

Development and field evaluation of transgenic ryegrass (*Lolium* spp.) with down-regulation of main pollen allergens

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Introduction Ryegrass (*Lolium* spp.) pollen is a widespread source of airborne allergens and is a major cause of hayfever and seasonal allergic asthma, which affect approximately 25% of the population in cool temperate climates. The main allergens of ryegrass pollen are the proteins Lol p 1 and Lol p 2. These proteins belong to two major classes of grass pollen allergens to which over 90% of pollen-allergic patients are sensitive. The functional role *in planta* of these pollen allergen proteins remains largely unknown. The generation, analysis and field evaluation of transgenic plants with reduced levels of the main ryegrass pollen allergens, Lol p 1 and Lol p 2 in the most important worldwide cultivated ryegrass species, perennial ryegrass (*L. perenne* L.) and Italian ryegrass (*L. multiflorum* Lam.) are described.

Materials and methods *Lol p 1* and *Lol p 2* cDNA and genomic clones were isolated. Transformation vectors were generated with *Lol p 1* and *Lol p 2* cDNA sequences in antisense orientation under the control of maize and ryegrass pollen-specific promoters.

Results and conclusions Embryogenic suspension cells of perennial and Italian ryegrass were subjected to biolistic transformation with *Lol p 1* and *Lol p 2* antisense vectors and transgenic plants were recovered. The transgenic nature of the perennial and Italian ryegrass plants was confirmed by Southern hybridisation analysis. Transgenic antisense *Lol p 1* and *Lol p 2* ryegrass plants showed a reduction in the levels of the respective pollen allergens assessed with antibodies raised against the recombinant allergenic proteins. Hypo-allergenicity of Lol p 1 down-regulated pollen was confirmed by immunoblots using IgE sera from Lol p 1 sensitised patients. Transgenic antisense *Lol p 1* ryegrass plants showed normal reproductive development and pollen viability. Selected antisense *Lol p 1* transformation events were evaluated in a small-scale field release carried out in Ardmore, Oklahoma, USA in 2004. Mitotic and meiotic stability of transgene integration and expression, as well as general morphology of the field-grown transgenic ryegrass plants were assessed. A more comprehensive assessment of pollen and gene flow of these transgenic ryegrass plants will be evaluated in a planned large-scale field release in USA in 2005. This will complement detailed gene and pollen flow studies undertaken in Hamilton, Australia using non-transgenic novel perennial ryegrass genotypes.

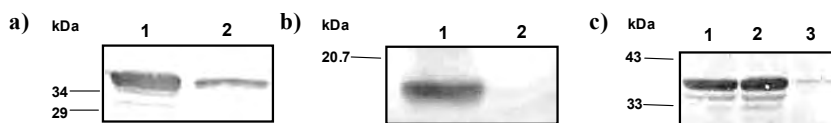


Figure 1 Western blot analysis of pollen protein extracts: a) using anti-Lol p 1 antibodies; untransformed control ryegrass plant (1) and Lol p 1 down-regulated transgenic ryegrass plant; b) using anti-Lol p 2 antibodies; untransformed control ryegrass plant (1) and Lol p 2 down-regulated transgenic ryegrass plant; c) using IgE antibodies from serum of grass pollen allergic patient; perennial ryegrass control plant (1), Italian ryegrass control plant (2) and Lol p 1 down-regulated transgenic ryegrass plant (3)

Reference

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