T. cruzi infection may be via dual mechanisms that operate during different phases of disease.

### Results

**Global inhibition of eicosanoid production early in the course of T. cruzi infection results in increased parasitemia and mortality**

Infected CD-1 mice were treated with either 20 or 50 mg/kg ASA from 5 dpi to address the involvement of COX-derived mediators during acute infection. Over the subsequent 50 days of infection 40% of untreated mice died (Figure 1A). ASA treatment increased mortality during acute infection in a dose dependent manner with 60% and 80% mortality (50 dpi) in the groups treated with 20 and 50 mg/kg ASA, respectively (Figure 1A). Similarly, ASA treatment increased the parasitemia during acute infection by 2.7 and 5.6 fold in the 20 and 50 mg/kg ASA treated groups, respectively (Figure 1B). Conversely, treatment of mice with ASA (20 mg/kg) during the chronic phase (60 dpi) produced no exacerbation of disease (Figure 1C). Delayed administration of ASA did not increase peripheral parasitemia nor did it augment mortality (Figure 1C). Thus, eicosanoid production during acute infection appears to modulate host response and disease evolution in favor of progression to the chronic state.

**ASA treatment during the chronic disease improves ejection fraction**

Declining cardiac function is a significant source of mortality in both experimental models and in patients suffering from Chagas disease. The effects of early (beginning at 5 dpi) or delayed (beginning at 60 dpi) treatment with ASA (20 mg/kg) on cardiac structure and function were determined using magnetic resonance imaging (MRI). Significant dilatation of the right ventricle was observed in mice in response to infection (Figure 2A). Neither early nor delayed treatment with ASA attenuated the right ventricular dilation (Figure 2A and B) with a 2 fold increase in internal diameter in all infected groups. Moreover, a 10% decrease in left ventricular diastolic diameter was also noted across all groups (Figure 2B). Despite this, delayed treatment significantly reversed the reduction in ejection fraction observed in infected mice as determined by echocardiography. The 38% decrease in ejection fraction in infected mice was attenuated to 15% upon delayed treatment with ASA. The improved ejection fraction in the delayed treatment group may be due to reduced inflammation during the chronic stage of the disease. Using microPET imaging we previously observed increased glucose uptake in the hearts of infected mice [26] which correlated with increased inflammation. Thus, our microPET data demonstrating reduced left ventricular glucose uptake (LVSUV; Figure 2D) in the delayed treatment group suggest reduced inflammation in that group compared with untreated infected mice. Unlike the mortality data, treatment at the early timepoints did not restore cardiac function indicating that the therapeutic window for the preservation of cardiac function and host survival are not the same.

The effective preservation of myocardial function by ASA prompted us to examine cardiac tissue from infected mice for hallmarks of disease (Figure 3). Compared to hearts from uninfected mice (Figure 3A and B) infected mice displayed increased inflammation and pseudocysts along with increased cellularity, mostly resulting from influx of inflammatory cells (Figure 3C and D). No significant differences in myocardial histopathology were observed between infected mice with or without ASA treatment with parasite pseudocysts present in both groups (arrows, Figure 3C through F). Thus, the action of ASA did not prevent the pathological changes to the heart induced by experimental T. cruzi infection.
ASA treatment ablates eicosanoid production by both the parasite and host.

The improvements to cardiac function with ASA without alteration to cardiac pathology or structure indicated a humoral mediator in the suppression of cardiac function. TXA2 is an ASA sensitive mediator with robust links to cardiac damage post-infarction and during failure [27,28]. To determine the extent of COX suppression by ASA treatment we measured TXA2 levels (as the stable hydrolytic product TXB2) in the plasma of infected mice by ELISA. TXB2 levels in infected CD-1 mice increased linearly from 10 dpi and peaked at 27 ng/ml plasma at 45 dpi (Figure 4A). In ASA-treated, infected mice TXA2 levels remained at or below those observed in uninfected mice (constant at 8 ng/ml plasma over 45 dpi). ASA treatment ablated TXA2 release in uninfected mice (Figure 4A) and the augmentation of TXA2 release in ASA treated mice due to T. cruzi infection was also blunted when compared to vehicle treated controls. Thus, both host- and parasite-derived eicosanoid synthesis in infected mice appear to be sensitive to COX-inhibition by ASA.

Previously we demonstrated that in T. cruzi-infected TXA2 synthase null mice the majority of TXB2 in the plasma is derived from the parasite, and not the host [21]. TXA2 release in infected TXA2 synthase null mice (Figure 4B) was also ablated by treatment with ASA [20 mg/kg]. The reduction in total TXA2 levels indicated that either the TXA2 biosynthetic pathway in the parasite is significantly inhibited by ASA or that the parasite is dependent upon host-cell derived PGH2. Treatment of cultured epimastigotes, the extracellular life-stage of T. cruzi, with ASA had no effect on parasite proliferation in vitro (data not shown). These results were consistent with previous data suggesting that the biosynthetic pathways of the parasite are resistant to the effects of ASA [18,20,29]. Thus, it appears that the scavenging of prostanoid precursors by the parasite from the host was the most likely hypothesis for the observed effects.

Infection of COX-1 and TXA2 Synthase null mice mimics only some of the changes in ASA treated mice

To confirm the mechanism of ASA action in vivo we examined the pathogenesis in genetically modified mice with attenuated biosynthetic capacity. ASA has a 66 fold preference for COX-1 over COX-2 [30,31]; therefore, COX-1 null mice were chosen to test the hypothesis. TXA2 synthase null mice (normal COX-1 activity by ablated TXA2 synthesis) were used for comparison. Infection of COX-1 and TXA2 synthase null mice with T. cruzi yielded small alterations in the mortality curves but no significant change in the overall survival compared to wild-type (WT) littermates (Figure 5A). Conversely, peripheral parasitemia in infected COX-1 null mice was increased 9.2 fold compared to WT littermates with peak parasitemia increased 5.7 fold and prolongation of parasite levels from 33 dpi to 42 dpi. Similarly, parasitemia in T. cruzi infected TXA2 synthase null mice was increased 7.1 fold over the 50 dpi and accompanied by a 7- fold increase in peak parasitemia and prolongation of parasite persistence until 48 dpi. The kinetics in both genetically modified mouse strains were almost identical suggesting that the majority of these changes were accounted for by a lack of TXA2 production by the host in COX-1 null mice.

Eicosanoid release from infected COX-1 and TXA2 synthase null mice was significantly different from their WT controls. Plasma of infected COX-1 null mice exhibited a 2.1 fold decreased rate of

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Figure 1. Early administration of ASA increases mortality and parasitemia in response to T. cruzi infection. A and B. CD-1 mice were infected with the Brazil strain of T. cruzi and mortality (A) and parasitemia (B) assessed in vehicle (●) and ASA treated mice (□, 20 mg/kg; ▼, 50 mg/kg) over 55 days post infection (dpi). Treatment with ASA or vehicle started at 5 dpi. C. Table showing the effects of delayed ASA treatment on chronic experimental T. cruzi infection. ASA treatment (20 mg/kg) was initiated 60 dpi until 120 dpi. Survival and parasitemia were assessed 120 and 75 dpi respectively. Data are represented as mean ± SD are representative of at least 20 mice per group. * and # indicates significance (P<0.05) from uninfected and infected mice, respectively. ND = not detected.

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synthesis and an overall 70% decrease in PGF<sub>2α</sub> production (Figure 5C) compared to infected WT mice. Unlike WT mice, the plasma of COX-1 null mice did not contain significant levels of TXA<sub>2</sub> (Figure 5D). As stated above, our previous data suggested that most of the circulating TXA<sub>2</sub> in experimental infection is parasite derived [21]. Thus, our current data suggest that scavenging of metabolic intermediates from the host is likely required for parasite eicosanoid biosynthesis. To further support this hypothesis the release of PGF<sub>2α</sub> from infected TXA<sub>2</sub> synthase null mice, which have unaffected generation of precursor molecules, was normal (Figure 5C). Collectively, these data indicate that decreased eicosanoid biosynthesis by ASA only accounts for the control of parasite proliferation in experimental infection while the enhanced mortality observed with early administration of ASA may be a response to alterations in other unrelated pathways.

Effect of ASA on cytokines in T. cruzi infected mice

The enhanced mortality when ASA treatment was initiated early resulted in the hypothesis that ASA treatment might inhibit priming of the innate immune system in order to decrease host response to the parasite. Plasma of mice treated with ASA (20 mg/kg) during acute T. cruzi infection displayed a reduction in TNF-α and IFN-γ levels 15 dpi (data not shown). Immunoblotting of lysates from the spleen of infected mice confirmed that TNF-α expression was significantly reduced by ASA treatment (Figure 6). These findings were not replicated in cardiac tissue (no change in TNF-α production compared to uninfected mice) indicating that the spleen was the likely source of the cytokines present in the plasma and the target organ of ASA treatment. ASA has many off-target effects that influence the inflammatory response including inhibition of NFκB [32], augmentation of SOCS-2 and decrease of TRAF6 expression [33,34]. Treatment with ASA early in infection not only ablated TNF-α release from spleen but also increased SOCS-2 and reduced TRAF6 expression (Figure 6). Changes were observed as early as 15 dpi and were even more pronounced at 30 dpi indicating they were present throughout the acute phase of disease. Thus, the enhanced mortality observed in infected mice treated with ASA early in infection may be due to suppression of the innate immune response as a result of suppression of cytokine synthesis/release and receptor signaling associated with the initial “cytokine storm” during acute infection.

To examine the above hypothesis more directly, we treated infected WT and COX-1 null mice with ASA (20 mg/kg) and examined the effects on survival (Figure 7). Infection of COX-1 null mice with T. cruzi had little effect on host survival with 23.3% mortality over the 50 days of infection. However, infected COX-1 null mice treated with ASA beginning 5 dpi significantly exacerbated mortality with 65% of mice dying in response to
T. cruzi infection over the subsequent 50 days and with a linear rate of loss (1.995 mice/day) from day 26 onwards. As an important target of ASA was removed in these mice we can only speculate that the “off target” effects of ASA we have observed are those that mediate the mortality in response to infection.

Discussion

It is now appreciated that the release of eicosanoids during infection with T. cruzi regulates host responses and controls disease progression [21]. The role of these bioactive lipids in acute and chronic Chagas disease is largely unexplored and potentially further complicated by whether the host or the parasite is the primary source of synthesis. In this study we found that ASA treatment increased mortality and parasitemia in a dose-dependent manner during acute infection with the Brazil-strain of T. cruzi in mice. These changes were due, in part, to suppression of eicosanoid production (primarily TXA2) which controls parasite proliferation and may participate in cytokine release/signaling during early disease. Importantly, our data strongly suggest that control of fever and pain with ASA during acute Chagas disease should be used with caution. Conversely, use of ASA in the chronic phase of disease may improve cardiac function suggesting the same COX-1 products that mediate host-survival during the acute disease are likely to contribute to the progression of cardiac damage and heart failure in the chronic phase. Importantly, we established an essential host-parasite interdependence that dictates the biosynthetic activity of the parasite. This interdependence is an exploitable target for therapy to manage the chronic phase of disease and potentially prevent disease progression.

Previous studies have attempted to document the role of eicosanoids in early disease using pharmacological intervention with mixed results [22,24,25,35,36]. Pharmacological antagonists selective for either COX-1 (ASA), COX-2 (celecoxib) or both (indomethacin) increase mortality and parasitemia (both peripheral blood counts and cardiac parasite nests) regardless of mouse or parasite strain used [23–25,36]. Conversely, others have found inhibition of prostaglandin synthesis/release ablates parasitemia and extends survival in mice infected with T. cruzi [22,35,37]. Our data correlates well with the former group of studies regarding the changes in parasitemia and degree of mortality. Moreover, use of the COX-1 null mice in this study confirms that COX-1 derived mediators from the host contribute to the suppression of parasite proliferation but perhaps not mortality in acute disease. None of the other studies have utilized null mice to confirm the observed effects and therefore it is difficult to know whether mortality and parasitemia are coordinately regulated in other reports or the response to separate properties of the pharmacological antagonists used (as suggested by our study).

The mechanism for the enhanced mortality with NSAID treatment during acute disease may lie with more complete inhibition of prostaglandin synthesis or “off-target” effects of these agents. ASA is not mono-specific and will also inhibit COX-2 [38].

Figure 3. Cardiac pathology in ASA-treated mice is no different to vehicle treated controls. Representative histopathology of infected CD-1 mice with and without ASA treatment (20 mg/kg) at 35 dpi compared with uninfected controls. Sections were stained with H&E. Parasite pseudocysts are observed (arrows). Total magnification of either 100× (A, C, E) or 400× (B, D, F). Images are representative of at least five mice in each group.

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Our data with COX-1 null mice and pharmacological antagonism strongly indicate that host-derived PGH2 is involved in PG synthesis throughout infection. A key question is whether the host or parasite is the primary source of the lipid mediators regulating the pathogenesis of disease. Pharmacological inhibition does not distinguish between these two sources of eicosanoids. The reduction in PGF2α release in COX-1 null, but not TXA2 synthase null, mice indicates that COX activity in the host provides precursor molecules required for the biosynthetic pathways of this parasite. This “scavenging” hypothesis is confirmed by the inability of the parasite (the primary source of TXA2 during infection) to sustain TXA2 release in the COX-1 null mice. If the parasite is scavenging precursors from the host then they would only need the terminal synthases to produce bioactive lipids. Fatty acid biosynthetic pathways in trypanosomes are poorly defined and little homology is reported between the mammalian enzymes and their trypanosomal homologues [20]. Some putative candidates, such as the PGF2α synthase “old yellow enzyme” [19], have been identified. However, reports have indicated that parasitic biosynthetic pathways are resistant to mammalian antagonists, such as ASA, which have little effect on parasite biology [18,20,29]. Conversely, the recent report of anti-parasitic activity of indomethacin derivatives [41] indicates that the active sites of parasite enzymes, if not their primary sequences, are sufficiently homologous to their mammalian counterparts. Recent structural characterization of the target enzyme (TcCYP51), which participates in sterol biosynthesis of T. cruzi, has facilitated understanding of the integral nature of this enzyme to T. cruzi and has revealed much of the kinetics of the mechanism of action of indomethacin amides [42]. Interestingly, no enzyme other than COX isoforms has been identified as sensitive to indomethacin. However, it remains to be determined whether TcCYP51 is an integral component of the eicosanoid biosynthetic pathway in T. cruzi [41].

The identification of the PGH2 derivatives that are most important for disease remains unsolved. Several species of eicosanoids have been implicated in both acute and chronic Chagas disease. Plasma from infected mice displayed increased levels of PGF2α, PGL2, TXA2 [43] and PGE2 [36] compared to uninfected mice from 10 dpi onwards. Previously, we determined that the main prostaglandins derived from T. cruzi are TXA2 and PGF2α [21], indicating that host is the likely source of the elevated PGI2 and PGE2. No specific role has been delineated for the TXA2 synthase null and WT mice were similar indicating this prostaglandin was likely not involved with the augmentation of parasitemia observed in the COX-1 null and ASA treated mice or in the regulation of mortality. This leaves the potential role of PGF2α in Chagas disease largely unexplored; however, the significant amounts of PGF2α produced by T. cruzi, and the fact that all members of the trypanosomatids have an identifiable synthase for PGF2α, indicate that it is of significant value to the parasite.

During acute infection, PGE2 has been shown to modulate the virulence of the T. cruzi strain. A non-lethal strain (K98) provoked elevated circulating PGE2 while lethal strains (RA or K98-2) did not [24]. Inhibition of COX activity (and therefore PGE2 release) increased mortality in K98-strain infected mice but PGE2 infusion did not attenuate the virulence of the RA strain. Inhibition of PGE2 synthesis reduces both inflammatory infiltrates and cardiac fibrosis during acute infection [37]. Conversely, preventing host response to parasite-derived TXA2 augmented death and parasitemia [21]. TXA2 likely regulates vasospasm, thrombosis, vascular permeability and endothelial cell dysfunction during acute disease. TXA2 also displays immunosuppressive properties as WT mice display minimal pathology but TXA2 receptor null mice exhibited pronounced myocardial inflammation with an almost 3-fold...
increased in parasite load in cardiac tissue. Thus, it appears that the eicosanoids present during acute infection largely act as immunomodulators that aid in the transition to and maintenance of the chronic phase of the disease [23]. It is unclear whether T. cruzi generates prostaglandins as a defense against host immune system or whether it hijacks the host prostaglandin metabolic pathway in its favor. To this end, further studies using null mice missing biosynthetic enzymes or receptors are required to fully elucidate the role of the identified prostaglandins in Chagas disease.

In contrast to acute infection, where plasma levels of multiple PGs are elevated, only increased levels of TXA2 are observed in chronic disease (Fig. 5C, D) [43]. In chronic disease the effects of TXA2 largely promote tissue damage, especially in the heart where it may exacerbate myocyte apoptosis and enhance progression to dilated cardiomyopathy and heart failure, a major cause of death in patients with this disease. Thus, disproving the adage that the things that don’t kill you make you stronger. In addition to the maelstrom of changes that TXA2 mediates during acute infection, the secretion of TXA2 would prevent the initiation of an adaptive immune response by the host [44], enabling progression to and maintenance of the chronic phase of the disease. Finally, the role for TXA2 in chronic disease is made more complicated by its control of parasite proliferation. While we have confirmed that TXA2 plays a prominent role in Chagas disease the hypothesis that parasite-derived TXA2 release does not function to suppress peripheral parasitemia in the long term with overall parasite load in the TXA2 synthase null mice 7-fold higher than WT littermates. This produced a late increase in the mortality of TXA2 synthase null mice which was not significant but may point to a need for host-derived TXA2 for control of the severity of the chronic disease. In fact, in the group with delayed ASA treatment there was an improvement in the infection-associated decrease in ejection fraction which may have resulted from negating the detrimental effects of TXA2 on myocyte contractility, platelet function and vascular tone.

In conclusion, our results demonstrate for the first time, that parasitemia and mortality in response to COX blockade during T. cruzi infection may be due to different pathways related to inhibition of prostaglandin synthesis and cytokine release respectively. Our data also show, for the first time, an interdependence of the parasite on host metabolism for prostanoid biosynthesis. These findings advance our understanding of host–parasite relationships and reveal a potential new avenue for pharmacological treatment for a disease with few therapeutic options.

Materials and Methods

Mice

Male CD-1 mice were obtained from Charles River Laboratories (Wilmington, MA). C57Bl/6 and CH3/HeJ mice were obtained from Jackson Laboratories (Bar Harbor, ME). COX-1
null mice and their wild type counterparts (originally from Taconic Farms [Hudson, NY]), TXA2 synthase null (originally from Dr. Kenneth Wu, University of Texas Health Science Center, Houston, TX), were bred in our facility. This study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. All experiments involving mice were approved by the Albert Einstein College of Medicine Institutional Animal Care and Use Committee (Approval Number: 20100204). All efforts were made to minimize suffering during surgical procedures.

**Parasitology and pathology**

The Brazil strain of *T. cruzi* was used in our experiments. The Brazil strain was maintained in C3H/HeJ. Hearts were obtained from infected and uninfected mice, fixed in 10% buffered formalin, paraffin embedded and stained with H&E. Male mice (8–10 weeks) were infected by an intra-peritoneal route with $5 \times 10^4$ trypomastigotes Brazil strain of *T. cruzi* at the inoculums indicated. ASA (Sigma-Aldrich, Saint Louis, MO) was administered daily via intraperitoneal route at a dose of either 20 or 50 mg/kg of body weight beginning 5 (early) or 60 (delayed) dpi and was continued for 60 days. Mortality was recorded and blood drawn for the determination of parasitemia at the intervals stated. Parasitemia was assessed by counting in a hemocytometer chamber.

**Cardiac magnetic resonance imaging (MRI)**

Cardiac MRI of mice infected with *T. cruzi* was first described by our laboratory group [45]. Briefly, the mice were anesthetized with 1.5% isoflurane. A set of standard, shielded, nonmagnetic electrocardiographic leads ending in silver wires were attached to the four limbs. The ECG signal was fed to a Gould ECG amplifier.

**Figure 6. Treatment of *T. cruzi*-infected mice with ASA modulates cytokine signaling in the spleen of infected mice.** Splenic extracts were prepared from uninfected and infected CD1 mice treated with ASA (20 mg/kg) from 5 dpi. Immunoblotting for TNF-α, SOCS-2 and TRAF-6 was performed on 15 and 30 dpi. β-actin was used as a loading control. Immunoblots are representative from at least three separate experiments. doi:10.1371/journal.pone.0016959.g006
right ventricular cavity. The right ventricular internal dimension is the widest point of the wall. The right ventricular internal dimension is the average of the anterior, posterior, lateral, and septal walls. The right ventricular internal dimension is the widest point of the wall is the average of the anterior, posterior, lateral, and septal walls.

The right ventricular (RV) dimensions in millimeters were determined from the images representing end-diastole. The left ventricular (LV) and right ventricle (RV) dimensions in millimeters were determined using MATLAB-based software. Left ventricle (LV) and right ventricle (RV) dimensions in millimeters were determined using MATLAB-based software. The following parameters were used to obtain 8 short axis images of the heart. The gating delay was adjusted to collect data in systole or diastole. The following parameters were used to obtain 8 short axis images of the heart. The gating delay was adjusted to collect data in systole or diastole.

Echocardiography

Echocardiography of mice infected with T. cruzi has been described previously by our laboratory [46]. Briefly, the mice were anesthetized with 1.5% isoflurane-oxygen mixture, which continued throughout the imaging portion of the procedure. Each mouse was placed on a heating pad before and during scanning to maintain normal body temperature. Mice were administered 300–400 μCi (12–15 MBq) in 0.1 ml. normal saline. [18F] fluoro-2-deoxyglucose (FDG), via tail vein and imaging was started at 1 hour after injection as previously described. Imaging was performed using a Concorde Microsystems R4 microPET Scanner, with 24 detector modules, without septa, providing 7.9 cm axial and 12 cm transaxial field of view. Acquisitions were performed in three-dimensional (3D) list mode. Image analysis was performed using ASIPRO VM (Concorde Microsystems, LLC) dedicated software. Manual regions of interest (ROI) were defined around areas of visually identified heart activity in the LV. Successive scrolling through 2 dimensional slices (each 1.2 mm thick in the axial images) permitted identification of a pixel of maximum measured decay-corrected uptake, termed the standardized uptake value, or SUV max. The SUV max is the maximum value of the percentage injected dose per gram of cardiac tissue multiplied by the body weight of each animal.

Measurement of PGF$_2\alpha$ and TXB$_2$ by ELISA

Blood was drawn from the retro orbital plexus in anti-coagulated tubes with heparin (100 U/ml). The blood was centrifuged at 1500g to remove cellular components and the platelet-poor plasma used for the assessment of prostaglandins. Plasma TXA$_2$ and PGF$_2\alpha$ levels were determined using sensitive ELISA kits according to manufacturer’s instructions (Cayman Chemical Company, Ann Arbor, MI). TXA$_2$ levels were determined by measuring the stable hydrolytic product TXB$_2$.

Immunoblotting of Signaling Molecules

For immunoblotting, aliquots of whole cell lysates (30 μg) from spleen and heart were separated by SDS-PAGE under reducing conditions. Proteins were transferred onto nitrocellulose membrane (Protran BA 85 Nitrocellulose from Whatman, Dassel, Germany) and analyzed by immunoblotting using antibodies against SOCS-2, TNFα, and TRAF6 (Santa Cruz Biotechnology, Santa Cruz, CA). Antibodies against β-actin (Ana Spec Inc, San Jose, CA) was used to control for loading.

Statistical Analysis

Data were pooled and statistical analysis was performed using the Mann-Whitney U-test using Sigma Stat Version 2.0. Statistical differences (p<0.05) are indicated on each figure using * or # to denote significance from control and infected groups, respectively.

Author Contributions

Conceived and designed the experiments: LMW HBT AWA. Performed the experiments: SMF HH DZ HSO. Analyzed the data: LAJ SMF LMW HBT AWA. Contributed reagents/materials/analysis tools: CMP WK EJF JEC. Wrote the paper: LAJ LMW HBT AWA.
References


