## University of Kentucky

# **UKnowledge**

[Biosystems and Agricultural Engineering](https://uknowledge.uky.edu/bae_facpub) 

Biosystems and Agricultural Engineering

12-17-2021

# A Literature Review of Wetland Treatment Systems Used to Treat Runoff Mixtures Containing Antibiotics and Pesticides from Urban and Agricultural Landscapes

Emily R. Nottingham University of Kentucky, emily.nottingham@uky.edu

Tiffany L. Messer University of Kentucky, tiffany.messer@uky.edu

Follow this and additional works at: [https://uknowledge.uky.edu/bae\\_facpub](https://uknowledge.uky.edu/bae_facpub?utm_source=uknowledge.uky.edu%2Fbae_facpub%2F252&utm_medium=PDF&utm_campaign=PDFCoverPages)

Part of the [Bioresource and Agricultural Engineering Commons](https://network.bepress.com/hgg/discipline/1056?utm_source=uknowledge.uky.edu%2Fbae_facpub%2F252&utm_medium=PDF&utm_campaign=PDFCoverPages), and the Water Resource Management **[Commons](https://network.bepress.com/hgg/discipline/1057?utm_source=uknowledge.uky.edu%2Fbae_facpub%2F252&utm_medium=PDF&utm_campaign=PDFCoverPages)** 

[Right click to open a feedback form in a new tab to let us know how this document benefits you.](https://uky.az1.qualtrics.com/jfe/form/SV_0lgcRp2YIfAbzvw)

### Repository Citation

Nottingham, Emily R. and Messer, Tiffany L., "A Literature Review of Wetland Treatment Systems Used to Treat Runoff Mixtures Containing Antibiotics and Pesticides from Urban and Agricultural Landscapes" (2021). Biosystems and Agricultural Engineering Faculty Publications. 252. [https://uknowledge.uky.edu/bae\\_facpub/252](https://uknowledge.uky.edu/bae_facpub/252?utm_source=uknowledge.uky.edu%2Fbae_facpub%2F252&utm_medium=PDF&utm_campaign=PDFCoverPages)

This Review is brought to you for free and open access by the Biosystems and Agricultural Engineering at UKnowledge. It has been accepted for inclusion in Biosystems and Agricultural Engineering Faculty Publications by an authorized administrator of UKnowledge. For more information, please contact [UKnowledge@lsv.uky.edu](mailto:UKnowledge@lsv.uky.edu).

## A Literature Review of Wetland Treatment Systems Used to Treat Runoff Mixtures Containing Antibiotics and Pesticides from Urban and Agricultural Landscapes

Digital Object Identifier (DOI) https://doi.org/10.3390/w13243631

## Notes/Citation Information

Published in Water, v. 13, issue 24, 3631.

© 2021 by the authors. Licensee MDPI, Basel, Switzerland.

This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license ([https://creativecommons.org/licenses/by/4.0/\)](https://creativecommons.org/licenses/by/4.0/).



*Review*



# **A Literature Review of Wetland Treatment Systems Used to Treat Runoff Mixtures Containing Antibiotics and Pesticides from Urban and Agricultural Landscapes**

**Emily R. Nottingham [\\*](https://orcid.org/0000-0002-4781-1236) and Tiffany L. Messer**

Department of Biosystems and Agricultural Engineering, University of Kentucky, Lexington, KY 40506, USA; tiffany.messer@uky.edu

**\*** Correspondence: emily.nottingham@uky.edu

**Abstract:** Wetland treatment systems are used extensively across the world to mitigate surface runoff. While wetland treatment for nitrogen mitigation has been comprehensively reviewed, the implications of common-use pesticides and antibiotics on nitrogen reduction remain relatively unreviewed. Therefore, this review seeks to comprehensively assess the removal of commonly used pesticides and antibiotics and their implications for nitrogen removal in wetland treatment systems receiving non-point source runoff from urban and agricultural landscapes. A total of 181 primary studies were identified spanning 37 countries. Most of the reviewed publications studied pesticides  $(n = 153)$  entering wetlands systems, while antibiotics  $(n = 29)$  had fewer publications. Even fewer publications reviewed the impact of influent mixtures on nitrogen removal processes in wetlands (n = 16). Removal efficiencies for antibiotics (35–100%), pesticides (−619–100%), and nitrate-nitrogen (−113–100%) varied widely across the studies, with pesticides and antibiotics impacting microbial communities, the presence and type of vegetation, timing, and hydrology in wetland ecosystems. However, implications for the nitrogen cycle were dependent on the specific emerging contaminant present. A significant knowledge gap remains in how wetland treatment systems are used to treat non-point source mixtures that contain nutrients, pesticides, and antibiotics, resulting in an unknown regarding nitrogen removal efficiency as runoff contaminant mixtures evolve.

**Keywords:** constructed wetlands; nitrogen removal; contaminants of emerging concern; pesticides; antibiotics; non-point source pollution

#### **1. Introduction**

Water quality degradation in rivers and streams across the globe is becoming a major concern, especially as stresses from climate change increase [\[1\]](#page-17-0). The leading cause for surface water impairments in the United States is due to non-point source pollution [\[2\]](#page-17-1). Non-point sources include urban, agricultural, and construction runoff [\[3\]](#page-17-2), which often contain nutrients, suspended sediment, pesticides, antibiotics, and other pharmaceuticals, fecal coliform, and metals [\[3](#page-17-2)[–6\]](#page-18-0). Nutrient contamination, such as nitrate-N ( $NO<sub>3</sub>$ -N) from fertilizer applications, is persistent in rivers and is the most common form globally of chemical contamination in groundwater, resulting in  $NO<sub>3</sub>-N$  being a major cause of drinking water impairment across the United States and the globe [\[7](#page-18-1)[–11\]](#page-18-2). In addition to nutrients, chemically derived common-use pesticide (CUPs) applications throughout the world have increased by 71% from 1990 to 2017, with 11% used for non-agricultural application in the United States [\[12,](#page-18-3)[13\]](#page-18-4). The United States Geological Survey (USGS) recently reported that urban pesticides (e.g., 4-Hydroxychlorothalonil, azoxystrobin, carbendazim, propiconazole, 2,4-D, diuron, prometon, triclopyr, fipronil, imidacloprid) accounted for 83% of the pesticides found in urban sites where the mixture complexity and potential toxicity of the pesticides increased with increasing urbanization [\[14\]](#page-18-5). Additionally, most antibiotics used in human and veterinary medicine (e.g., macrolides, sulfonamides, trimethoprim, quinolones)



**Citation:** Nottingham, E.R.; Messer, T.L. A Literature Review of Wetland Treatment Systems Used to Treat Runoff Mixtures Containing Antibiotics and Pesticides from Urban and Agricultural Landscapes. *Water* **2021**, *13*, 3631. [https://doi.org/](https://doi.org/10.3390/w13243631) [10.3390/w13243631](https://doi.org/10.3390/w13243631)

Academic Editors: Víctor Matamoros and Pedro N. Carvalho

Received: 20 October 2021 Accepted: 14 December 2021 Published: 17 December 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:/[/](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/)  $4.0/$ ).

are excreted via urine and feces. Antibiotics used by humans often enter wastewater treatment systems where removal efficiencies ranged from 15.8–78.4% from 2010 to 2019 [\[15\]](#page-18-6). Veterinary antibiotic residues in livestock products result in 24,000,000–72,000,000 kg of antibiotic residue being introduced into agricultural soil and downstream aquatic environments from manure applications annually [\[16\]](#page-18-7). While CUPs and antibiotics are important for agricultural productivity to support an ever-increasing global population, these chemicals have become ubiquitous in waterways worldwide, resulting in significant effects on agroecosystem food webs (e.g., honeybee colony collapse) [\[17\]](#page-18-8) and human health (e.g., reproductive and development disruption, carcinogens, antibiotic-resistant genes) [\[18](#page-18-9)[,19\]](#page-18-10).

Wetland treatment systems, including natural and constructed wetlands, are now being used extensively across the United States and the world to treat non-point source pollution. This is due to their cost-effectiveness and low energy consumption compared to other surface water treatment methods (e.g., coagulation, membrane filtration, ion exchange) [\[20–](#page-18-11)[22\]](#page-18-12). Wetlands utilize plants, soils, and associated microbial assemblages to remove pollutants through biodegradation, substrate adsorption, and plant uptake [\[23](#page-18-13)[,24\]](#page-18-14). These wetland treatment processes take place in different biogeochemical compartments of the wetland (e.g., aqueous, sediment, detritus), which have separate roles as either sinks or sources [\[25\]](#page-18-15). However, there are limitations to wetland treatment systems including the requirement of large areas and the high variability in performance, as design and operation can be challenging because of the complexity of environmental processes, changing influent concentrations, seasonal changes in weather, and system clogging [\[22\]](#page-18-12).

The use of wetlands as a treatment approach for  $NO<sub>3</sub>-N$  is well known [\[11,](#page-18-2)[26–](#page-18-16)[29\]](#page-18-17); however, N removal performance is significantly impacted by wetland design, operation, and internal and external environmental factors (e.g., microbial communities, plants, type and concentration of contaminants) [\[30\]](#page-18-18). Specifically, contaminants of emerging concerns (CECs), which include CUPs and veterinary antibiotics, in surface waters have only recently started to be investigated for their impact on ecologically important processes (e.g., NO<sub>2</sub>-N oxidizing bacteria population, NH<sub>4</sub><sup>+</sup>-N oxidizing bacteria population) [\[31](#page-18-19)[,32\]](#page-18-20). Exposure to specific CUPs (e.g., imidacloprid) in runoff waters and exposure to antibiotic residues contained in livestock manure and wastewater (e.g., tetracycline, lincomycin, chlortetracycline, sulfamethazine) may influence the population structure of denitrifying bacteria communities [\[33](#page-18-21)[–36\]](#page-19-0), and thus, the activity of microbial denitrification in adjacent wetland treatment systems.

Therefore, this review sought to comprehensively assess past reports related to CUPs and antibiotic treatment in wetlands along with the implications of runoff mixtures for the nitrogen (N) cycle and specifically  $NO<sub>3</sub>-N$  removal in wetland treatment systems receiving runoff from urban and agricultural landscapes. The objectives of this review were to: 1. Evaluate the current scientific status of the topic, 2. Quantify the scale and type of wetlands used to treat runoff mixtures, 3. Identify the source and mixture of antibiotics and CUPs reported to be entering wetlands along with removal mechanisms, and 4. Review implications to N removal processes (e.g., denitrification, plant uptake) in the presence of antibiotics and/or CUPs.

#### **2. Materials and Methods**

This review was designed to better understand the current state of knowledge on the implications of runoff mixtures to the N cycle in wetland treatment systems (specifically the removal of  $NO<sub>3</sub>-N$ ). The review was completed by identifying the methodology and major results about nutrient (N) and CEC (pesticides and antibiotics) removal efficiencies along with implications to the N cycle. The Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) standard was used for analyzing and reporting [\[37\]](#page-19-1). Herein, studies used in this review are referred to as primary studies.

#### *2.1. Eligibility Criteria*

This review sought to include all searchable studies that have researched the ability of wetlands to treat runoff from urban and agricultural landscapes. The primary studies eligible for inclusion in this review needed to meet the following criteria: (1) the study performed analysis on the ability of wetlands to treat CUPs and/or antibiotics from nonpoint source pollution; (2) the study was from a peer-reviewed journal.

#### *2.2. Identification of Records*

The primary studies were obtained from searching Web of Science and SCOPUS databases. A search string was determined through an iterative process to identify studies using any of the search terms in the title, abstract, and/or keywords. The final search string used was the following: (treatment wetland AND antibiotic OR treatment wetland AND pesticide OR constructed wetland AND antibiotic OR constructed wetland AND pesticide). The search was further limited to full-text peer-reviewed journal articles available in English. The official search ended in July 2021 and resulted in 1224 studies meeting the criteria.

Limitations of the search included evaluating only English studies, inevitably underrepresenting the sample of collected studies by excluding those that may have met all other search criteria but were not available in English. Additionally, the keywords used were important to define papers retrieved in the database search; however, it was inevitable that the keywords did not entirely encompass all relevant papers even with careful consideration and several iterations of keywords. These are inherent limitations to all reviews; thus, even though the search was performed systematically, the primary studies may still be biased. However, the limitations do not necessarily take away from the importance and applicability of the findings.

#### *2.3. Screening and Coding of Studies Based upon Eligibility Criteria*

After the initial database search, a three-phase process was used to screen primary studies that met all eligibility criteria (Figure [1\)](#page-5-0) [\[38\]](#page-19-2). Using Rayann [\[39\]](#page-19-3), one coder independently read the title and abstract for all the studies obtained from the initial database search after duplicates were removed. One coder was used throughout the process to maintain consistency and reduce bias; however, if there was any uncertainty regarding the inclusion of a primary study, a second coder was sought for a final decision.

Each study was evaluated to ensure it pertained to wetland treatment of non-point sources. For example, Zheng et al. (2021) was removed from further analysis because the authors simulated domestic sewage waste (point source pollution) to study the removal efficiency of sulfamethoxazole and nitrogen in wetland mesocosms [\[40\]](#page-19-4). Additionally, Maryniuk et al. (2016) considered how high-contaminant loads in wetlands impact largemouth bass [\[41\]](#page-19-5), whereas Sen et al. (2019) focused on the impact of crows that roosted in constructed wetlands and their ability to be carriers of *E. coli* [\[42\]](#page-19-6). Neither of these studies quantified or identified wetland contaminant removal efficiencies or N cycle pathways, so these studies were removed from further analysis. Studies were also removed if only nutrient removal was analyzed without the presence of pesticides or antibiotics. For example, Hussain et al. (2011) analyzed N removal in the presence of antibiotics; thus, the study was retained for further analysis even though the antibiotic removal potential was not specified [\[43\]](#page-19-7); however, Jia et al. (2021) investigated N removal and poly-3-hydroxybutyrate-co-3hydroxyvalerate/polylactic acid removal in the absence of any pesticides or antibiotics, so this study was removed from further analysis [\[44\]](#page-19-8). Review articles were also excluded from the analysis, given that new research on the topic was not conducted; rather, a summary of the existing research was reviewed and/or meta-analysis was completed. From this, there were 181 primary studies used for further analysis [\[3](#page-17-2)[,16](#page-18-7)[,23](#page-18-13)[,25](#page-18-15)[,30](#page-18-18)[,32](#page-18-20)[,43,](#page-19-7)[45–](#page-19-9)[218\]](#page-25-0).

<span id="page-5-0"></span>

**Figure 1.** PRISMA flow diagram detailing the review process (adapted from [38]). **Figure 1.** PRISMA flow diagram detailing the review process (adapted from [\[38\]](#page-19-2)).

The final step of the screening process was to code the primary studies using a com-mon coding schema (Table [1\)](#page-6-0). Key information was coded, including the publication year, location of the study, the scale of the wetland system, number and type of contaminant(s) of concern, removal mechanisms studied, source of runoff, and removal efficiencies. Concerns on correct coding were resolved by the second coder, which included defining wetland systems (e.g., excluding vegetated ditches and riparian buffers), non-point source identification, and identifying removal mechanisms. Throughout the screening and coding process, coders met weekly to check in on progress and to resolve any issues that arose. parameter studies were removed from  $\frac{1}{2}$  and  $\frac$ 



<span id="page-6-0"></span>**Table 1.** Coding schema used for each primary study.

#### **3. Results**

Following the screening, 181 primary studies were identified, with 153 on CUP removal, 29 on antibiotic removal, and 31 on a combination of contaminants. Primary study publication dates ranged from 1995 to 2021, with a linear increase in primary studies on the topics over the last 15 years. The primary studies were drawn from 37 countries.

### *3.1. Bibliometric Source Overview of the Primary Studies*

Publication information, such as geography, was analyzed to provide insight into the annual scientific production growth of research about wetland treatment systems of runoff mixtures to assess Objective 1. Annual scientific production growth on the topic was approximately 29%, showing that this research area is growing, with the peak year of production being 2019 (Figure [2\)](#page-7-0). The decline in publications in 2020 and 2021 could be due to delayed publications from the COVID-19 pandemic. However, 2021 was close behind with 16 publications, even without a complete representative sample of the year, since the studies were collected up to July of 2021. These results demonstrate that research about CUP and antibiotic treatment from wetland systems is growing as more of these contaminants are introduced into the environment and being detected in downstream best management practices and waterways.

The publication sources for the primary studies were indicative of the interdisciplinary nature, broad application, and relevance of the topic, dependent on region. The top publication sources for the primary studies included Chemosphere ( $n = 25$ ), Science of the Total Environment ( $n = 16$ ), and Ecological Engineering ( $n = 16$ ; Table [2\)](#page-7-1). Table [2](#page-7-1) only reports the top five sources. However, 61 total sources were identified. These findings indicate that wetland treatment systems treating non-point source runoff mixtures have broad applicability to a variety of journal sources, which ranged from chemistry, environmental hazards and contaminations, engineering, biology, and ecology-focused journals.

<span id="page-7-0"></span>

contaminants are introduced into the environment and being detected in downstream

**Figure 2.** Annual scientific production of primary studies. **Figure 2.** Annual scientific production of primary studies.

<span id="page-7-1"></span>

Table 2. Results by publication source for the top 5 sources of the primary studies.		



Full primary studies took place intoughout the world, occurring in 37 countries.<br>Countries with the highest number of primary studies included the United States (U.S.;  $\frac{1}{2}$  chemosphere 25, 2 14  $\frac{1}{2}$  25, 2 14  $\frac{1}{2}$  25, 2 14  $\frac{1}{2}$  could also be indicative of water quality concerns and policies limiting the use of emerging contaminants in different regions of the world. For example, the U.S. has approved the use of pesticides and antibiotics (e.g., phorate, dicrotophos, tribufos, oxytetracycline, streptomycin) for outdoor agricultural use, of which many have been banned or phased out by the European Union, Brazil, and China [\[219\]](#page-25-1). In the U.S., the Mississippi River Basin is the largest, most intensively farmed The primary studies took place throughout the world, occurring in 37 countries.  $n = 52$ , China (n = 30), and France (n = 26; Figure [3\)](#page-8-0). This could undoubtedly be due to the region with phorate, dictophos, tirubfos, and oxytetracycline primarily applied to land in the Southeastern region of the United States [\[220\]](#page-25-2). This is represented in Figure [4,](#page-8-1) where the state of Mississippi was the location of 48% of the primary studies in the U.S. Furthermore,

<span id="page-8-0"></span>

these studies were further analyzed based upon experimental design and implications to wetland treatment processes in the sections below.

applied to land in the Southeastern region of the Southeastern region of the United States  $\mathcal{Z}$ 

applied to land in the Southeastern region of the United States [220]. This is represented

**Figure 3.** The spatial distribution of the primary studies across the world.

<span id="page-8-1"></span>

**Figure 4.** The spatial distribution of the primary studies within the continental United States (U.S.).

# 3.2. Scale and Type of Wetlands Used

The scale and type of wetland system used for analysis within the primary studies were coded to determine the scope of the studies along with their impact on the study<br>recalls to see to Okiatine 2. The seals has the relatively impact was real officially of results to assess Objective 2. The scale has the potential to impact removal efficiencies of contaminants by increasing the number of variables in the system, moving from a controlled<br>continuous of the contaminants by increasing the number of variables in the system, moving from a controlled in microcosm and mesocosm experiments) as studies progress from microcosm to full-scale [\[221](#page-26-0)[,222\]](#page-26-1). Microcosm studies utilize repeatable small-scale systems to understand a specific biogeochemical process (e.g., sorption, phytoremediation, microbial activity) in a controlled environment [\[222](#page-26-1)[,223\]](#page-26-2). On the other hand, full-scale systems include major ential the potential to be interested to the potential to the potential to interest environment to a natural environment, and introducing artificial impacts (i.e., wall-effects processes in the natural environment; however, these studies are harder to replicate due to land and cost constraints [\[221\]](#page-26-0). Mesocosm studies are a more cost-effective tool to study contaminant removal in wetlands compared to a full-scale system, while still providing useful predictions using treatment replicates and controls to study wetland efficacy in controlled and natural environments [\[221](#page-26-0)[,224\]](#page-26-3). As for the primary studies, 43% studied full-scale systems, 37% studied mesocosms, and 21% studied microcosms (Figure [5\)](#page-9-0), with only two studies investigating more than one scale [\[53](#page-19-10)[,76\]](#page-20-0).

<span id="page-9-0"></span>

 $21.1\pm 1.0$  studied microcosms (Figure 5), with only two studies investigating more than one than  $\alpha$ 

**Figure 5.** Count of the scale used in the primary studies for the wetland system. **Figure 5.** Count of the scale used in the primary studies for the wetland system.

The primary studies that showed high removal efficiencies (greater than 75%) were The primary studies that showed high removal efficiencies (greater than 75%) were larger for mesocosm studies. Of the studies that analyzed the mesocosm scale, 68% had at larger for mesocosm studies. Of the studies that analyzed the mesocosm scale, 68% had at least one contaminant removed at an efficiency greater than or equal to 75% least one contaminant removed at an efficiency greater than or equal to 75% [\[16](#page-18-7)[,23,](#page-18-13)[45](#page-19-9)[,48,](#page-19-11)[53–](#page-19-10) [16,23,45,48,53–55,58,59,67,69,70,76,79,86,102,104,105,112– [55,](#page-19-12)[58](#page-19-13)[,59,](#page-20-1)[67](#page-20-2)[,69,](#page-20-3)[70,](#page-20-4)[76](#page-20-0)[,79,](#page-20-5)[86](#page-21-0)[,102,](#page-21-1)[104,](#page-21-2)[105](#page-21-3)[,112](#page-21-4)[–114](#page-22-0)[,124,](#page-22-1)[144](#page-23-0)[,151,](#page-23-1)[154,](#page-23-2)[161](#page-23-3)[,163,](#page-23-4)[169](#page-24-0)[–171](#page-24-1)[,175,](#page-24-2)[178](#page-24-3)[–180,](#page-24-4) [182,](#page-24-5)[183](#page-24-6)[,185](#page-24-7)[,192](#page-24-8)[,193,](#page-25-3)[195,](#page-25-4)[201–](#page-25-5)[203,](#page-25-6)[209](#page-25-7)[,210,](#page-25-8)[217](#page-25-9)[–219\]](#page-25-1). Additionally, 54% of microcosm studies 203,209,210,217–219]. Additionally, 54% of microcosm studies resulted in high removal resulted in high removal efficiencies [\[30,](#page-18-18)[60,](#page-20-6)[76,](#page-20-0)[78,](#page-20-7)[83,](#page-20-8)[90,](#page-21-5)[94,](#page-21-6)[97,](#page-21-7)[119,](#page-22-2)[129,](#page-22-3)[131,](#page-22-4)[143,](#page-23-5)[146,](#page-23-6)[164](#page-23-7)[,168,](#page-24-9) efficiencies [30,60,76,78,83,90,94,97,119,129,131,143,146,164,168,177,181,188,194,200,207] [177,](#page-24-10)[181,](#page-24-11)[188](#page-24-12)[,194](#page-25-10)[,200,](#page-25-11)[207\]](#page-25-12) and 59% of field studies resulted in high removal efficiencies [\[3,](#page-17-2)[46,](#page-19-14) [50,](#page-19-15)[53,](#page-19-10)[62](#page-20-9)[,63,](#page-20-10)[68,](#page-20-11)[74](#page-20-12)[,77](#page-20-13)[,80,](#page-20-14)[82,](#page-20-15)[84](#page-20-16)[,85](#page-20-17)[,87,](#page-21-8)[92,](#page-21-9)[95](#page-21-10)[,99,](#page-21-11)[103,](#page-21-12)[106](#page-21-13)[,107,](#page-21-14)[117,](#page-22-5)[119,](#page-22-2)[121,](#page-22-6)[122](#page-22-7)[,126](#page-22-8)[,128,](#page-22-9)[133](#page-22-10)[,135,](#page-22-11)[139,](#page-22-12)<br>114 119 117 150 150 150 150 150 151 160 100 105 105 100 101 100 000 000 000 101 J [141,](#page-23-8)[142](#page-23-9)[,147](#page-23-10)[,150,](#page-23-11)[156](#page-23-12)[,159](#page-23-13)[,160,](#page-23-14)[173,](#page-24-13)[174](#page-24-14)[,184,](#page-24-15)[186,](#page-24-16)[187](#page-24-17)[,189,](#page-24-18)[191,](#page-24-19)[196](#page-25-13)[,206,](#page-25-14)[208,](#page-25-15)[212\]](#page-25-16). However, efficiencies between studies for specific contaminants and/or classes were not able to undergo statistical<br>circles for short-formate disclosure the series contaminants and (or class contacted ward of the two specifical studies for specific contaminants and/or classes were not be two specifical studies evaluation due to few studies having the same contaminant and/or class evaluated.

The type of wetland system was another principal factor to consider because the type of system in a statistical factor the same contaminant of interest  $[225]$ and *or class examples following enforcement* of the communities of interest  $\frac{1}{2\pi\sigma}$ . Freduces are often defined sused apon tryatology (e.g.) free water surface and substitute flow) along with vegetation (e.g., emergent, submerged, floating, and free-floating) [\[226\]](#page-26-5). In this review, 19 different wetland systems were identified throughout the primary studies. These ranged from natural systems to different types of constructed systems including flow) along with vegetation (e.g., emergent, submerged, floating, and free-floating) [226]. agricultural field practices (e.g., rice fields and wetland buffers). Natural wetlands were defined as wetlands that existed naturally in the environment. These included, but were not limited to, floodplains, prairie potholes, depressions, salt marshes, and ephemeral wetlands. Constructed wetland systems included horizontal flow wetlands, subsurface flow wetlands, man-made reservoirs (e.g., ponds, lakes, lagoons, dugouts, and storage dams), free-water surface wetlands, vertical flow wetlands, wetlands in a series, floating treatment wetlands, stormwater basins, and recirculating wetlands. Wetlands used as an of system impacts removal efficiencies, dependent on the contaminant of interest [\[225\]](#page-26-4). agricultural field practice included rice fields, which acted as temporary wetlands with some of the same species as temporary ponds [\[227\]](#page-26-6). Additionally, wetland buffers were defined as a wetland system constructed near agricultural fields to remove contaminants in the water before entering receiving waterbodies.

The most common wetland systems identified were horizontal subsurface flow constructed wetlands ( $n = 42$ ; Table [3\)](#page-10-0). However, while horizontal subsurface flow constructed wetlands were the most common wetland system reviewed, removal efficiencies for CUPs and antibiotics varied widely from 0% norlurazon removal at the field-scale [\[61\]](#page-20-18) to 100% imidacloprid removal at the mesocosm scale [\[210\]](#page-25-8). Additionally, natural wetland systems and reservoirs were analyzed in 15% and 14% of the primary studies, respectively. Both wetlands were used to treat a variety of contaminants (e.g., atrazine, metolachlor, chlorpyrifos, clothianidin, endosulfan, permethrin, prosulfocarb, fluometuron, isoproturon) in full-scale systems. Removal efficiencies ranged from 10% (clothianidin) [\[187\]](#page-24-17) to 100% (permethrin) [\[77\]](#page-20-13) for natural wetlands and 0% (prosulfocarb) [\[128\]](#page-22-9) to 100% (isoproturon) [\[126\]](#page-22-8) for reservoirs. The least studied systems were depressions, ephemeral wetlands, wetland buffers, recirculating constructed wetlands, and salt marshes each appearing only once throughout the primary studies. However, six studies investigated the ability of rice fields to mitigate nutrients, pesticides, and antibiotics as a wetland system in Mississippi, U.S. [\[63,](#page-20-10)[160,](#page-23-14)[174\]](#page-24-14), India [\[56\]](#page-19-16), China [\[115\]](#page-22-13), and Spain [\[103\]](#page-21-12) with high removal efficiencies (58–100%) for several different contaminants (e.g., diazinon, benthocarb, carbofuran, atrazine, permethrin, NH<sub>4</sub><sup>+</sup>-N, NO<sub>3</sub>-N, nitrate-N).



<span id="page-10-0"></span>**Table 3.** Results by wetland type analyzed in the primary studies.

#### *3.3. Source and Mixture of Contaminant Types Entering Wetlands*

Specific design approaches used in the primary studies were analyzed to identify study length (e.g., days, years), type of water analyzed (e.g., urban runoff, rural runoff), and contaminants evaluated to address Objective 3. This was completed to determine which methodological approaches were used most often throughout the primary studies, along with attempting to understand the impact of seasonality, type of landscape runoff, contaminant type, the presence of mixtures of contaminants, and wetland plant type in the primary study results. The length of the study was important to identify the impact of time on wetland treatment. Water type provided insight into runoff from urban and agricultural landscapes and into which landscapes have been studied the most. The specific contaminants studied and the presence of mixtures were coded to determine the most common contaminants and how the contaminant (CUPs and/or antibiotics) impacted  $NO<sub>3</sub>$ -N removal efficiencies. Finally, the type of wetland plants recorded in the primary studies assisted with identifying which wetland plants were most commonly used and if there was an impact on the removal efficiencies of different contaminants based on planting plan.

The length of the study was coded in five different categories: hours, days, weeks, months, and years (Table [4\)](#page-11-0). Hours were defined as any study that took place less than 24 h. Only one study fell into this category, which evaluated removal efficiencies of permethrin in a mesocosm study with four different wetland plant species (i.e., *Leersia oryzoides, Typha latifolia, Sparaganium americanum, Thalia dealbata*) over 12 h [\[54\]](#page-19-17). The second category, days, was defined as an experiment that lasted between 1 to 7 days, which included 13% of the primary studies. The category of weeks was then defined as an experiment that was between 7 to 30 days, with months defined as an experiment that took place between 30 to 365 days, and years being an experiment that was longer than 365 days.



<span id="page-11-0"></span>**Table 4.** Results by the length of the study analyzed by the primary studies.

Months were the most common length of study (53%), and weeks were the second most commonly evaluated (26%). Most of the microcosm studies (67%) addressed periods shorter than months, while a majority of the mesocosm studies (64%) used longer periods. Additionally, several contaminants (e.g., methyl-parathion, malathion, endosulfan, chlorpyrifos, diazinon, tetracycline) were removed within a relatively short period (i.e., less than 10 days) [\[144](#page-23-0)[,168,](#page-24-9)[171,](#page-24-1)[174,](#page-24-14)[177\]](#page-24-10). However, other studies found accumulation of contaminants (e.g., pyrethroid, 2,4-MCPA, glyphosate, propoxycarbazone-Na, NO3- N) [\[68,](#page-20-11)[114](#page-22-0)[,126\]](#page-22-8) with recommendations that the long-term impact on wetlands needs to be further investigated.

The runoff from landscapes defined the water type and included agricultural runoff, urban runoff, both agriculture and urban runoff, or nature reserve. This was coded based on land cover in the watershed for field-scale systems. For mesocosm and microcosm experiments, the primary study either stated where the water was collected from, or the type and concentration of contaminants used for synthetic water. For example, Birch et al. (2004) studied a wetland located in an urban watershed in Sydney, Australia to determine the removal efficiencies of organochlorine pesticides, polycyclic aromatic hydrocarbons, trace metals, nitrogen, phosphorous, and stormwater effluent [\[213\]](#page-25-17). On the other hand, Butkovskyi et al. (2021) used synthetic wastewater to simulate agricultural runoff by applying tap water, fertilizer solution, and different types of pesticides (bentazone, MCPA, metalaxyl, propiconazole, pencycuron, imidacloprid) to potted microcosms containing *Phalaris arundinacea* sp. *Larsa* [\[181\]](#page-24-11). For the primary studies, the most common water type was from agricultural runoff ( $n = 148$ ), with both agriculture and urban runoff encompassing 18 studies, and urban runoff alone assessed in 15 studies (Figure [6\)](#page-12-0). Studies investigating urban and agricultural runoff were either not specific about the water type and instead reported contaminant(s) applied in upstream regions (e.g., sulfonamides used to treat both human and animal infections [\[76](#page-20-0)[,90](#page-21-5)[,170\]](#page-24-20), or the field site for the primary studies' watershed contained both urban and agricultural runoff [\[74,](#page-20-12)[80\]](#page-20-14). Finally, nature reserve was the least studied landscape  $(n = 1)$ ; Tsui et al. (2008) assessed glyphosate concentrations in the Mia Po Nature Reserve in Hong Kong [\[87\]](#page-21-8).

Seven different types of contaminants were studied: nutrients, pesticides, antibiotics, other pharmaceuticals besides antibiotics, metals, minerals, and industrial by-products (Table S1). Pesticides included herbicides, insecticides, and fungicides and were the most common type of contaminant analyzed, appearing 556 times throughout the primary studies with peak study counts occurring between 2016 and 2021 (Figure [7\)](#page-12-1). The most common pesticides studied were atrazine  $(n = 25)$ , chlorpyrifos  $(n = 22)$ , s-metolachlor  $(n = 21)$ , alachlor  $(n = 16)$ , and isoproturon  $(n = 14)$ , along with 158 other pesticides analyzed. Atrazine is a commonly used herbicide, introduced in 1985, and is mainly used for agricultural landscapes while also being used on residential lawns and golf courses, particularly in the Southeastern United States [\[228\]](#page-26-7). Additionally, chlorpyrifos, an insecticide introduced in 1965, and s-metolachlor, an herbicide introduced in 1976, are used for agricultural and non-agricultural landscapes, while alachlor and isoproturon are herbicides mainly used for agriculture.

<span id="page-12-0"></span>

**Figure 6.** Count of non-point source water used by primary studies. **Figure 6.** Count of non-point source water used by primary studies. *2021* **2021 2021 2021 2022 2022 2022 2023 20** 

<span id="page-12-1"></span>

**Figure 7.** Count of contaminant type and mixture by year. **Figure 7.** Count of contaminant type and mixture by year.

Antibiotics appeared throughout the primary studies 60 times, starting in 2007, with Antibiotics appeared throughout the primary studies 60 times, starting in 2007, with peak counts of primary studies  $(n_{\text{antibiotic}} = 9, n_{\text{nutrients and antibiotic}} = 11, n_{\text{nutrients, pesticides, and antibiotic}}$  $=$  1) between 2016 and 2021. The most common antibiotics studied were tetracycline (n = 11), sulfamethoxazole (n = 5), monensin (n = 3), narasin (n = 3), and ciprofloxacin (n = 5), out of a total of 29 primary studies. Of these, narasin and monensin are used as veterinary of a total of 29 primary studies. Of these, narasin and monensin are used as veterinary antibiotics and were found primarily in agricultural runoff, whereas sulfamethoxazole, antibiotics and were found primarily in agricultural runoff, whereas sulfamethoxazole, tetracycline, and ciprofloxacin are used more for humans and were found in urban runoff.

tetracycline, and ciprofloxacin are used more for humans and were found in urban runoff. Additional contaminants found in the primary studies included other pharmaceu-Additional contaminants found in the primary studies included other ticals, metals, minerals, and industrial byproducts. Other pharmaceuticals included any pharmaceuticals, metals, minerals, and industrial byproducts. Other pharmaceuticals pharmaceutical that was not considered an antibiotic (e.g., carbamazepine, caffeine, dipharmaceutical that was not considered an antibiotic (e.g., carbamazepine, canonic) at clofenac, fluoxetine, naproxen, ibuprofen). Industrial byproducts were defined as concaffeine, diclofenac, fluoxetine, naproxen, ibuprofen). Industrial byproducts were defined taminants that are commonly used in industry such as fragrances (e.g., cashmeran, dias contained in the contaminant contained in  $(a, \alpha)$  is induced in induction  $(a, \alpha)$  in  $(a, \alpha)$  as fragrances (e.g., cashmeran, cashmeran, cashmeran, cashmeran, cashmeran, cashmeran, cashmeran, cashmeran, cashmeran, cashm dihydrojasmonate), plastic production (e.g., bisphenol A, di-n-butyl phthalate), dyes (e.g., hydrojasmonate), plastic production (e.g., bisphenol A, di-n-butyl phthalate), dyes (e.g., uranine), corrosion inhibitors (e.g., benzotriazole), and de-icing fluids (e.g., 5-methyl-1Hbenzotriazole). While these contaminants were not the focus of the review, they were observed in the primary studies and are often found in runoff from urban and agricultural landscapes [\[80](#page-20-14)[,99](#page-21-11)[,101](#page-21-15)[,103,](#page-21-12)[112,](#page-21-4)[137,](#page-22-14)[151,](#page-23-1)[213\]](#page-25-17).

There was a total of 9 nutrients analyzed throughout the primary studies, with the most common being TN ( $n = 29$ ), NO<sub>3</sub>-N ( $n = 15$ ), NH<sub>4</sub><sup>+</sup>-N ( $n = 12$ ), total phosphorus (TP,  $n = 10$ ), and nitrite-N (NO<sub>2</sub>-N;  $n = 4$ ). Other nutrients found in the primary studies were orthophosphate, sulfate, chloride, and urea. As for the presence of mixtures, only 30 primary studies looked at contaminant mixtures. Nutrient and pesticide mixtures were the most common mixtures studied, peaking between 2016 and 2021 with 11 primary studies. Only one study in 2020 considered a runoff mixture containing antibiotics, nutrients, pesticides, and other pharmaceuticals [\[103\]](#page-21-12). Of these studies that looked at mixtures, only 16 specifically analyzed the impact on nitrogen removal in the presence of different contaminants [\[23](#page-18-13)[,30](#page-18-18)[,58](#page-19-13)[,59](#page-20-1)[,67](#page-20-2)[,97](#page-21-7)[,102](#page-21-1)[,103](#page-21-12)[,112](#page-21-4)[–114](#page-22-0)[,120](#page-22-15)[,131,](#page-22-4)[169,](#page-24-0)[179,](#page-24-21)[181,](#page-24-11)[201\]](#page-25-5). Thus, the data shows that only recently have some of these contaminants started to be studied individually, in the case of antibiotics, but also as mixtures. This has resulted in the limitation of this review to systematically analyze the implications of wetland design parameters and mechanisms on N removal processes.

#### *3.4. Removal Mechanisms and Efficiencies*

Removal mechanisms were coded based upon the specific processes that were the focus of the primary study to further address Objective 3. Fifteen removal mechanisms were evaluated (Table [5\)](#page-13-0). Varying mechanisms provide insight into the impact on contaminant removal efficiencies, particularly the impact on nitrogen removal processes. Design parameters included size, depth, and aspect ratio of the wetland in addition to the impact of timing (the removal efficiencies over time from contaminant exposure) and space (distance from wetland inlet). If a specific process was not listed and instead removal efficiency of contaminants was analyzed for the wetland system as a whole, then "holistic approach" was coded.



<span id="page-13-0"></span>**Table 5.** Results by removal mechanisms studied for impact on removal efficiencies of contaminants by primary studies.

For the primary studies, the most common removal mechanisms studied were biological processes such as phytoremediation ( $n = 90$ ), sorption ( $n = 69$ ), and microbial activity  $(n = 43)$ . Phytoremediation was coded if the primary study looked at the impact of vegetated vs. non-vegetated systems [\[55,](#page-19-12)[161](#page-23-3)[,175\]](#page-24-2), or analyzed the plant roots [\[23](#page-18-13)[,119](#page-22-2)[,127,](#page-22-16)[180](#page-24-4)[,210\]](#page-25-8), leaves [\[127](#page-22-16)[,210\]](#page-25-8), and stems [\[119,](#page-22-2)[210\]](#page-25-8) for contaminant concentrations. The type of wetland plant used throughout the primary studies helps to identify which wetland plants

were most common and if there was an impact on the removal efficiencies for different contaminants dependent on plant species. There were 92 genera of plants identified, with *Typha* spp. being the most common, comprising 14% of the primary studies, and *Phragmites* spp. used in 11% of the studies (Table [6\)](#page-14-0). It is important to note that not all studies used wetland plants [\[43](#page-19-7)[,83](#page-20-8)[,89\]](#page-21-16) or identified the specific plant species within the wetland system [\[148](#page-23-15)[,159](#page-23-13)[,179\]](#page-24-21). Also, not all primary studies that specified the plant species present in the wetland system analyzed the impact of phytoremediation as a removal mechanism [\[21](#page-18-22)[,173](#page-24-13)[,205\]](#page-25-18). For the studies that did investigate phytoremediation, vegetated systems increased removal efficiencies compared to non-vegetated systems [\[51](#page-19-18)[,55](#page-19-12)[,66](#page-20-19)[,95](#page-21-10)[,154](#page-23-2)[,161](#page-23-3)[,175](#page-24-2)[,181](#page-24-11)[,217\]](#page-25-9), with mature plants out-performing younger ones [\[158\]](#page-23-16).

Plant	Count	<b>Percentage of Primary Studies</b>
Typha spp.	53	14
Phragmites spp.	43	11
Iris spp.	15	4
Carex spp.	14	4
Cyperus sp.	12	З
Phalaris sp.	11	З
Glyceria spp.	10	3
Canna sp.	9	
Scripoides sp.	9	
Other ( $n = 92$ )	210	54

<span id="page-14-0"></span>**Table 6.** Results of the top 10 plant genus used by the primary studies.

As for the impact of the type of plant on removal efficiencies, Lv et al. (2016) concluded that *Typha latifolia*, *Phragmites australis*, *Iris pseudacorus*, and *Juncus effusus* were all able to take up and metabolize imazalil and tebuconazole with removal efficiencies between 46–96% and 25–41%, respectively [\[78\]](#page-20-7). Additionally, Tang et al. (2019) concluded that there were no significant differences in planted systems (*Cyperus alternifolius*, *Canna indica*, *Iris pseudacorus*, *Juncus effusus*, and *Typha orientalis*) and that plants with high biomass and transpiration were able to accelerate the removal of chlorpyrifos and conventional pollutants with removal efficiencies between 94–98% [\[86\]](#page-21-0). On the other hand, some primary studies found that specific species outperformed others, with *Lemna minor* having high removal efficiencies for dimethomorph (17%) and pyrimethanil (12%) compared to *Spirodela polyrhiza* (11–15%) [\[111\]](#page-21-17). Additionally, *Phalaris arundinacea* was better at up taking dicamba, dimethoate, trifloxystrobin, metamitron, and tebuconazole (mean removal of 4%) compared to *Typha latifolia* (2%) [\[95\]](#page-21-10), *Eleocharis mutata* retained less imidacloprid in the plant material and roots (0.5%) compared to *Nymphaea amazonum* (78.9%) [\[210\]](#page-25-8), and *Pontederia cordata* reduced greater amounts of azoxystrobin (51.7%) compared to *Juncus effusus* (24.9%) and *Silene latifolia* (28.7%), while *Silene latifolia* was the best at removing imidacloprid (79.3%) [\[154\]](#page-23-2).

Sorption to wetland media was a removal mechanism of focus for 18% of the primary studies (Table [5\)](#page-13-0). This included studies that compared different media types [\[67,](#page-20-2)[104](#page-21-2)[,170\]](#page-24-20) or analyzed the amount of contaminant sorbed to the wetland media by determining the concentrations of contaminants [\[74,](#page-20-12)[87,](#page-21-8)[135,](#page-22-11)[170,](#page-24-20)[229\]](#page-26-8) or determining sorption isotherm coefficients for the media [\[137,](#page-22-14)[196,](#page-25-13)[198\]](#page-25-19). The different media types found in the primary studies included biochar [\[59,](#page-20-1)[170,](#page-24-20)[179,](#page-24-21)[192\]](#page-24-8), straw [\[133,](#page-22-10)[134\]](#page-22-17), compost [\[170\]](#page-24-20), different types of soil [\[67,](#page-20-2)[145](#page-23-17)[,196,](#page-25-13)[197\]](#page-25-20), gravel [\[104](#page-21-2)[,161](#page-23-3)[,229\]](#page-26-8), pebbles [\[179\]](#page-24-21), zeolite [\[59,](#page-20-1)[114,](#page-22-0)[147,](#page-23-10)[218\]](#page-25-0), and cobbles [\[104\]](#page-21-2).

Soil types and properties (e.g., organic matter content, porosity, structure, moisture content, electrical conductivity) are important components of wetland systems and have been found to impact microbial communities, N cycling, and vegetation growth [\[229\]](#page-26-8). However, these soil properties are dependent upon wetland type (natural vs. constructed wetlands) and age. For example, newly developed, constructed wetlands may have to

overcome soil compaction, resulting in decreased porosity and redox potential, which in turn impacts N cycling [\[230\]](#page-26-9). Overall, the type of contaminants present has been reported to impact sorption processes due to competing cations [\[87\]](#page-21-8) and differences between highly vs. weakly sorbing contaminants [\[54\]](#page-19-17). Additionally, sedimentation or sorption was not the primary removal mechanism in wetland systems with the presence of multiple removal mechanisms (e.g., increased hydraulic retention time, vegetation, microbial activity); instead, it enhanced contaminant removal [\[67](#page-20-2)[,74](#page-20-12)[,84](#page-20-16)[,115](#page-22-13)[,135](#page-22-11)[,196](#page-25-13)[,218\]](#page-25-0). For example, Uddin et al. (2019) found electrical conductivity, total organic carbon, and total nitrogen in the soil significantly impacted microbial richness and diversity [\[115\]](#page-22-13).

Microbial activity is another important removal mechanism in wetland systems because microbial communities facilitate water treatment through metabolic actions (e.g., anabolism, catabolism) [\[231\]](#page-26-10). Microbial communities are mainly found in the rhizospheres, the biofilms around the media, and the water. For the primary studies, 11% focused on microbial activity as a specific removal mechanism. The increased microbial activity enhanced the removal capabilities for CUPS and antibiotics [\[59](#page-20-1)[,60\]](#page-20-6), with microbial degradation being a leading mechanism for removal [\[194\]](#page-25-10). However, the microbial communities were impacted by the substrate [\[59](#page-20-1)[,194\]](#page-25-10), the type and concentration of contaminants present [\[58,](#page-19-13)[114](#page-22-0)[,179\]](#page-24-21), and the physicochemical properties of the water [\[179\]](#page-24-21). In particular, Lu et al. (2021) reported the presence of sulfamethoxazole improved interactions for denitrifying bacteria, but also decreased network complexity and microbial interaction on the whole molecular network, thus altering the community structure of nitrogen-transforming microorganisms [\[114\]](#page-22-0). Yuan et al. (2020) observed the addition of Mn ore impacted microbial diversity, causing increased removal potential for antibiotics (ciprofloxacin hydrochloride and sulfamethazine), TN,  $NH_4^+$ -N, and  $NO_3$ -N [\[59\]](#page-20-1).

#### *3.5. The Impact of CUPs and Antibiotics on Nitrogen Removal*

Overall, 31 primary studies analyzed and reported contaminant removal efficiencies for runoff mixtures containing nitrogen, pesticides, and/or antibiotics to address Objective 4 (Table S2). The removal efficiencies for each contaminant varied widely, with total nitrogen ranging from 5% to 99% removal and a mean of 58%. Ammonia-N had a similar range of 7% to 100% and a mean of 75% removal. Nitrate-N removal varied widely from  $-113%$  to 98%; however, only one primary study found an accumulation of NO<sub>3</sub>-N, which was attributed to the amount of zeolite used in the mesocosm cells [\[114\]](#page-22-0). As for the CUPs and antibiotics studied, the removal efficiencies varied from −619% to 100% and 35% to 100% for pesticides and antibiotics respectively, with negative removal efficiencies being attributed to runoff and remobilization of pesticides in full-scale systems.

The location of the wetlands for these 31 primary studies with the highest removal efficiencies of TN and  $NO<sub>3</sub>-N$  occurred in the United States [\[67](#page-20-2)[,99\]](#page-21-11), while the highest removal efficiencies of NH4-N and pesticides occurred in the United States and Greece [\[77](#page-20-13)[,99](#page-21-11)[,218\]](#page-25-0). In contrast, the highest efficiencies of antibiotic removal occurred in China [\[58\]](#page-19-13). However, the variation in removal efficiencies reported throughout the primary studies was likely due to the wide range of CUPS and antibiotics studied, the type of wetland systems (e.g., scale, wetland type, plant type), and the climatic conditions of the wetland analyzed. For example, Lu et al. (2021) reported that removal efficiencies of NH<sub>4</sub><sup>+</sup>-N were most affected by temperature, rather than the concentration of contaminants, with increasing removal efficiencies occurring at higher temperatures and with increased contact time [\[113\]](#page-22-18). For the 31 primary studies identified as analyzing runoff mixtures containing nitrogen, CUPs, and/or antibiotics, only two studies analyzed natural wetland systems and found relatively high removal efficiencies (43–100%) for TN, NO<sub>3</sub>-N, NH<sub>4</sub><sup>+</sup>-N, atrazine, S-metolachlor, and Permethrin [\[77,](#page-20-13)[184\]](#page-24-15). The lowest removal efficiencies for TN and  $NH_4^+$ -N were found with full-scale horizontal flow constructed wetlands [\[3,](#page-17-2)[213\]](#page-25-17), while the lowest removal efficiencies for  $NO<sub>3</sub>-N$  and antibiotics were found with mesocosm vertical flow constructed wetlands [\[114\]](#page-22-0). In contrast, the lowest removal efficiencies for pesticides were associated with a full-scale wetland buffer [\[126\]](#page-22-8). Despite these external climatic and wetland

design parameters, the leading cause for N-removal disruptions is thought to be due to contaminant mixture type and concentration.

Several primary studies reported the impact of CUPs' and antibiotics' presence on nitrogen removal efficiencies. Of these, seven studies reported that the specific contaminant analyzed decreased nitrogen removal [\[30,](#page-18-18)[58,](#page-19-13)[67,](#page-20-2)[113,](#page-22-18)[114,](#page-22-0)[169\]](#page-24-0). The impact of CUPS and antibiotics on nutrient removal was mainly attributed to a decline in the microbial communities responsible for nutrient metabolism and degradation [\[30](#page-18-18)[,114](#page-22-0)[,131,](#page-22-4)[169\]](#page-24-0), where the presence of plants and type of plant species [\[23,](#page-18-13)[102,](#page-21-1)[120,](#page-22-15)[201\]](#page-25-5), saturated vs. unsaturated conditions [\[120,](#page-22-15)[201\]](#page-25-5), weather conditions [\[201\]](#page-25-5), and timing [\[48](#page-19-11)[,169\]](#page-24-0) also played a role in N removal in the presence of CUPS or antibiotics. Ohore et al. (2021) reported that the presence of tetracycline decreased nitrogen removal initially, but observed an increase in total N removal with an increasing number of days due to the degradation of antibiotics in the wetland system [\[169\]](#page-24-0). Additionally, Tong et al. (2019) observed the presence of plants protected the microbial communities, limiting ofloxacin's ability to negatively impact  $NO<sub>3</sub>$ -N and  $NH<sub>4</sub>$ <sup>+</sup>-N removal [\[30\]](#page-18-18).

However, the presence of CUPs and antibiotics has also been observed not to affect nitrogen removal in wetland treatment systems [\[23,](#page-18-13)[158,](#page-23-16)[201\]](#page-25-5), and in some cases, the CUPs or antibiotics increased N removal [\[43](#page-19-7)[,131\]](#page-22-4). These increases in N removal are presumed to be due to an increase in microbial community diversity and richness in the presence of CUPS or antibiotics [\[23](#page-18-13)[,171\]](#page-24-1). Yu et al. (2019) observed that tetracycline had a slightly negative effect on nitrogen removal (91% to 71%); however, the presence of copper and tetracycline led to higher microbial richness with increases in microbial variations [\[171\]](#page-24-1). In contrast, Lu et al. (2021) observed that the presence of sulfamethoxazole had positive influences on denitrifying bacteria interactions, but reduced the network complexity and microbial interactions in the wetland mesocosms [\[114\]](#page-22-0).

Runoff mixtures not only impact N removal efficiencies, but also impact the wetlands' ability to remove CUPs and antibiotics. Recent investigations on the impact of N on CUPs indicated that nitrifying bacteria can also degrade certain pesticides (e.g., metribuzin, imazalil, tebuconazole) [\[59,](#page-20-1)[102,](#page-21-1)[120\]](#page-22-15). However, in some cases, the presence of nutrients decreased CUP or antibiotic removal efficiencies in the wetland [\[171](#page-24-1)[,205\]](#page-25-18). For example, Matamoros et al. (2020) reported both the presence of nutrients impacted CECs removal and that the presence of other CECs (caffeine, tributyl phosphate, 5TTri, bisphenol A, benzotriazole, carbamazepine, diclofenac, ibuprofen, lorazepam, naproxen, oxazepam, primidone, and triclosan) can reduce pesticide (sulfonyl 104, alachlor, bentazone, chlorpyrifos, DEET, molinate, oxadiazone, propanil, tebuconazole, and MCPA) removal in rice fields [\[103\]](#page-21-12).

#### **4. Conclusions**

This review sought to analyze 181 primary studies that used wetlands of varying scales to treat runoff from urban and agricultural landscapes; this was accomplished by assessing the bibliometric information of the primary studies, identifying wetland scale and type, quantifying the source and mixture of CUPs and antibiotics entering wetlands, identifying mechanisms of removal used to treat runoff mixtures, and analyzing the implications of these mixtures to nitrogen removal processes (e.g., denitrification, plant uptake). While the scientific production of wetland treatment systems used to treat runoff has increased by 29%, of the primary studies reviewed, only 16 wetland treatment systems received urban runoff, while 82% of the studies used wetlands to treat agricultural runoff. Additionally, antibiotics and runoff mixtures have only recently begun to be studied. This has resulted in only 31 of the primary studies evaluating removal efficiencies in the presence of runoff mixtures, with only one study analyzing mixtures containing nutrients, antibiotics, and pesticides.

The impacts on CUP (−619–100%), antibiotic (35–100%), and N (TN = 5–99%,  $NO<sub>3</sub>-N=-113-98%$ ,  $NH<sub>4</sub>-N = 7-100%$  removal efficiencies varied greatly over the primary studies. Variations were likely due to the wide range of different types of

CUPs ( $n = 556$ ) and antibiotics ( $n = 60$ ) analyzed. Additionally, the location, scale of the wetland system (microcosm, mesocosm, field-scale), presence and type of plants, the substrate used, weather conditions, wetland type, and design parameters all impacted the efficacy of the wetland system for water quality treatment. However, the largest impact on nitrogen removal was the shift in microbial community diversity and richness in the presence of CECs; however, this shift was seen to both increase and decrease microbial communities depending on the contaminant mixture, showing a lack of understanding of how microbial communities are impacted by different types and mixtures of CUPs and antibiotics. In addition, long-term exposure analyses of wetland treatment processes and efficacy were limited.

Throughout the primary studies, a lack of uniform reporting on wetland removal performance appeared, with a portion of the studies reporting removal efficiencies, defined as the difference between the influent and effluent mean concentrations over the influent concentration, while others reported removal loads, defined as a mass balance removal rate. Some primary studies also reported on sorption isotherms, microbial populations, or toxicity. In addition, the wide variety of different classes of CUPs and antibiotics analyzed throughout the primary studies made it difficult to perform further statistical analysis to compare wetland type, scale, length of study, and removal mechanisms with removal efficiencies. A significant knowledge gap remains in urban wetland treatment systems, which are continuing to become more important as urban centers continue to grow. Even more, a significant knowledge gap remains in how wetland treatment systems are used to treat non-point source mixtures containing antibiotics and CUPs, resulting in a significant unknown regarding nitrogen removal efficiency in wetland systems as contaminant runoff mixtures and mechanisms for treatment evolve.

**Supplementary Materials:** The following are available online at [https://www.mdpi.com/article/10](https://www.mdpi.com/article/10.3390/w13243631/s1) [.3390/w13243631/s1,](https://www.mdpi.com/article/10.3390/w13243631/s1) Table S1: Contaminant type and appearances in the primary studies. Table S2: Contaminant removal efficiencies for the primary studies that analyzed both nitrogen and pesticide and/or antibiotics in wetland treatment systems.

**Author Contributions:** Conceptualization, T.L.M. and E.R.N.; methodology, E.R.N. and T.L.M.; validation, E.R.N. and T.L.M.; formal analysis, E.R.N.; investigation, E.R.N.; data curation, E.R.N.; writing—original draft preparation, E.R.N.; writing—review and editing, E.R.N. and T.L.M.; visualization, E.R.N.; supervision, T.L.M.; project administration, T.L.M.; funding acquisition, T.L.M. All authors have read and agreed to the published version of the manuscript.

**Funding:** This material is based upon work supported by the National Science Foundation under Grant No. (2042761). This project was also supported with funds by the United States Hatch multistate capacity funding grant (W-4045). Any opinions, findings, and conclusions, or recommendations expressed in this material are those of the author(s) and do not necessarily reflect the views of the National Science Foundation or United States Department of Agriculture.

**Informed Consent Statement:** Not applicable.

**Acknowledgments:** A special thanks to Mathew Russel, Dayana Nayelly Rodriguez Jimenez, Jacob Richardson, Kyra Sigler, and William Rud for their support and encouragement throughout the project.

**Conflicts of Interest:** The authors declare no conflict of interest.

#### **References**

- <span id="page-17-0"></span>1. Murdoch, P.S.; Baron, J.S.; Miller, T.L. Potential Effects of Climate Change on Surface-Water Quality in North America. *JAWRA* **2000**, *36*, 347–366. [\[CrossRef\]](http://doi.org/10.1111/j.1752-1688.2000.tb04273.x)
- <span id="page-17-1"></span>2. EPA. Protecting Water Quality from Agricultural Runoff. Available online: [https://nepis.epa.gov/Exe/ZyPDF.cgi/P10039OH.](https://nepis.epa.gov/Exe/ZyPDF.cgi/P10039OH.PDF?Dockey=P10039OH.PDF) [PDF?Dockey=P10039OH.PDF](https://nepis.epa.gov/Exe/ZyPDF.cgi/P10039OH.PDF?Dockey=P10039OH.PDF) (accessed on 9 September 2021).
- <span id="page-17-2"></span>3. White, K.D.; Meyers, A.L. Above par storm-water management. *Civ. Eng.* **1997**, *67*, 50.
- 4. Panthi, S.; Sapkota, A.R.; Raspanti, G.; Allard, S.M.; Bui, A.; Craddock, H.A.; Murray, R.; Zhu, L.; East, C.; Handy, E.; et al. Pharmaceuticals, herbicides, and disinfectants in agricultural water sources. *Environ. Res.* **2019**, *174*, 1–8. [\[CrossRef\]](http://doi.org/10.1016/j.envres.2019.04.011)
- 5. Jaimes-Correa, J.C.; Snow, D.D.; Bartelt-Hunt, S.L. Seasonal occurrence of antibiotics and a beta agonist in an agriculturallyintensive watershed. *Environ. Pol.* **2015**, *205*, 87–96. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2015.05.023)
- <span id="page-18-0"></span>6. Vymazal, J.; Brezinova, T. The use of constructed wetlands for removal of pesticides from agricultural runoff and drainage: A review. *Environ. Int.* **2015**, *75*, 11–20. [\[CrossRef\]](http://doi.org/10.1016/j.envint.2014.10.026)
- <span id="page-18-1"></span>7. Dwivedi, D.; Mohanty, B. Hot Spots and Persistence of Nitrate in Aquifers Across Scales. *Entropy* **2016**, *18*, 25. [\[CrossRef\]](http://doi.org/10.3390/e18010025)
- 8. Molenat, J.; Gascuel-Odoux, C.; Ruiz, L.; Gruau, G. Role of water table dynamics on stream nitrate export and concentration in agricultural headwater catchment (France). *J. Hydrol.* **2008**, *348*, 363–378. [\[CrossRef\]](http://doi.org/10.1016/j.jhydrol.2007.10.005)
- 9. Mittelstet, A.R.; Gilmore, T.E.; Messer, T.; Rudnick, D.R.; Heatherly, T. Evaluation of selected watershed characteristics to identify best management practices to reduce Nebraskan nitrate loads from Nebraska to the Mississippi/Atchafalaya River basin. *Agric. Ecosyst. Environ.* **2019**, *277*, 1–10. [\[CrossRef\]](http://doi.org/10.1016/j.agee.2019.02.018)
- 10. Katz, B.G.; Berndt, M.P.; Crandall, C.A. Factors affecting the movement and persistence of nitrate and pesticides in the surficial and upper Floridan aquifers in two agricultural areas in the southeastern United States. *Environ. Earth Sci.* **2013**, *71*, 2779–2795. [\[CrossRef\]](http://doi.org/10.1007/s12665-013-2657-8)
- <span id="page-18-2"></span>11. Karpuzcu, M.E.; Stringfellow, W.T. Kinetics of nitrate removal in wetlands receiving agricultural drainage. *Ecol. Eng.* **2012**, *42*, 295–303. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2012.02.015)
- <span id="page-18-3"></span>12. Roser, M. Pesticides—Our World in Data. Available online: <https://ourworldindata.org/pesticides> (accessed on 9 September 2021).
- <span id="page-18-4"></span>13. Atwood, D.; Paisley-Jones, C. Pesticides Industry Sales and Usage, 2008–2012 Market Estimates. Available online: [https:](https://www.epa.gov/pesticides/pesticides-industry-sales-and-usage-2008-2012-market-estimates) [//www.epa.gov/pesticides/pesticides-industry-sales-and-usage-2008-2012-market-estimates](https://www.epa.gov/pesticides/pesticides-industry-sales-and-usage-2008-2012-market-estimates) (accessed on 9 September 2021).
- <span id="page-18-5"></span>14. Nowell, L.H.; Moran, P.W.; Bexfield, L.M.; Mahler, B.J.; Van Metre, P.C.; Bradley, P.M.; Schmidt, T.S.; Button, D.T.; Qi, S.L. Is there an urban pesticide signature? Urban streams in five U.S. regions share common dissolved-phase pesticides but differ in predicted aquatic toxicity. *Sci. Total Environ.* **2021**, *793*, 148453. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2021.148453)
- <span id="page-18-6"></span>15. Wang, J.; Chu, L.; Wojnarovits, L.; Takacs, E. Occurrence and fate of antibiotics, antibiotic resistant genes (ARGs) and antibiotic resistant bacteria (ARB) in municipal wastewater treatment plant: An overview. *Sci. Total Environ.* **2020**, *744*, 140997. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2020.140997) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32755790)
- <span id="page-18-7"></span>16. Zhao, C.; Xie, H.J.; Mu, Y.; Xu, X.; Zhang, J.; Liu, C.; Liang, S.; Ngo, H.H.; Guo, W.; Xu, J.; et al. Bioremediation of endosulfan in laboratory-scale constructed wetlands: Effect of bioaugmentation and biostimulation. *Environ. Sci. Pol. Res.* **2014**, *21*, 12827–12835. [\[CrossRef\]](http://doi.org/10.1007/s11356-014-3107-1) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24969425)
- <span id="page-18-8"></span>17. Chauzat, M.P.; Faucon, J.P. Pesticide residues in beeswax samples collected from honey bee colonies (*Apis mellifera* L.) in France. *Pest Manag. Sci.* **2007**, *63*, 1100–1106. [\[CrossRef\]](http://doi.org/10.1002/ps.1451) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17879980)
- <span id="page-18-9"></span>18. Anderson, J.C.; Dubetz, C.; Palace, V.P. Neonicotinoids in the Canadian aquatic environment: A literature review on current use products with a focus on fate, exposure, and biological effects. *Sci. Total Environ.* **2015**, *505*, 409–422. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2014.09.090)
- <span id="page-18-10"></span>19. Ben, Y.; Fu, C.; Hu, M.; Liu, L.; Wong, M.H.; Zheng, C. Human health risk assessment of antibiotic resistance associated with antibiotic residues in the environment: A review. *Environ. Res.* **2019**, *169*, 483–493. [\[CrossRef\]](http://doi.org/10.1016/j.envres.2018.11.040)
- <span id="page-18-11"></span>20. Trepel, M. Assessing the cost-effectiveness of the water purification function of wetlands for environmental planning. *Ecol. Complex.* **2010**, *7*, 320–326. [\[CrossRef\]](http://doi.org/10.1016/j.ecocom.2010.02.006)
- <span id="page-18-22"></span>21. Weerakoon, G.M.P.R.; Jinadasa, K.B.S.N.; Herath, G.B.B.; Mowjood, M.I.M.; Ng, W.J. Applicability of constructed wetlands for water quality improvement in a tea estate catchment: The Pussellawa case study. *Water* **2018**, *10*, 332. [\[CrossRef\]](http://doi.org/10.3390/w10030332)
- <span id="page-18-12"></span>22. Fitch, M.W. Constructed Wetlands. *Compr. Water Qual. Purif.* **2014**, *3*, 268–295. [\[CrossRef\]](http://doi.org/10.1016/B978-0-12-382182-9.00053-0)
- <span id="page-18-13"></span>23. Tong, X.N.; Wang, X.Z.; He, X.J.; Wang, Z.; Li, W.X. Effects of antibiotics on microbial community structure and microbial functions in constructed wetlands treated with artificial root exudates. *Environ. Sci. Process. Impacts* **2020**, *22*, 217–226. [\[CrossRef\]](http://doi.org/10.1039/C9EM00458K)
- <span id="page-18-14"></span>24. Qasaimeh, A.; AlSharie, H.; Masoud, T. A Review on Constructed Wetlands Components and Heavy Metal Removal from Wastewater. *J. Environ. Prot.* **2015**, *6*, 710–718. [\[CrossRef\]](http://doi.org/10.4236/jep.2015.67064)
- <span id="page-18-15"></span>25. Moore, M.T.; Schulz, R.; Cooper, C.M.; Smith, S.; Rodgers, J.H., Jr. Mitigation of chlorpyrifos runoff using constructed wetlands. *Chemosphere* **2002**, *46*, 827–835. [\[CrossRef\]](http://doi.org/10.1016/S0045-6535(01)00189-8)
- <span id="page-18-16"></span>26. Kadlec, R.H.; Tanner, C.C.; Hally, V.M.; Gibbs, M.M. Nitrogen spiraling in subsurface-flow constructed wetlands: Implications for treatment response. *Ecol. Eng.* **2005**, *25*, 365–381. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2005.06.009)
- 27. Borin, M.; Tocchetto, D. Five year water and nitrogen balance for a constructed surface flow wetland treating agricultural drainage waters. *Sci. Total Environ.* **2007**, *380*, 38–47. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2006.12.039)
- 28. Uwimana, A.; van Dam, A.A.; Irvine, K. Effects of conversion of wetlands to rice and fish farming on water quality in valley bottoms of the Migina catchment, southern Rwanda. *Ecol. Eng.* **2018**, *125*, 76–86. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2018.10.019)
- <span id="page-18-17"></span>29. Bachand, P.A.M.; Horne, A.J. Denitrification in constructed free-water surface wetlands: I. very high nitrate removal rates in a macrocosm study. *Ecol. Eng.* **2000**, *14*, 9–15. [\[CrossRef\]](http://doi.org/10.1016/S0925-8574(99)00016-6)
- <span id="page-18-18"></span>30. Tong, X.; Wang, X.; He, X.; Xu, K.; Mao, F. Effects of ofloxacin on nitrogen removal and microbial community structure in constructed wetland. *Sci. Total Environ.* **2019**, *656*, 503–511. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2018.11.358) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30522033)
- <span id="page-18-19"></span>31. Pontius, F.W. Emerging Contaminants in Water: Detection, Treatment, and Regulation. *Water* **2021**, *13*, 1470. [\[CrossRef\]](http://doi.org/10.3390/w13111470)
- <span id="page-18-20"></span>32. Stoler, A.; Walker, B.; Hintz, W.; Jones, D.; Lind, L.; Mattes, B.; Schuler, M.; Relyea, R. Combined Effects of Road Salt and an Insecticide on Wetland Communities. *Environ. Toxicol. Chem.* **2017**, *36*, 771–779. [\[CrossRef\]](http://doi.org/10.1002/etc.3639)
- <span id="page-18-21"></span>33. Tu, C.M. Effect of five insecticides on microbial and enzymatic activities in sandy soil. *J. Environ. Sci. Health, Part B* **1995**, *30*, 289–306. [\[CrossRef\]](http://doi.org/10.1080/03601239509372940)
- 34. Shan, J.; Yang, P.; Rahman, M.M.; Shang, X.; Yan, X. Tetracycline and sulfamethazine alter dissimilatory nitrate reduction processes and increase N2O release in rice fields. *Environ. Pol.* **2018**, *242*, 788–796. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2018.07.061)
- 35. Zhang, M.; Wang, W.; Tang, L.; Heenan, M.; Xu, Z. Effects of nitrification inhibitor and herbicides on nitrification, nitrite and nitrate consumptions and nitrous oxide emission in an Australian sugarcane soil. *Biol. Fertil. Soils* **2018**, *54*, 697–706. [\[CrossRef\]](http://doi.org/10.1007/s00374-018-1293-6)
- <span id="page-19-0"></span>36. Robson, S.V.; Rosi, E.J.; Richmond, E.K.; Grace, M.R. Environmental concentrations of pharmaceuticals alter metabolism, denitrification, and diatom assemblages in artificial streams. *Freshw. Sci.* **2020**, *39*, 256–267. [\[CrossRef\]](http://doi.org/10.1086/708893)
- <span id="page-19-1"></span>37. Moher, D.; Liberati, A.; Tetzlaff, J.; Altman, D.G.; Group, P. Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. *PLoS Med.* **2009**, *6*, e1000097. [\[CrossRef\]](http://doi.org/10.1371/journal.pmed.1000097) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19621072)
- <span id="page-19-2"></span>38. Page, M.J.; McKenzie, J.E.; Bossuyt, P.M.; Boutron, I.; Hoffmann, T.C.; Mulrow, C.D.; Shamseer, L.; Tetzlaff, J.M.; Akl, E.A.; Brennan, S.E.; et al. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. *BMJ* **2021**, *372*, n71. [\[CrossRef\]](http://doi.org/10.1136/bmj.n71) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33782057)
- <span id="page-19-3"></span>39. Ouzzani, M.; Hammady, H.; Fedorowicz, Z.; Elmagarmid, A. Rayyan-a web and mobile app for systematic reviews. *Syst. Rev.* **2016**, *5*, 210. [\[CrossRef\]](http://doi.org/10.1186/s13643-016-0384-4)
- <span id="page-19-4"></span>40. Zheng, Y.; Liu, Y.; Qu, M.; Hao, M.; Yang, D.; Yang, Q.; Wang, X.C.; Dzakpasu, M. Fate of an antibiotic and its effects on nitrogen transformation functional bacteria in integrated vertical flow constructed wetlands. *Chem. Eng. J.* **2021**, *417*, 129272. [\[CrossRef\]](http://doi.org/10.1016/j.cej.2021.129272)
- <span id="page-19-5"></span>41. Martyniuk, C.J.; Doperalski, N.J.; Prucha, M.S.; Zhang, J.L.; Kroll, K.J.; Conrow, R.; Barber, D.S.; Denslow, N.D. High contaminant loads in Lake Apopka's riparian wetland disrupt gene networks involved in reproduction and immune function in largemouth bass. *Comp. Biochem. Physiol. Part D Genom. Proteom.* **2016**, *19*, 140–150. [\[CrossRef\]](http://doi.org/10.1016/j.cbd.2016.06.003)
- <span id="page-19-6"></span>42. Sen, K.; Berglund, T.; Soares, M.A.; Taheri, B.; Ma, Y.; Khalil, L.; Fridge, M.; Lu, J.; Turner, R.J. Antibiotic resistance of *E. coli* Isolated from a Constructed Wetland Dominated by a Crow Roost, with Emphasis on ESBL and AmpC Containing *E. coli*. *Front. Microbiol.* **2019**, *10*, 1034. [\[CrossRef\]](http://doi.org/10.3389/fmicb.2019.01034) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31156579)
- <span id="page-19-7"></span>43. Hussain, S.A.; Prasher, S.O.; Chenier, M.; Arya, G. Removal of nitrate-N by antibiotic exposed bacterial isolates from constructed wetlands. *World J. Microbiol. Biotechnol.* **2011**, *27*, 2061–2069. [\[CrossRef\]](http://doi.org/10.1007/s11274-011-0668-8)
- <span id="page-19-8"></span>44. Jia, L.; Sun, H.; Zhou, Q.; Zhao, L.; Wu, W. Pilot-scale two-stage constructed wetlands based on novel solid carbon for rural wastewater treatment in southern China: Enhanced nitrogen removal and mechanism. *J. Environ. Manag.* **2021**, *292*, 112750. [\[CrossRef\]](http://doi.org/10.1016/j.jenvman.2021.112750)
- <span id="page-19-9"></span>45. Moore, M.T.; Rodgers, J.H., Jr.; Smith, S., Jr.; Cooper, C.M. Mitigation of metolachlor-associated agricultural runoff using constructed wetlands in Mississippi, USA. *Agric. Ecosyst. Environ.* **2001**, *84*, 169–176. [\[CrossRef\]](http://doi.org/10.1016/S0167-8809(00)00205-X)
- <span id="page-19-14"></span>46. Schultz, R.; Colletti, J.; Isenhart, T.; Simpkins, W.; Mize, C.; Thompson, M. Design and placement of a multi-species riparian buffer strip system. *Agroforest. Syst.* **1995**, *29*, 201–226. [\[CrossRef\]](http://doi.org/10.1007/BF00704869)
- 47. Runes, H.; Jenkins, J.; Bottomley, P. Atrazine degradation by bioaugmented sediment from constructed wetlands. *Appl. Microbiol. Biotechnol.* **2001**, *57*, 427–432. [\[CrossRef\]](http://doi.org/10.1007/s002530100792) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11759697)
- <span id="page-19-11"></span>48. Nguyen, H.; Chao, H.; Chen, K. Treatment of Organic Matter and Tetracycline in Water by Using Constructed Wetlands and Photocatalysis. *Appl. Sci.* **2019**, *9*, 2680. [\[CrossRef\]](http://doi.org/10.3390/app9132680)
- 49. Cedillo-Herrera, C.I.G.; Roé-Sosa, A.; Pat-Espadas, A.M.; Ramírez, K.; Rochín-Medina, J.; Amabilis-Sosa, L.E. Efficient malathion removal in constructed wetlands coupled to UV/H2O<sup>2</sup> pretreatment. *Appl. Sci.* **2020**, *10*, 5036. [\[CrossRef\]](http://doi.org/10.3390/app10155306)
- <span id="page-19-15"></span>50. Bouldin, J.; Farris, J.; Moore, M.; Smith, S.; Cooper, C. Assessment of diazinon toxicity in sediment and water of constructed wetlands using deployed Corbicula fluminea and laboratory testing. *Arch. Environ. Contam. Toxicol.* **2007**, *53*, 174–182. [\[CrossRef\]](http://doi.org/10.1007/s00244-006-0180-6)
- <span id="page-19-18"></span>51. Lizotte, R.E., Jr.; Moore, M.T.; Locke, M.A.; Kröger, R. Role of vegetation in a constructed wetland on nutrient-pesticide mixture toxicity to Hyalella azteca. *Arch. Environ. Contam. Toxicol.* **2011**, *60*, 261–271. [\[CrossRef\]](http://doi.org/10.1007/s00244-010-9596-0)
- 52. Lizotte, R.; Testa, S.; Locke, M.; Steinriede, R. Responses of Phytoplankton and Hyalella azteca to Agrichemical Mixtures in a Constructed Wetland Mesocosm. *Arch. Environ. Contam. Toxicol.* **2013**, *65*, 474–485. [\[CrossRef\]](http://doi.org/10.1007/s00244-013-9927-z)
- <span id="page-19-10"></span>53. Mazanti, L.; Rice, C.; Bialek, K.; Sparling, D.; Stevenson, C.; Johnson, W.E.; Kangas, P.; Rheinstein, J. Aqueous-phase disappearance of atrazine, metolachlor, and chlorpyrifos in laboratory aquaria and outdoor macrocosms. *Arch. Environ. Contam. Toxicol.* **2003**, *44*, 67–76. [\[CrossRef\]](http://doi.org/10.1007/s00244-002-1259-3)
- <span id="page-19-17"></span>54. Moore, M.T.; Kröger, R.; Cooper, C.M.; Smith, S., Jr. Ability of four emergent macrophytes to remediate permethrin in mesocosm experiments. *Arch. Environ. Contam. Toxicol.* **2009**, *57*, 282–288. [\[CrossRef\]](http://doi.org/10.1007/s00244-009-9334-7)
- <span id="page-19-12"></span>55. Schulz, R.; Moore, M.T.; Bennett, E.R.; Milam, C.D.; Bouldin, J.L.; Farris, J.L.; Smith, S., Jr.; Cooper, C.M. Acute toxicity of methyl-parathion in wetland mesocosms: Assessing the influence of aquatic plants using laboratory testing with Hyalella azteca. *Arch. Environ. Contam. Toxicol.* **2003**, *45*, 331–336. [\[CrossRef\]](http://doi.org/10.1007/s00244-003-2170-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/14674585)
- <span id="page-19-16"></span>56. Dash, N.P.; Kumar, A.; Kaushik, M.S.; Abraham, G.; Singh, P.K. Agrochemicals influencing nitrogenase, biomass of N2-fixing cyanobacteria and yield of rice in wetland cultivation. *Biocatal. Agric. Biotechnol.* **2017**, *9*, 28–34. [\[CrossRef\]](http://doi.org/10.1016/j.bcab.2016.11.001)
- 57. Guo, X.; Wang, P.; Li, Y.; Zhong, H.; Li, P.; Zhang, C.; Zhao, T. Effect of copper on the removal of tetracycline from water by Myriophyllum aquaticum: Performance and mechanisms. *Bioresour. Technol.* **2019**, *291*, 121916. [\[CrossRef\]](http://doi.org/10.1016/j.biortech.2019.121916) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31377514)
- <span id="page-19-13"></span>58. Ohore, O.E.; Zhang, S.; Guo, S.; Addo, F.G.; Manirakiza, B.; Zhang, W. Ciprofloxacin increased abundance of antibiotic resistance genes and shaped microbial community in epiphytic biofilm on Vallisneria spiralis in mesocosmic wetland. *Bioresour. Technol.* **2021**, *323*, 124574. [\[CrossRef\]](http://doi.org/10.1016/j.biortech.2020.124574)
- <span id="page-20-1"></span>59. Yuan, Y.; Yang, B.; Wang, H.; Lai, X.; Li, F.; Salam, M.M.A.; Pan, F.; Zhao, Y. The simultaneous antibiotics and nitrogen removal in vertical flow constructed wetlands: Effects of substrates and responses of microbial functions. *Bioresour. Technol.* **2020**, *310*, 123419. [\[CrossRef\]](http://doi.org/10.1016/j.biortech.2020.123419) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32361200)
- <span id="page-20-6"></span>60. Zhao, X.; Bai, S.; Li, C.; Yang, J.; Ma, F. Bioaugmentation of atrazine removal in constructed wetland: Performance, microbial dynamics, and environmental impacts. *Bioresour. Technol.* **2019**, *289*, 121618. [\[CrossRef\]](http://doi.org/10.1016/j.biortech.2019.121618)
- <span id="page-20-18"></span>61. Wilson, P.; Lu, H.; Lin, Y. Norflurazon and Simazine Removal from Surface Water Using a Constructed Wetland. *Bull. Environ. Contam. Toxicol.* **2011**, *87*, 426–430. [\[CrossRef\]](http://doi.org/10.1007/s00128-011-0380-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21833732)
- <span id="page-20-9"></span>62. Moore, M.; Kroger, R.; Locke, M.; Lizotte, R.; Testa, S.; Cooper, C. Diazinon and Permethrin Mitigation Across a Grass-Wetland Buffer. *Bull. Environ. Contam. Toxicol.* **2014**, *93*, 574–579. [\[CrossRef\]](http://doi.org/10.1007/s00128-014-1357-8)
- <span id="page-20-10"></span>63. Moore, M.; Lizotte, R.; Kroger, R. Efficiency of Experimental Rice (*Oryza sativa* L.) Fields in Mitigating Diazinon Runoff Toxicity to Hyalella azteca. *Bull. Environ. Contam. Toxicol.* **2009**, *82*, 777–780. [\[CrossRef\]](http://doi.org/10.1007/s00128-009-9696-6)
- 64. Moore, M.T.; Lizotte, R.E., Jr.; Smith, S., Jr. Responses of Hyalella azteca to a pyrethroid mixture in a constructed wetland. *Bull. Environ. Contam. Toxicol.* **2007**, *78*, 245–248. [\[CrossRef\]](http://doi.org/10.1007/s00128-007-9135-5)
- 65. Gikas, G.; Vryzas, Z.; Tsihrintzis, V. S-metolachlor herbicide removal in pilot-scale horizontal subsurface flow constructed wetlands. *Chem. Eng. J.* **2018**, *339*, 108–116. [\[CrossRef\]](http://doi.org/10.1016/j.cej.2018.01.056)
- <span id="page-20-19"></span>66. Gikas, G.D.; Pérez-Villanueva, M.; Tsioras, M.; Alexoudis, C.; Pérez-Rojas, G.; Masís-Mora, M.; Lizano-Fallas, V.; Rodríguez-Rodríguez, C.E.; Vryzas, Z.; Tsihrintzis, V.A. Low-cost approaches for the removal of terbuthylazine from agricultural wastewater: Constructed wetlands and biopurification system. *Chem. Eng. J.* **2018**, *335*, 647–656. [\[CrossRef\]](http://doi.org/10.1016/j.cej.2017.11.031)
- <span id="page-20-2"></span>67. Lizotte, R.E., Jr.; Locke, M.A.; Testa Iii, S. Influence of varying nutrient and pesticide mixtures on abatement efficiency using a vegetated free water surface constructed wetland mesocosm. *Chem. Ecol.* **2014**, *30*, 280–294. [\[CrossRef\]](http://doi.org/10.1080/02757540.2013.861823)
- <span id="page-20-11"></span>68. Budd, R.; O'Geen, A.; Goh, K.S.; Bondarenko, S.; Gan, J. Removal mechanisms and fate of insecticides in constructed wetlands. *Chemosphere* **2011**, *83*, 1581–1587. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2011.01.012) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21296378)
- <span id="page-20-3"></span>69. Caldelas, C.; Gurí, R.; Araus, J.; Sorolla, A. Effect of ZnO nanoparticles on Zn, Cu, and Pb dissolution in a green bioretention system for urban stormwater remediation. *Chemosphere* **2021**, *282*, 131045. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2021.131045)
- <span id="page-20-4"></span>70. Cheng, S.; Vidakovic-Cifrek, Ž.; Grosse, W.; Karrenbrock, F. Xenobiotics removal from polluted water by a multifunctional constructed wetland. *Chemosphere* **2002**, *48*, 415–418. [\[CrossRef\]](http://doi.org/10.1016/S0045-6535(02)00097-8)
- 71. Dosnon-Olette, R.; Trotel-Aziz, P.; Couderchet, M.; Eullaffroy, P. Fungicides and herbicide removal in Scenedesmus cell suspensions. *Chemosphere* **2010**, *79*, 117–123. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2010.02.005)
- 72. Elsayed, O.; Maillard, E.; Vuilleumier, S.; Nijenhuis, I.; Richnow, H.; Imfeld, G. Using compound-specific isotope analysis to assess the degradation of chloroacetanilide herbicides in lab-scale wetlands. *Chemosphere* **2014**, *99*, 89–95. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2013.10.027)
- 73. Gebremariam, S.Y.; Beutel, M.W. Effects of drain-fill cycling on chlorpyrifos mineralization in wetland sediment-water microcosms. *Chemosphere* **2010**, *78*, 1337–1341. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2010.01.006)
- <span id="page-20-12"></span>74. Imfeld, G.; Lefrancq, M.; Maillard, E.; Payraudeau, S. Transport and attenuation of dissolved glyphosate and AMPA in a stormwater wetland. *Chemosphere* **2013**, *90*, 1333–1339. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2012.04.054) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22633860)
- 75. Kuchta, S.L.; Cessna, A.J. Fate of lincomycin in snowmelt runoff from manure-amended pasture. *Chemosphere* **2009**, *76*, 439–446. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2009.03.069)
- <span id="page-20-0"></span>76. Kurade, M.; Xiong, J.; Govindwar, S.; Roh, H.; Saratale, G.; Jeon, B.; Lim, H. Uptake and biodegradation of emerging contaminant sulfamethoxazole from aqueous phase using Ipomoea aquatica. *Chemosphere* **2019**, *225*, 696–704. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2019.03.086)
- <span id="page-20-13"></span>77. Lizotte, R.; Shields, F.; Murdock, J.; Knight, S. Responses of Hyalella azteca and phytoplankton to a simulated agricultural runoff event in a managed backwater wetland. *Chemosphere* **2012**, *87*, 684–691. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2011.12.058)
- <span id="page-20-7"></span>78. Lv, T.; Zhang, Y.; Casas, M.E.; Carvalho, P.N.; Arias, C.A.; Bester, K.; Brix, H. Phytoremediation of imazalil and tebuconazole by four emergent wetland plant species in hydroponic medium. *Chemosphere* **2016**, *148*, 459–466. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2016.01.064)
- <span id="page-20-5"></span>79. Maillard, E.; Lange, J.; Schreiber, S.; Dollinger, J.; Herbstritt, B.; Millet, M.; Imfeld, G. Dissipation of hydrological tracers and the herbicide S-metolachlor in batch and continuous-flow wetlands. *Chemosphere* **2016**, *144*, 2489–2496. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2015.11.027) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26630289)
- <span id="page-20-14"></span>80. Matamoros, V.; Arias, C.A.; Nguyen, L.X.; Salvadó, V.; Brix, H. Occurrence and behavior of emerging contaminants in surface water and a restored wetland. *Chemosphere* **2012**, *88*, 1083–1089. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2012.04.048) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22633078)
- 81. Page, D.; Dillon, P.; Mueller, J.; Bartkow, M. Quantification of herbicide removal in a constructed wetland using passive samplers and composite water quality monitoring. *Chemosphere* **2010**, *81*, 394–399. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2010.07.002)
- <span id="page-20-15"></span>82. Passeport, E.; Tournebize, J.; Chaumont, C.; Guenne, A.; Coquet, Y. Pesticide contamination interception strategy and removal efficiency in forest buffer and artificial wetland in a tile-drained agricultural watershed. *Chemosphere* **2013**, *91*, 1289–1296. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2013.02.053) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23535469)
- <span id="page-20-8"></span>83. Rodríguez-Liébana, J.A.; ElGouzi, S.; Peña, A. Laboratory persistence in soil of thiacloprid, pendimethalin and fenarimol incubated with treated wastewater and dissolved organic matter solutions. Contribution of soil biota. *Chemosphere* **2017**, *181*, 508–517. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2017.04.111)
- <span id="page-20-16"></span>84. Rose, M.T.; Crossan, A.N.; Kennedy, I.R. The effect of vegetation on pesticide dissipation from ponded treatment wetlands: Quantification using a simple model. *Chemosphere* **2008**, *72*, 999–1005. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2008.04.059)
- <span id="page-20-17"></span>85. Rose, M.T.; Sanchez-Bayo, F.; Crossan, A.N.; Kennedy, I.R. Pesticide removal from cotton farm tailwater by a pilot-scale ponded wetland. *Chemosphere* **2006**, *63*, 1849–1858. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2005.10.024)
- <span id="page-21-0"></span>86. Tang, X.Y.; Yang, Y.; McBride, M.B.; Tao, R.; Dai, Y.N.; Zhang, X.M. Removal of chlorpyrifos in recirculating vertical flow constructed wetlands with five wetland plant species. *Chemosphere* **2019**, *216*, 195–202. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2018.10.150) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30368084)
- <span id="page-21-8"></span>87. Tsui, M.; Chu, L.M. Environmental fate and non-target impact of glyphosate-based herbicide (Roundup®) in a subtropical wetland. *Chemosphere* **2008**, *71*, 439–446. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2007.10.059) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18155747)
- 88. Vandermeeren, P.; Baken, S.; Vanderstukken, R.; Diels, J.; Springael, D. Impact of dry-wet and freeze-thaw events on pesticide mineralizing populations and their activity in wetland ecosystems: A microcosm study. *Chemosphere* **2016**, *146*, 85–93. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2015.11.089)
- <span id="page-21-16"></span>89. Vilas-Boas, J.A.; Arenas-Sánchez, A.; Vighi, M.; Romo, S.; Brink, P.J.V.D.; Dias, R.J.P.; Rico, A. Multiple stressors in Mediterranean coastal wetland ecosystems: Influence of salinity and an insecticide on zooplankton communities under different temperature conditions. *Chemosphere* **2021**, *269*, 129381. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2020.129381) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33383245)
- <span id="page-21-5"></span>90. Yin, T.; Te, S.H.; Reinhard, M.; Yang, Y.; Chen, H.; He, Y.; Gin, K.Y.H. Biotransformation of Sulfluramid (N-ethyl perfluorooctane sulfonamide) and dynamics of associated rhizospheric microbial community in microcosms of wetland plants. *Chemosphere* **2018**, *211*, 379–389. [\[CrossRef\]](http://doi.org/10.1016/j.chemosphere.2018.07.157) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30077934)
- 91. Kanjilal, T.; Bhattacharjee, C.; Datta, S. Utilization of *S. aureusstrain* 502A in biodegradation of insecticide acetamiprid from wetland wastewater. *Desalination Water Treat.* **2015**, *57*, 13190–13206. [\[CrossRef\]](http://doi.org/10.1080/19443994.2015.1056839)
- <span id="page-21-9"></span>92. Lizotte, R.; Shields, F.; Knight, S.; Bryant, C. Efficiency of a modified backwater wetland in trapping a pesticide mixture. *Ecohydrology* **2009**, *2*, 287–293. [\[CrossRef\]](http://doi.org/10.1002/eco.52)
- 93. Bundschuh, M.; Elsaesser, D.; Stang, C.; Schulz, R. Mitigation of fungicide pollution in detention ponds and vegetated ditches within a vine-growing area in Germany. *Ecol. Eng.* **2016**, *89*, 121–130. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2015.12.015)
- <span id="page-21-6"></span>94. Douglass, J.F.; Radosevich, M.; Tuovinen, O.H. Molecular analysis of atrazine-degrading bacteria and catabolic genes in the water column and sediment of a created wetland in an agricultural/urban watershed. *Ecol. Eng.* **2016**, *83*, 405–412. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2015.06.041)
- <span id="page-21-10"></span>95. Elsaesser, D.; Blankenberg, A.-G.B.; Geist, A.; Mæhlum, T.; Schulz, R. Assessing the influence of vegetation on reduction of pesticide concentration in experimental surface flow constructed wetlands: Application of the toxic units approach. *Ecol. Eng.* **2011**, *37*, 955–962. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2011.02.003)
- 96. Gaullier, C.; Baran, N.; Dousset, S.; Devau, N.; Billet, D.; Kitzinger, G.; Coisy, E. Wetland hydrodynamics and mitigation of pesticides and their metabolites at pilot-scale. *Ecol. Eng.* **2019**, *136*, 185–192. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2019.06.019)
- <span id="page-21-7"></span>97. Hongbin, L.U.; Wang, H.; Shaoyong, L.U.; Jiaxin, L.I.; Wang, T. Response mechanism of typical wetland plants and removal of water pollutants under different levofloxacin concentration. *Ecol. Eng.* **2020**, *158*, 106023. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2020.106023)
- 98. Hussain, S.A.; Prasher, S.O.; Patel, R.M. Removal of ionophoric antibiotics in free water surface constructed wetlands. *Ecol. Eng.* **2012**, *41*, 13–21. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2011.12.006)
- <span id="page-21-11"></span>99. Kohler, E.A.; Poole, V.L.; Reicher, Z.J.; Turco, R.F. Nutrient, metal, and pesticide removal during storm and nonstorm events by a constructed wetland on an urban golf course. *Ecol. Eng.* **2004**, *23*, 285–298. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2004.11.002)
- 100. Krone-Davis, P.; Watson, F.; Los Huertos, M.; Starner, K. Assessing pesticide reduction in constructed wetlands using a tanks-inseries model within a Bayesian framework. *Ecol. Eng.* **2013**, *57*, 342–352. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2013.04.053)
- <span id="page-21-15"></span>101. Lebrun, J.D.; Ayrault, S.; Drouet, A.; Bordier, L.; Fechner, L.C.; Uher, E.; Chaumont, C.; Tournebize, J. Ecodynamics and bioavailability of metal contaminants in a constructed wetland within an agricultural drained catchment. *Ecol. Eng.* **2019**, *136*, 108–117. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2019.06.012)
- <span id="page-21-1"></span>102. Lv, T.; Zhang, Y.; Carvalho, P.N.; Zhang, L.; Button, M.; Arias, C.A.; Weber, K.P.; Brix, H. Microbial community metabolic function in constructed wetland mesocosms treating the pesticides imazalil and tebuconazole. *Ecol. Eng.* **2017**, *98*, 378–387. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2016.07.004)
- <span id="page-21-12"></span>103. Matamoros, V.; Caiola, N.; Rosales, V.; Hernández, O.; Ibáñez, C. The role of rice fields and constructed wetlands as a source and a sink of pesticides and contaminants of emerging concern: Full-scale evaluation. *Ecol. Eng.* **2020**, *156*, 105971. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2020.105971)
- <span id="page-21-2"></span>104. Papaevangelou, V.A.; Gikas, G.D.; Vryzas, Z.; Tsihrintzis, V.A. Treatment of agricultural equipment rinsing water containing a fungicide in pilot-scale horizontal subsurface flow constructed wetlands. *Ecol. Eng.* **2017**, *101*, 193–200. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2017.01.045)
- <span id="page-21-3"></span>105. Romain, V.; Sylvie, D.; David, B. Water residence time and pesticide removal in pilot-scale wetlands. *Ecol. Eng.* **2015**, *85*, 76–84. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2015.09.040)
- <span id="page-21-13"></span>106. Schulz, R.; Peall, S.K.C.; Hugo, C.; Krause, V. Concentration, load and toxicity of spraydrift-borne azinphos-methyl at the inlet and outlet of a constructed wetland. *Ecol. Eng.* **2001**, *18*, 239–245. [\[CrossRef\]](http://doi.org/10.1016/S0925-8574(01)00073-8)
- <span id="page-21-14"></span>107. Tournebize, J.; Passeport, E.; Chaumont, C.; Fesneau, C.; Guenne, A.; Vincent, B. Pesticide de-contamination of surface waters as a wetland ecosystem service in agricultural landscapes. *Ecol. Eng.* **2013**, *56*, 51–59. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2012.06.001)
- 108. Yu, X.; Zhen, S.; Zheng, M.; Ma, X.; Wang, G.; Zou, Y. Herbicide accumulations in the Xingkai lake area and the use of restored wetland for agricultural drainage treatment. *Ecol. Eng.* **2018**, *120*, 260–265. [\[CrossRef\]](http://doi.org/10.1016/j.ecoleng.2018.06.009)
- 109. Alvord, H.; Kadlec, R. Atrazine fate and transport in the Des Plaines Wetlands. *Ecol. Model.* **1996**, *90*, 97–107. [\[CrossRef\]](http://doi.org/10.1016/0304-3800(95)00150-6)
- 110. Milenkovski, S.; Bååth, E.; Lindgren, P.E.; Berglund, O. Toxicity of fungicides to natural bacterial communities in wetland water and sediment measured using leucine incorporation and potential denitrification. *Ecotoxicology* **2010**, *19*, 285–294. [\[CrossRef\]](http://doi.org/10.1007/s10646-009-0411-5) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19768538)
- <span id="page-21-17"></span>111. Dosnon-Olette, R.; Couderchet, M.; Eullaffroy, P. Phytoremediation of fungicides by aquatic macrophytes: Toxicity and removal rate. *Ecotoxicol. Environ. Saf.* **2009**, *72*, 2096–2101. [\[CrossRef\]](http://doi.org/10.1016/j.ecoenv.2009.08.010)
- <span id="page-21-4"></span>112. Guo, X.; Zhong, H.; Li, P.; Zhang, C. Microbial communities responded to tetracyclines and Cu(II) in constructed wetlands microcosms with Myriophyllum aquaticum. *Ecotoxicol. Environ. Saf.* **2020**, *205*, 111362. [\[CrossRef\]](http://doi.org/10.1016/j.ecoenv.2020.111362)
- <span id="page-22-18"></span>113. Lu, H.; Wang, T.; Lu, S.; Liu, H.; Wang, H.; Li, C.; Liu, X.; Guo, X.; Zhao, X.; Liu, F. Performance and bacterial community dynamics of hydroponically grown Iris pseudacorus L. during the treatment of antibiotic-enriched wastewater at low/normal temperature. *Ecotoxicol. Environ. Saf.* **2021**, *213*, 111997. [\[CrossRef\]](http://doi.org/10.1016/j.ecoenv.2021.111997)
- <span id="page-22-0"></span>114. Lu, S.; Zhang, Y.; Liu, X.; Xu, J.; Liu, Y.; Guo, W.; Liu, X.; Chen, J. Effects of sulfamethoxazole on nitrogen removal and molecular ecological network in integrated vertical-flow constructed wetland. *Ecotoxicol. Environ. Saf.* **2021**, *219*, 112292. [\[CrossRef\]](http://doi.org/10.1016/j.ecoenv.2021.112292) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34022628)
- <span id="page-22-13"></span>115. Uddin, M.; Chen, J.; Qiao, X.; Tian, R.; Arafat, Y.; Yang, X. Bacterial community variations in paddy soils induced by application of veterinary antibiotics in plant-soil systems. *Ecotoxicol. Environ. Saf.* **2019**, *167*, 44–53. [\[CrossRef\]](http://doi.org/10.1016/j.ecoenv.2018.09.101)
- 116. Ramos, A.; Whelan, M.J.; Guymer, I.; Villa, R.; Jefferson, B. On the potential of on-line free-surface constructed wetlands for attenuating pesticide losses from agricultural land to surface waters. *Environ. Chem.* **2019**, *16*, 563–576. [\[CrossRef\]](http://doi.org/10.1071/EN19026)
- <span id="page-22-5"></span>117. Cryder, Z.; Wolf, D.; Carlan, C.; Gan, J. Removal of urban-use insecticides in a large-scale constructed wetland. *Environ. Pollut.* **2021**, *268*, 115586. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2020.115586)
- 118. Guo, X.; Mu, Q.; Zhong, H.; Li, P.; Zhang, C.; Wei, D.; Zhao, T. Rapid removal of tetracycline by Myriophyllum aquaticum: Evaluation of the role and mechanisms of adsorption. *Environ. Pollut.* **2019**, *254*, 113101. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2019.113101)
- <span id="page-22-2"></span>119. Lv, T.; Carvalho, P.N.; Casas, M.E.; Bollmann, U.E.; Arias, C.A.; Brix, H.; Bester, K. Enantioselective uptake, translocation and degradation of the chiral pesticides tebuconazole and imazalil by Phragmites australis. *Environ. Pollut.* **2017**, *229*, 362–370. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2017.06.017)
- <span id="page-22-15"></span>120. Lyu, T.; Zhang, L.; Xu, X.; Arias, C.A.; Brix, H.; Carvalho, P.N. Removal of the pesticide tebuconazole in constructed wetlands: Design comparison, influencing factors and modelling. *Environ. Pollut.* **2018**, *233*, 71–80. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2017.10.040)
- <span id="page-22-6"></span>121. Moore, M.T.; Bennett, E.R.; Cooper, C.M.; Smith, S., Jr.; Farris, J.L.; Drouillard, K.G.; Schulz, R. Influence of vegetation in mitigation of methyl parathion runoff. *Environ. Pollut.* **2006**, *142*, 288–294. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2005.10.009)
- <span id="page-22-7"></span>122. Moore, M.T.; Cooper, C.M.; Smith, S.; Cullum, R.F.; Knight, S.S.; Locke, M.A.; Bennett, E.R. Mitigation of two pyrethroid insecticides in a Mississippi Delta constructed wetland. *Environ. Pollut.* **2009**, *157*, 250–256. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2008.07.025) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18789833)
- 123. Moore, M.T.; Rodgers, J.H., Jr.; Cooper, C.M.; Smith, S., Jr. Constructed wetlands for mitigation of atrazine-associated agricultural runoff. *Environ. Pollut.* **2000**, *110*, 393–399. [\[CrossRef\]](http://doi.org/10.1016/S0269-7491(00)00034-8)
- <span id="page-22-1"></span>124. Sherrard, R.M.; Bearr, J.S.; Murray-Gulde, C.L.; Rodgers, J.H., Jr.; Shah, Y.T. Feasibility of constructed wetlands for removing chlorothalonil and chlorpyrifos from aqueous mixtures. *Environ. Pollut.* **2004**, *127*, 385–394. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2003.08.017)
- 125. Stoler, A.; Mattes, B.; Hintz, W.; Jones, D.; Lind, L.; Schuler, M.; Relyea, R. Effects of a common insecticide on wetland communities with varying quality of leaf litter inputs. *Environ. Pollut.* **2017**, *226*, 452–462. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2017.04.019)
- <span id="page-22-8"></span>126. Vallée, R.; Dousset, S.; Schott, F.X.; Pallez, C.; Ortar, A.; Cherrier, R.; Munoz, J.F.; Benoît, M. Do constructed wetlands in grass strips reduce water contamination from drained fields? *Environ. Pollut.* **2015**, *207*, 365–373. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2015.09.027) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26452003)
- <span id="page-22-16"></span>127. Papadopoulos, N.; Zalidis, G. The Use of *Typha latifolia* L. in Constructed Wetland Microcosms for the Remediation of Herbicide Terbuthylazine. *Environ. Process* **2019**, *6*, 985–1003. [\[CrossRef\]](http://doi.org/10.1007/s40710-019-00398-3)
- <span id="page-22-9"></span>128. Gaillard, J.; Thomas, M.; Lazartigues, A.; Bonnefille, B.; Pallez, C.; Dauchy, X.; Feidt, C.; Banas, D. Potential of barrage fish ponds for the mitigation of pesticide pollution in streams. *Environ. Sci. Pollut. Res.* **2016**, *23*, 23–35. [\[CrossRef\]](http://doi.org/10.1007/s11356-015-5378-6)
- <span id="page-22-3"></span>129. Gaullier, C.; Dousset, S.; Billet, D.; Baran, N. Is pesticide sorption by constructed wetland sediments governed by water level and water dynamics? *Environ. Sci. Pollut. Res.* **2018**, *25*, 14324–14335. [\[CrossRef\]](http://doi.org/10.1007/s11356-017-9123-1) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28508331)
- 130. Tang, S.; Liang, J.; Gong, J.; Song, B.; Yang, Z.; Fang, S.; Zhang, P.; Cao, W.; Li, J.; Luo, Y. The effects of biochar/compost for adsorption behaviors of sulfamethoxazole in amended wetland soil. *Environ. Sci. Pollut. Res.* **2021**, *28*, 49289–49301. [\[CrossRef\]](http://doi.org/10.1007/s11356-021-13959-7)
- <span id="page-22-4"></span>131. Tong, X.; Wang, X.; He, X.; Sui, Y.; Shen, J.; Feng, J. Effects of antibiotics on nitrogen uptake of four wetland plant species grown under hydroponic culture. *Environ. Sci. Pollut. Res.* **2019**, *26*, 10621–10630. [\[CrossRef\]](http://doi.org/10.1007/s11356-019-04184-4)
- 132. Ulrich, U.; Lange, J.; Pfannerstill, M.; Loose, L.; Fohrer, N. Hydrological tracers, the herbicide metazachlor and its transformation products in a retention pond during transient flow conditions. *Environ. Sci. Pollut. Res.* **2019**, *26*, 26706–26720. [\[CrossRef\]](http://doi.org/10.1007/s11356-019-05815-6) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31297708)
- <span id="page-22-10"></span>133. Vallée, R.; Dousset, S.; Billet, D. Influence of substrate water saturation on pesticide dissipation in constructed wetlands. *Environ. Sci. Pollut. Res.* **2016**, *23*, 109–119. [\[CrossRef\]](http://doi.org/10.1007/s11356-015-4367-0) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25813638)
- <span id="page-22-17"></span>134. Vallée, R.; Dousset, S.; Billet, D.; Benoit, M. Sorption of selected pesticides on soils, sediment and straw from a constructed agricultural drainage ditch or pond. *Environ. Sci. Pollut. Res.* **2014**, *21*, 4895–4905. [\[CrossRef\]](http://doi.org/10.1007/s11356-013-1840-5) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23784054)
- <span id="page-22-11"></span>135. Budd, R.; O'Geen, A.; Goh, K.S.; Bondarenko, S.; Gan, J. Efficacy of constructed wetlands in pesticide removal from tailwaters in the central valley, California. *Environ. Sci. Technol.* **2009**, *43*, 2925–2930. [\[CrossRef\]](http://doi.org/10.1021/es802958q) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19475972)
- 136. Carley, L.N.; Panchagavi, R.; Song, X.; Davenport, S.; Bergemann, C.M.; McCumber, A.W.; Gunsch, C.K.; Simonin, M. Long-Term Effects of Copper Nanopesticides on Soil and Sediment Community Diversity in Two Outdoor Mesocosm Experiments. *Environ. Sci. Technol.* **2020**, *54*, 8878–8889. [\[CrossRef\]](http://doi.org/10.1021/acs.est.0c00510) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32543178)
- <span id="page-22-14"></span>137. Jeppe, K.; Kellar, C.; Marshall, S.; Colombo, V.; Sinclair, G.; Pettigrove, V. Bifenthrin Causes Toxicity in Urban Stormwater Wetlands: Field and Laboratory Assessment Using Austrochiltonia (Amphipoda). *Environ. Sci. Technol.* **2017**, *51*, 7254–7262. [\[CrossRef\]](http://doi.org/10.1021/acs.est.7b01472) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28493716)
- 138. Kuechle, K.J.; Webb, E.B.; Mengel, D.; Main, A.R.; Webb, L. Factors Influencing Neonicotinoid Insecticide Concentrations in Floodplain Wetland Sediments across Missouri. *Environ. Sci. Technol.* **2019**, *53*, 10591–10600. [\[CrossRef\]](http://doi.org/10.1021/acs.est.9b01799)
- <span id="page-22-12"></span>139. Maillard, E.; Imfeld, G. Pesticide Mass Budget in a Stormwater Wetland. *Environ. Sci. Technol.* **2014**, *48*, 8603–8611. [\[CrossRef\]](http://doi.org/10.1021/es500586x)
- 140. Main, A.; Michel, N.; Headley, J.; Peru, K.; Morrissey, C. Ecological and Landscape Drivers of Neonicotinoid Insecticide Detections and Concentrations in Canada's Prairie Wetlands. *Environ. Sci. Technol.* **2015**, *49*, 8367–8376. [\[CrossRef\]](http://doi.org/10.1021/acs.est.5b01287)
- <span id="page-23-8"></span>141. Schulz, R.; Hahn, C.; Bennett, E.R.; Dabrowski, J.M.; Thiere, G.; Peall, S.K.C. Fate and effects of azinphos-methyl in a flow-through wetland in South Africa. *Environ. Sci. Technol.* **2003**, *37*, 2139–2144. [\[CrossRef\]](http://doi.org/10.1021/es026029f)
- <span id="page-23-9"></span>142. Schulz, R.; Peall, S.K.C. Effectiveness of a constructed wetland for retention of nonpoint-source pesticide pollution in the Lourens River catchment, South Africa. *Environ. Sci. Technol.* **2001**, *35*, 422–426. [\[CrossRef\]](http://doi.org/10.1021/es0001198)
- <span id="page-23-5"></span>143. Bouldin, J.L.; Farris, J.L.; Moore, M.T.; Smith, S., Jr.; Stephens, W.W.; Cooper, C.M. Evaluated fate and effects of atrazine and lambda-cyhalothrin in vegetated and unvegetated microcosms. *Environ. Toxicol.* **2005**, *20*, 487–498. [\[CrossRef\]](http://doi.org/10.1002/tox.20137) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16161102)
- <span id="page-23-0"></span>144. Milam, C.D.; Bouldin, J.L.; Farris, J.L.; Schulz, R.; Moore, M.T.; Bennett, E.R.; Cooper, C.M.; Smith, S., Jr. Evaluating acute toxicity of methyl parathion application in constructed wetland mesocosms. *Environ. Toxicol.* **2004**, *19*, 471–479. [\[CrossRef\]](http://doi.org/10.1002/tox.20052)
- <span id="page-23-17"></span>145. Passeport, E.; Benoit, P.; Bergheaud, V.; Coquet, Y.; Tournebize, J. Selected pesticides adsorption and desorption in substrates from artificial wetland and forest buffer. *Environ. Toxicol. Chem.* **2011**, *30*, 1669–1676. [\[CrossRef\]](http://doi.org/10.1002/etc.554)
- <span id="page-23-6"></span>146. Thomas, K.; Hand, L. Assessing the potential for algae and macrophytes to degrade crop protection products in aquatic ecosystems. *Environ. Toxicol. Chem.* **2011**, *30*, 622–631. [\[CrossRef\]](http://doi.org/10.1002/etc.412)
- <span id="page-23-10"></span>147. Pappalardo, S.; Otto, S.; Gasparini, V.; Zanin, G.; Borin, M. Mitigation of herbicide runoff as an ecosystem service from a constructed surface flow wetland. *Hydrobiologia* **2016**, *774*, 193–202. [\[CrossRef\]](http://doi.org/10.1007/s10750-015-2375-1)
- <span id="page-23-15"></span>148. Greiwe, J.; Olsson, O.; Kummerer, K.; Lange, J. Pesticide peak concentration reduction in a small vegetated treatment system controlled by chemograph shape. *Hydrol. Earth Syst. Sci.* **2021**, *25*, 497–509. [\[CrossRef\]](http://doi.org/10.5194/hess-25-497-2021)
- 149. Blankenberg, A.-G.B.; Braskerud, B.; Haarstad, K. Pesticide retention in two small constructed wetlands: Treating non-point source pollution from agriculture runoff. *Int. J. Environ. Anal. Chem.* **2006**, *86*, 225–231. [\[CrossRef\]](http://doi.org/10.1080/03067310500247470)
- <span id="page-23-11"></span>150. Maillard, E.; Payraudeau, S.; Ortiz, F.; Imfeld, G. Removal of dissolved pesticide mixtures by a stormwater wetland receiving runoff from a vineyard catchment: An inter-annual comparison. *Int. J. Environ. Anal. Chem.* **2011**, *92*, 979–994. [\[CrossRef\]](http://doi.org/10.1080/03067319.2011.609935)
- <span id="page-23-1"></span>151. Turcios, A.E.; Hielscher, M.; Duarte, B.; Fonseca, V.F.; Caçador, I.; Papenbrock, J. Screening of emerging pollutants (Eps) in estuarine water and phytoremediation capacity of tripolium pannonicum under controlled conditions. *Int. J. Environ. Res. Public Health* **2021**, *18*, 943. [\[CrossRef\]](http://doi.org/10.3390/ijerph18030943) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33499038)
- 152. Barchanska, H.; Plonka, J.; Jaros, A.; Ostrowska, A. Potential application of Pistia stratiotes for the phytoremediation of mesotrione and its degradation products from water. *Int. J. Phytoremed.* **2019**, *21*, 1090–1097. [\[CrossRef\]](http://doi.org/10.1080/15226514.2019.1606780) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31084361)
- 153. Cheng, S.; Xiao, J.; Xiao, H.; Zhang, L.; Wu, Z. Phytoremediation of triazophos by Canna indica Linn. in a hydroponic system. *Int. J. Phytoremed.* **2007**, *9*, 453–463. [\[CrossRef\]](http://doi.org/10.1080/15226510701709531) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18246772)
- <span id="page-23-2"></span>154. McKnight, A.M.; Gannon, T.W.; Yelverton, F. Phytoremediation of azoxystrobin and imidacloprid by wetland plant species Juncus effusus, Pontederia cordata and Sagittaria latifolia. *Int. J. Phytoremed.* **2021**, 1–9. [\[CrossRef\]](http://doi.org/10.1080/15226514.2021.1932726)
- 155. Mercado-Borrayo, B.; Heydrich, S.; Perez, I.; Quiroz, M.; Hill, C. Organophosphorus and Organochlorine Pesticides Bioaccumulation by Eichhornia crassipes in Irrigation Canals in an Urban Agricultural System. *Int. J. Phytoremed.* **2015**, *17*, 701–708. [\[CrossRef\]](http://doi.org/10.1080/15226514.2014.964841) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25976884)
- <span id="page-23-12"></span>156. Gaillard, J.; Thomas, M.; Iuretig, A.; Pallez, C.; Feidt, C.; Dauchy, X.; Banas, D. Barrage fishponds: Reduction of pesticide concentration peaks and associated risk of adverse ecological effects in headwater streams. *J. Environ. Manag.* **2016**, *169*, 261–271. [\[CrossRef\]](http://doi.org/10.1016/j.jenvman.2015.12.035)
- 157. Gaullier, C.; Dousset, S.; Baran, N.; Kitzinger, G.; Coureau, C. Influence of hydrodynamics on the water pathway and spatial distribution of pesticide and metabolite concentrations in constructed wetlands. *J. Environ. Manag.* **2020**, *270*. [\[CrossRef\]](http://doi.org/10.1016/j.jenvman.2020.110690) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32721282)
- <span id="page-23-16"></span>158. Knuteson, S.; Whitwell, T.; Klaine, S. Influence of plant age and size on simazine toxicity and uptake. *J. Environ. Qual.* **2002**, *31*, 2096–2103. [\[CrossRef\]](http://doi.org/10.2134/jeq2002.2096) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12469861)
- <span id="page-23-13"></span>159. McMaine, J.T.; Vogel, J.R.; Belden, J.B.; Schnelle, M.A.; Morrison, S.A.; Brown, G.O. Field studies of pollutant removal from nursery and greenhouse runoff by constructed wetlands. *J. Environ. Qual.* **2020**, *49*, 106–118. [\[CrossRef\]](http://doi.org/10.1002/jeq2.20024)
- <span id="page-23-14"></span>160. Moore, M.T.; Locke, M.A.; Cullum, R.F. Expanding wetland mitigation: Can rice fields remediate pesticides in agricultural runoff? *J. Environ. Qual.* **2018**, *47*, 1564–1571. [\[CrossRef\]](http://doi.org/10.2134/jeq2018.04.0154)
- <span id="page-23-3"></span>161. Stearman, G.K.; George, D.B.; Carlson, K.; Lansford, S. Pesticide removal from container nursery runoff in constructed wetland cells. *J. Environ. Qual.* **2003**, *32*, 1548–1556. [\[CrossRef\]](http://doi.org/10.2134/jeq2003.1548)
- 162. Sura, S.; Waiser, M.; Tumber, V.; Lawrence, J.; Cessna, A.; Glozier, N. Effects of Glyphosate and Two Herbicide Mixtures on Microbial Communities in Prairie Wetland Ecosystems: A Mesocosm Approach. *J. Environ. Qual.* **2012**, *41*, 732–743. [\[CrossRef\]](http://doi.org/10.2134/jeq2011.0376)
- <span id="page-23-4"></span>163. Casas-Zapata, J.C.; Ríos, K.; Florville-Alejandre, T.R.; Morató, J.; Peñuela, G. Influence of chlorothalonil on the removal of organic matter in horizontal subsurface flow constructed wetlands. *J. Environ. Sci. Health Part B Pestic. Food Contam. Agric. Wastes* **2013**, *48*, 122–132. [\[CrossRef\]](http://doi.org/10.1080/03601234.2013.726909)
- <span id="page-23-7"></span>164. Friesen-Pankratz, B.; Doebel, C.; Farenhorst, A.; Goldsborough, L.G. Interactions between algae (Selenastrum capricornutum) and pesticides: Implications for managing constructed wetlands for pesticide removal. *J. Environ. Sci. Health Part B Pestic. Food Contam. Agric. Wastes* **2003**, *38*, 147–155. [\[CrossRef\]](http://doi.org/10.1081/PFC-120018445) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12617553)
- 165. Verma, B.; Headley, J.; Robarts, R. Behaviour and fate of tetracycline in river and wetland waters on the Canadian Northern Great Plains. *J. Environ. Sci. Health Part A* **2007**, *42*, 109–117. [\[CrossRef\]](http://doi.org/10.1080/10934520601011163)
- 166. Munoz, A.; Trevisan, M.; Capri, E. Sorption and photodegradation of chlorpyrifos on riparian and aquatic macrophytes. *J. Environ. Sci. Health Part B* **2009**, *44*, 7–12. [\[CrossRef\]](http://doi.org/10.1080/03601230802519496)
- 167. Moore, M.T.; Locke, M.A.; Kröger, R. Mitigation of atrazine, S-metolachlor, and diazinon using common emergent aquatic vegetation. *J. Environ. Sci.* **2017**, *56*, 114–121. [\[CrossRef\]](http://doi.org/10.1016/j.jes.2016.09.009)
- <span id="page-24-9"></span>168. Matamoros, V.; Rodriguez, Y. Batch vs continuous-feeding operational mode for the removal of pesticides from agricultural run-off by microalgae systems: A laboratory scale study. *J. Hazard. Mater.* **2016**, *309*, 126–132. [\[CrossRef\]](http://doi.org/10.1016/j.jhazmat.2016.01.080)
- <span id="page-24-0"></span>169. Ohore, O.; Zhang, S.; Guo, S.; Manirakiza, B.; Addo, F.; Zhang, W. The fate of tetracycline in vegetated mesocosmic wetlands and its impact on the water quality and epiphytic microbes. *J. Hazard. Mater.* **2021**, *417*, 126148. [\[CrossRef\]](http://doi.org/10.1016/j.jhazmat.2021.126148)
- <span id="page-24-20"></span>170. Tang, X.; Liu, H.; Naïla, R.S.L.; Dai, Y.; Zhang, X.; Tam, N.F.Y.; Xiong, C.; Yang, Y. Irrigation using hybrid constructed wetland treated domestic sewage: Uptake of phthalic acid esters and antibiotics by Ipomoea aquatica forssk. *J. Hazard. Mater.* **2021**, *405*, 124025. [\[CrossRef\]](http://doi.org/10.1016/j.jhazmat.2020.124025) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33129603)
- <span id="page-24-1"></span>171. Yu, X.; Zhu, H.; Yan, B.; Xu, Y.; Banuelos, G.; Shutes, B.; Wen, H.; Cheng, R. Removal of chlorpyrifos and its hydrolytic metabolite 3,5,6-trichloro-2-pyridinol in constructed wetland mesocosms under soda saline-alkaline conditions: Effectiveness and influencing factors. *J. Hazard. Mater.* **2019**, *373*, 67–74. [\[CrossRef\]](http://doi.org/10.1016/j.jhazmat.2019.03.052)
- 172. Challis, J.K.; Carlson, J.C.; Friesen, K.J.; Hanson, M.L.; Wong, C.S. Aquatic photochemistry of the sulfonamide antibiotic sulfapyridine. *J. Photochem. Photobiol. A Chem.* **2013**, *262*, 14–21. [\[CrossRef\]](http://doi.org/10.1016/j.jphotochem.2013.04.009)
- <span id="page-24-13"></span>173. Bois, P.; Huguenot, D.; Norini, M.P.; Haque, M.F.U.; Vuilleumier, S.; Lebeau, T. Herbicide degradation and copper complexation by bacterial mixed cultures from a vineyard stormwater basin. *J. Soils Sed.* **2011**, *11*, 860–873. [\[CrossRef\]](http://doi.org/10.1007/s11368-011-0354-3)
- <span id="page-24-14"></span>174. Moore, M.T.; Kröger, R.; Cooper, C.M.; Cullum, R.F.; Smith, S., Jr.; Locke, M.A. Diazinon reduction and partitioning between water, sediment and vegetation in stormwater runoff mitigation through rice fields. *Pest Manag. Sci.* **2009**, *65*, 1182–1188. [\[CrossRef\]](http://doi.org/10.1002/ps.1805) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19548297)
- <span id="page-24-2"></span>175. Wang, C.; Liu, B.; Xu, D.; Zhang, D.; He, F.; Zhou, Q.; Wu, Z. Mitigation of wastewater-borne chlorpyrifos in constructed wetlands: The role of vegetation on partitioning. *Pol. J. Environ. Stud.* **2017**, *26*, 347–354. [\[CrossRef\]](http://doi.org/10.15244/pjoes/64375)
- 176. Wang, C.; Liu, B.; Xu, D.; Zhang, L.; He, F.; Zhou, Q.; Wu, Z. Mitigation of wastewater-borne chorpyrifos in constructed wetlands: An ecological suitability assessment by macrophyte and microbial responses. *Pol. J. Environ. Stud.* **2017**, *26*, 1279–1287. [\[CrossRef\]](http://doi.org/10.15244/pjoes/67656)
- <span id="page-24-10"></span>177. Topal, M.; Obek, E.; Senel, G.; Topal, E. Phytoremediation of Tetracycline and Degradation Products from Aqueous Solutions. *Pollution* **2018**, *4*, 471–480. [\[CrossRef\]](http://doi.org/10.22059/poll.2018.248824.359)
- <span id="page-24-3"></span>178. De Souza, T.D.; Borges, A.C.; de Matos, A.T.; Mounteer, A.H.; de Queiroz, M.E.L.R. Removal of chlorpyrifos insecticide in constructed wetlands with different plant species. *Rev. Bras. Eng. Agric. Ambient.* **2017**, *21*, 878–883. [\[CrossRef\]](http://doi.org/10.1590/1807-1929/agriambi.v21n12p878-883)
- <span id="page-24-21"></span>179. Sha, N.-Q.; Wang, G.-H.; Li, Y.-H.; Bai, S.-Y. Removal of abamectin and conventional pollutants in vertical flow constructed wetlands with Fe-modified biochar. *RSC Adv.* **2020**, *10*, 44171–44182. [\[CrossRef\]](http://doi.org/10.1039/D0RA08265A)
- <span id="page-24-4"></span>180. Agudelo, C.R.M.; Jaramillo, M.L.; Peñuela, G. Comparison of the removal of chlorpyrifos and dissolved organic carbon in horizontal sub-surface and surface flow wetlands. *Sci. Total Environ.* **2012**, *431*, 271–277. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2012.05.045) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22687437)
- <span id="page-24-11"></span>181. Butkovskyi, A.; Jing, Y.; Bergheim, H.; Lazar, D.; Gulyaeva, K.; Odenmarck, S.R.; Norli, H.R.; Nowak, K.M.; Miltner, A.; Kästner, M.; et al. Retention and distribution of pesticides in planted filter microcosms designed for treatment of agricultural surface runoff. *Sci. Total Environ.* **2021**, *778*, 146114. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2021.146114)
- <span id="page-24-5"></span>182. Chen, Z.; Chen, Y.; Vymazal, J.; Kule, L.; Koželuh, M. Dynamics of chloroacetanilide herbicides in various types of mesocosm wetlands. *Sci. Total Environ.* **2017**, *577*, 386–394. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2016.10.216)
- <span id="page-24-6"></span>183. Elsayed, O.F.; Maillard, E.; Vuilleumier, S.; Imfeld, G. Bacterial communities in batch and continuous-flow wetlands treating the herbicide S-metolachlor. *Sci. Total Environ.* **2014**, *499*, 327–335. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2014.08.048) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25201820)
- <span id="page-24-15"></span>184. Lizotte, R.; Shields, F.; Murdock, J.; Kroger, R.; Knight, S. Mitigating agrichemicals from an artificial runoff event using a managed riverine wetland. *Sci. Total Environ.* **2012**, *427*, 427–428. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2012.04.025) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22560749)
- <span id="page-24-7"></span>185. Lobson, C.; Luong, K.; Seburn, D.; White, M.; Hann, B.; Prosser, R.; Wong, C.; Hanson, M. Fate of thiamethoxam in mesocosms and response of the zooplankton community. *Sci. Total Environ.* **2018**, *637*, 637–638. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2018.05.087) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29801208)
- <span id="page-24-16"></span>186. Maillard, E.; Payraudeau, S.; Faivre, E.; Gregoire, C.; Gangloff, S.; Imfeld, G. Removal of pesticide mixtures in a stormwater wetland collecting runoff from a vineyard catchment. *Sci. Total Environ.* **2011**, *409*, 2317–2324. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2011.01.057) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21353289)
- <span id="page-24-17"></span>187. Main, A.; Fehr, J.; Liber, K.; Headley, J.; Peru, K.; Morrissey, C. Reduction of neonicotinoid insecticide residues in Prairie wetlands by common wetland plants. *Sci. Total Environ.* **2017**, *579*, 1193–1202. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2016.11.102)
- <span id="page-24-12"></span>188. Sahin, C.; Karpuzcu, M.E. Mitigation of organophosphate pesticide pollution in agricultural watersheds. *Sci. Total Environ.* **2020**, *710*, 136261. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2019.136261)
- <span id="page-24-18"></span>189. Solis, M.; Cappelletti, N.; Bonetto, C.; Franco, M.; Fanelli, S.; Amalvy, J.; Mugni, H. Attenuation of insecticide impact by a small wetland in a stream draining a horticultural basin in Argentina. *Sci. Total Environ.* **2021**, *785*, 147317. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2021.147317)
- 190. Sura, S.; Waiser, M.; Tumber, V.; Farenhorst, A. Effects of herbicide mixture on microbial communities in prairie wetland ecosystems: A whole wetland approach. *Sci. Total Environ.* **2012**, *435*, 435–436. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2012.07.003)
- <span id="page-24-19"></span>191. Sura, S.; Waiser, M.; Tumber, V.; Raina-Fulton, R.; Cessna, A. Effects of a herbicide mixture on primary and bacterial productivity in four prairie wetlands with varying salinities: An enclosure approach. *Sci. Total Environ.* **2015**, *512*, 512–513. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2015.01.064)
- <span id="page-24-8"></span>192. Tang, X.; Yang, Y.; Tao, R.; Chen, P.; Dai, Y.; Jin, C.; Feng, X. Fate of mixed pesticides in an integrated recirculating constructed wetland (IRCW). *Sci. Total Environ.* **2016**, *571*, 935–942. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2016.07.079)
- <span id="page-25-3"></span>193. Wu, J.; Feng, Y.; Dai, Y.; Cui, N.; Anderson, B.; Cheng, S. Biological mechanisms associated with triazophos (TAP) removal by horizontal subsurface flow constructed wetlands (HSFCW). *Sci. Total Environ.* **2016**, *553*, 13–19. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2016.02.067)
- <span id="page-25-10"></span>194. Zhu, H.; Yu, X.; Xu, Y.; Yan, B.; Banuelos, G.; Shutes, B.; Wen, Z. Removal of chlorpyrifos and its hydrolytic metabolite in microcosm-scale constructed wetlands under soda saline-alkaline condition: Mass balance and intensification strategies. *Sci. Total Environ.* **2021**, *777*, 145956. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2021.145956) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33676222)
- <span id="page-25-4"></span>195. Wu, J.; Li, Z.; Wu, L.; Zhong, F.; Cui, N.; Dai, Y.; Cheng, S. Triazophos (TAP) removal in horizontal subsurface flow constructed wetlands (HSCWs) and its accumulation in plants and substrates. *Sci. Rep.* **2017**, *7*, 5468. [\[CrossRef\]](http://doi.org/10.1038/s41598-017-05874-0)
- <span id="page-25-13"></span>196. D'Angelo, E.; Reddy, K. Effect of aerobic and anaerobic conditions on chlorophenol sorption in wetland soils. *Soil Sci. Soc. Am. J.* **2003**, *67*, 787–794. [\[CrossRef\]](http://doi.org/10.2136/sssaj2003.7870)
- <span id="page-25-20"></span>197. Passeport, E.; Tournebize, J.; Jankowfsky, S.; Prömse, B.; Chaumont, C.; Coquet, Y.; Lange, J. Artificial wetland and forest buffer zone: Hydraulic and tracer characterization. *Vadose Zone J.* **2010**, *9*, 73–84. [\[CrossRef\]](http://doi.org/10.2136/vzj2008.0164)
- <span id="page-25-19"></span>198. Lizotte, R.; Shields, F.; Testa, S. Effects of a Simulated Agricultural Runoff Event on Sediment Toxicity in a Managed Backwater Wetland. *Water Air Soil Pollut.* **2012**, *223*, 5375–5389. [\[CrossRef\]](http://doi.org/10.1007/s11270-012-1287-1)
- 199. Mauffrey, F.; Baccara, P.; Gruffaz, C.; Vuilleumier, S.; Imfeld, G. Bacterial Community Composition and Genes for Herbicide Degradation in a Stormwater Wetland Collecting Herbicide Runoff. *Water Air Soil Pollut.* **2017**, *228*, 452. [\[CrossRef\]](http://doi.org/10.1007/s11270-017-3625-9)
- <span id="page-25-11"></span>200. Bois, P.; Huguenot, D.; Jezequel, K.; Lollier, M.; Cornu, J.; Lebeau, T. Herbicide mitigation in microcosms simulating stormwater basins subject to polluted water inputs. *Water Res.* **2013**, *47*, 1123–1135. [\[CrossRef\]](http://doi.org/10.1016/j.watres.2012.11.029) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23246667)
- <span id="page-25-5"></span>201. Lv, T.; Carvalho, P.N.; Zhang, L.; Zhang, Y.; Button, M.; Arias, C.A.; Weber, K.P.; Brix, H. Functionality of microbial communities in constructed wetlands used for pesticide remediation: Influence of system design and sampling strategy. *Water Res.* **2017**, *110*, 241–251. [\[CrossRef\]](http://doi.org/10.1016/j.watres.2016.12.021) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28011364)
- 202. Lv, T.; Zhang, Y.; Zhang, L.; Carvalho, P.N.; Arias, C.A.; Brix, H. Removal of the pesticides imazalil and tebuconazole in saturated constructed wetland mesocosms. *Water Res.* **2016**, *91*, 126–136. [\[CrossRef\]](http://doi.org/10.1016/j.watres.2016.01.007)
- <span id="page-25-6"></span>203. McKinlay, R.G.; Kasperek, K. Observations on decontamination of herbicide-polluted water by marsh plant systems. *Water Res.* **1999**, *33*, 505–511. [\[CrossRef\]](http://doi.org/10.1016/S0043-1354(98)00244-9)
- 204. Borges, A.C.; Do Carmo Calijuri, M.; De Matos, A.T.; De Queiroz, M.E.L.R. Horizontal subsurface flow constructed wetlands for mitigation of ametryn-contaminated water. *Water SA* **2009**, *35*, 441–446. [\[CrossRef\]](http://doi.org/10.4314/wsa.v35i4.76804)
- <span id="page-25-18"></span>205. Braskerud, B.C.; Haarstad, K. Screening the retention of thirteen pesticides in a small constructed wetland. *Water Sci. Technol.* **2003**, *48*, 267–274. [\[CrossRef\]](http://doi.org/10.2166/wst.2003.0332)
- <span id="page-25-14"></span>206. Haarstad, K.; Braskerud, B.C. Pesticide retention in the watershed and in a small constructed wetland treating diffuse pollution. *Water Sci. Technol.* **2005**, *51*, 143–150. [\[CrossRef\]](http://doi.org/10.2166/wst.2005.0585)
- <span id="page-25-12"></span>207. Roseth, R.; Haarstad, K. Pesticide runoff from greenhouse production. *Water Sci. Technol.* **2010**, *61*, 1373–1381. [\[CrossRef\]](http://doi.org/10.2166/wst.2010.040)
- <span id="page-25-15"></span>208. Sudo, M.; Nishino, M.; Okubo, T. Decrease in herbicide concentrations and affected factors in lagoons located around Lake Biwa. *Water Sci. Technol.* **2006**, *53*, 131–138. [\[CrossRef\]](http://doi.org/10.2166/wst.2006.046) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16594331)
- <span id="page-25-7"></span>209. Lizotte, R.E., Jr.; Moore, M.T.; Locke, M.A.; Kröger, R. Effects of vegetation in mitigating the toxicity of pesticide mixtures in sediments of a wetland mesocosm. *Water Air Soil Pollut.* **2011**, *220*, 69–79. [\[CrossRef\]](http://doi.org/10.1007/s11270-010-0735-z)
- <span id="page-25-8"></span>210. Mahabali, S.; Spanoghe, P. Mitigation of two insecticides by wetland plants: Feasibility study for the treatment of agricultural runoff in suriname (South America). *Water Air Soil Pollut.* **2014**, *225*, 1771. [\[CrossRef\]](http://doi.org/10.1007/s11270-013-1771-2)
- 211. Moore, M.T.; Cooper, C.M.; Smith, S., Jr.; Cullum, R.F.; Knight, S.S.; Locke, M.A.; Bennett, E.R. Diazinon mitigation in constructed wetlands: Influence of vegetation. *Water Air Soil Pollut.* **2007**, *184*, 313–321. [\[CrossRef\]](http://doi.org/10.1007/s11270-007-9418-9)
- <span id="page-25-16"></span>212. Rose, M.T.; Crossan, A.N.; Kennedy, I.R. Dissipation of cotton pesticides from runoff water in glasshouse columns. *Water Air Soil Pollut.* **2007**, *182*, 207–218. [\[CrossRef\]](http://doi.org/10.1007/s11270-006-9333-5)
- <span id="page-25-17"></span>213. Birch, G.F.; Matthai, C.; Fazeli, M.S.; Suh, J. Efficiency of a constructed wetland in removing contaminants from stormwater. *Wetlands* **2004**, *24*, 459–466. [\[CrossRef\]](http://doi.org/10.1672/0277-5212(2004)024[0459:EOACWI]2.0.CO;2)
- 214. Henry, B.L.; Wesner, J.S.; Kerby, J.L. Cross-Ecosystem Effects of Agricultural Tile Drainage, Surface Runoff, and Selenium in the Prairie Pothole Region. *Wetlands* **2020**, *40*, 527–538. [\[CrossRef\]](http://doi.org/10.1007/s13157-019-01194-3)
- 215. Hussain, S.A.; Prasher, S.O. Understanding the sorption of ionophoric pharmaceuticals in a treatment wetland. *Wetlands* **2011**, *31*, 563–571. [\[CrossRef\]](http://doi.org/10.1007/s13157-011-0171-x)
- 216. Shan, A.; Wang, W.; Kang, K.J.; Hou, D.; Luo, J.; Wang, G.; Pan, M.; Feng, Y.; He, Z.; Yang, X. The Removal of Antibiotics in Relation to a Microbial Community in an Integrated Constructed Wetland for Tail Water Decontamination. *Wetlands* **2020**, *40*, 993–1004. [\[CrossRef\]](http://doi.org/10.1007/s13157-019-01262-8)
- <span id="page-25-9"></span>217. Schulz, R.; Moore, M.T.; Bennett, E.R.; Farris, J.L.; Smith, S., Jr.; Cooper, C.M. Methyl parathion toxicity in vegetated and nonvegetated wetland mesocosms. *Environ. Toxicol. Chem.* **2003**, *22*, 1262–1268. [\[CrossRef\]](http://doi.org/10.1002/etc.5620220611) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12785582)
- <span id="page-25-0"></span>218. Berberidou, C.; Kitsiou, V.; Lambropoulou, D.A.; Antoniadis, A.; Ntonou, E.; Zalidis, G.C.; Poulios, I. Evaluation of an alternative method for wastewater treatment containing pesticides using solar photocatalytic oxidation and constructed wetlands. *J. Environ. Manag.* **2017**, *195*, 133–139. [\[CrossRef\]](http://doi.org/10.1016/j.jenvman.2016.06.010)
- <span id="page-25-1"></span>219. Donley, N. The USA lags behind other agricultural nations in banning harmful pesticides. *Environ. Health* **2019**, *18*, 44. [\[CrossRef\]](http://doi.org/10.1186/s12940-019-0488-0)
- <span id="page-25-2"></span>220. Survey, U.S.G. Estimated Annual Agricultural Pesticide Use. Available online: [https://water.usgs.gov/nawqa/pnsp/usage/](https://water.usgs.gov/nawqa/pnsp/usage/maps/compound_listing.php) [maps/compound\\_listing.php](https://water.usgs.gov/nawqa/pnsp/usage/maps/compound_listing.php) (accessed on 24 September 2021).
- <span id="page-26-0"></span>221. Ahn, C.; Mitsch, W.J. Scaling considerations of mesocosm wetlands in simulating large created freshwater marshes. *Ecol. Eng.* **2020**, *18*, 327–342. [\[CrossRef\]](http://doi.org/10.1016/S0925-8574(01)00092-1)
- <span id="page-26-1"></span>222. Nally, R.M. Scaling artefacts in confinement experiments: A simulation model. *Ecol. Model.* **1997**, *99*, 229–245. [\[CrossRef\]](http://doi.org/10.1016/S0304-3800(97)01958-3)
- <span id="page-26-2"></span>223. Pilson, M.; Nixon, S. Marine Microcosms in Ecological Research. In *Department of Energy Symposium Series*; U.S. Environmental Protection Agency: Washington, DC, USA, 1980; EPA/600/D-82/218 (NTIS PB82181900).
- <span id="page-26-3"></span>224. Messer, T.; Burchell, M.; Bírgand, F. Comparison of Four Nitrate Removal Kinetic Models in Two Distinct Wetland Restoration Mesocosm Systems. *Water* **2017**, *9*, 517. [\[CrossRef\]](http://doi.org/10.3390/w9070517)
- <span id="page-26-4"></span>225. Parde, D.; Patwa, A.; Shukla, A.; Vijay, R.; Killedar, D.J.; Kumar, R. A review of constructed wetland on type, treatment and technology of wastewater. *Environ. Technol. Innov.* **2021**, *21*, 101261. [\[CrossRef\]](http://doi.org/10.1016/j.eti.2020.101261)
- <span id="page-26-5"></span>226. Vymazal, J. Constructed Wetlands for Wastewater Treatment. *Water* **2010**, *2*, 530–549. [\[CrossRef\]](http://doi.org/10.3390/w2030530)
- <span id="page-26-6"></span>227. Lawler, S. Rice fields as temporary wetlands. *Israel J. Zool.* **2001**, *47*, 513–528. [\[CrossRef\]](http://doi.org/10.1560/X7K3-9JG8-MH2J-XGX1)
- <span id="page-26-7"></span>228. Agency, E.P. Atrazine. Available online: <Epa.gov/ingredients-used-pesticide-products/atrazine> (accessed on 12 October 2021).
- <span id="page-26-8"></span>229. Wolf, K.L.; Ahn, C.; Noe, G.B. Development of soil properties and nitrogen cycling in created wetlands. *Wetlands* **2011**, *31*, 699–712. [\[CrossRef\]](http://doi.org/10.1007/s13157-011-0185-4)
- <span id="page-26-9"></span>230. Bruland, G.L.; Richardson, C.J. Spatial variability of soil properties in created, restored, and paired natural wetlands. *Soil Sci. Soc. Am. J.* **2005**, *69*, 273–284. [\[CrossRef\]](http://doi.org/10.2136/sssaj2005.0273a)
- <span id="page-26-10"></span>231. Weber, K. Microbial Community Assessment in Wetlands for Water Pollution Control: Past, Present, and Future Outlook. *Water* **2016**, *8*, 503. [\[CrossRef\]](http://doi.org/10.3390/w8110503)