University of Kentucky UKnowledge

**Chemistry Faculty Publications** 

Chemistry

10-22-2020

# Oxidation of Phenolic Aldehydes by Ozone and Hydroxyl Radicals at the Air-Water Interface

Md Sohel Rana University of Kentucky, mdsohelrana@uky.edu

Marcelo I. Guzman University of Kentucky, marcelo.guzman@uky.edu

Follow this and additional works at: https://uknowledge.uky.edu/chemistry\_facpub

Part of the Physical Chemistry Commons

Right click to open a feedback form in a new tab to let us know how this document benefits you.

# **Repository Citation**

Rana, Md Sohel and Guzman, Marcelo I., "Oxidation of Phenolic Aldehydes by Ozone and Hydroxyl Radicals at the Air-Water Interface" (2020). *Chemistry Faculty Publications*. 185. https://uknowledge.uky.edu/chemistry\_facpub/185

This Article is brought to you for free and open access by the Chemistry at UKnowledge. It has been accepted for inclusion in Chemistry Faculty Publications by an authorized administrator of UKnowledge. For more information, please contact UKnowledge@lsv.uky.edu.

# Oxidation of Phenolic Aldehydes by Ozone and Hydroxyl Radicals at the Air-Water Interface

Digital Object Identifier (DOI) https://doi.org/10.1021/acs.jpca.0c05944

# **Notes/Citation Information**

Published in The Journal of Physical Chemistry. A, v. 124, issue 42.

Copyright © 2020 American Chemical Society ACS AuthorChoice with CC-BY-NC-ND license

This article is available at UKnowledge: https://uknowledge.uky.edu/chemistry\_facpub/185



pubs.acs.org/JPCA

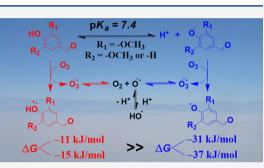
Article

# Oxidation of Phenolic Aldehydes by Ozone and Hydroxyl Radicals at the Air–Water Interface

Md. Sohel Rana and Marcelo I. Guzman\*

Cite This: J. F	Phys. Chem. A 2020, 124, 8822–8833	Read Online	
ACCESS	III Metrics & More	E Article Recommendations	s Supporting Information

**ABSTRACT:** Biomass burning releases highly reactive methoxyphenols into the atmosphere, which can undergo heterogeneous oxidation and act as precursors for secondary organic aerosol (SOA) formation. Understanding the reactivity of such methoxyphenols at the air—water interface is a matter of major atmospheric interest. Online electrospray ionization mass spectrometry (OESI-MS) is used here to study the oxidation of two methoxyphenols among three phenolic aldehydes, 4-hydroxybenzaldehyde, vanillin, and syringaldehyde, on the surface of water. The OESI-MS results together with cyclic voltammetry measurements at variable pH are integrated into a mechanism describing the heterogeneous oxidative processing of methoxyphenols by gaseous ozone (O<sub>3</sub>) and hydroxyl radicals (HO<sup>•</sup>). For a low



molar ratio of  $O_3 \le 66$  ppbv, the OESI-MS spectra show that the oxidation is dominated by in situ produced HO<sup>•</sup> and results in the production of polyhydroxymethoxyphenols. When the level of  $O_3$  increases (i.e.,  $\ge 78$  times), the ion count of polyhydroxymethoxyphenols increases, while new ring fragmentation products are generated, including conjugated aldehydes and double bonds as well as additional carboxylic acid groups. The interfacial reactivity of methoxyphenols with  $O_3$  and HO<sup>•</sup> is enhanced as the number of methoxy ( $-OCH_3$ ) groups increases (4-hydroxybenzaldehyde < vanillin < syringaldehyde). The experimental observations are summarized in two reaction pathways, leading to the formation of (1) hydroxylated methoxyphenols and (2) multifunctional carboxylic acids from fragmentation of the aromatic ring. The new highly oxygenated products with low volatility are excellent precursors for aqueous SOA formation.

# INTRODUCTION

Biomass burning of lignocellulosic plants and woods from natural wildfires and anthropogenic activities can contribute up to 90% of all primary organic carbon emitted globally from combustion sources.<sup>1</sup> Furthermore, biomass burning contributes 62% of the ca. 8.0 Tg  $y^{-1}$  black carbon and 93% of the ca. 33.9 Tg  $y^{-1}$  organic carbon emitted from combustion globally.<sup>2</sup> The polymeric composition of the cell wall of plants is dominated by lignin, which during biomass burning releases phenols and methoxyphenols (MPs) into the air. For example, the composition of aerosol obtained from a study on burning woods contained 21 and 45% by mass of phenols and MPs, respectively.<sup>3</sup> Thus, MPs constitute the most abundant aromatic species emitted into the atmosphere during biomass burning<sup>3-5</sup> at rates that have been quantified in the gas phase to vary between 1200 and 1500 mg per kg of fuel, depending on the type of wood.<sup>6</sup> MP markers for wood combustion have also been identified in aerosol, fog, and cloud water. $^{7-10}$  In addition, the atmospheric photo-oxidation of MPs can generate light-absorbing molecules in the UV-visible spectrum, which contributes to the brown carbon species found in secondary organic aerosol (SOA).<sup>11,12</sup> The interaction of SOA with light has been proposed as a key player altering the Earth's radiation budget.<sup>11-19</sup>

Recently, the interfacial oxidation of phenols and MPs derived from benzene, toluene, and anisole has been reported as important for the formation of SOA precursors.<sup>20,21</sup> However, the interfacial reactivity of phenolic aldehydes such as 4-hydroxybenzaldehyde (4-HBA), vanillin (VL), and syringaldehyde (SA), three important markers of biomass burning, needs to be elucidated. The large Henry's law constants at 298 K for 4-HBA  $(1.96 \times 10^{6} \text{ M atm}^{-1})$ , VL (4.65  $\times$  10<sup>5</sup> M atm<sup>-1</sup>), and SA (4.39  $\times$  10<sup>5</sup> M atm<sup>-1</sup>) suggest<sup>22</sup> that these surfactant species can accommodate at the air-water interface and even partition into the aqueous phase. Under such atmospheric conditions, these species can be transformed by oxidants such as gaseous ozone (O<sub>3</sub>) and hydroxyl radicals (HO<sup>•</sup>) on the surface of water.<sup>20,21</sup> The homogeneous photooxidative processing of SA dissolved in water has been reported to produce oligomeric and ring-opening products.<sup>23</sup>

Received: June 29, 2020 Revised: August 31, 2020 Published: September 15, 2020





In this work, the oxidative processing of the three phenolic aldehyde markers, 4-HBA, VL, and SA, is explored at the air–water interface to provide new mechanistic insights that explain the composition of aged biomass burning-derived SOA. The use of surface-sensitive online electrospray ionization mass spectrometry (OESI-MS) allows exploring directly the functionalization and fragmentation of the selected phenolic aldehydes by O<sub>3</sub> and HO<sup>•</sup> at the air–water interface.<sup>20,21,24,25</sup> The heterogeneous processes are studied at environmentally relevant low O<sub>3</sub> levels (e.g., molar ratio  $\leq 66$  ppbv) during reactions lasting a microsecond with ion detection within a millisecond. Mass spectrometric data are used to identify reaction products and propose reaction pathways that can form the polyfunctional carboxylic acids prevalent in atmospheric aerosols in dissimilar geographical locations.<sup>26–28</sup>

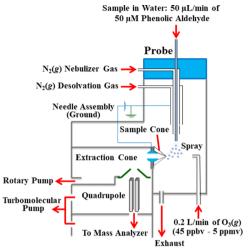
## EXPERIMENTAL SECTION

All experiments were similarly conducted as described in the methods of the previous related works.  $^{20,21,24}$  In brief, fresh 1.0 mM stock solutions of the phenolic aldehydes, VL (Alfa Aesar, 99.9%) and 4-HBA (Sigma-Aldrich, 98.7%), were directly prepared in ultrapure water (18.2 M $\Omega$  cm, ELGA PURELAB flex, Veolia), while fresh 50.0 mM SA (Alfa Aesar, 99.2%) was first prepared in acetonitrile (EMD Millipore, LC/MS grade). All stocks were then diluted in ultrapure water to provide a final phenolic aldehyde concentration of 50  $\mu$ M. These 50  $\mu$ M working concentrations for OESI-MS experiments are comparable to those used in the related photochemical studies in water.<sup>29,30</sup> The molar fraction of acetonitrile after final dilution of the SA stock,  $x_{CH_3CN} = 3.5 \times 10^{-4}$ , is negligible. Thus, the trace of acetonitrile present in the aqueous SA solution does not affect the reactivity and product distribution observed. All experiments were conducted with solutions freshly adjusted by addition of drops of 0.001, 0.010, or 0.100 M NaOH (AMRESCO,  $\geq$ 97%) to pH  $\geq$  6.00 or 0.001 M HCl (EMD, 37.7%) to pH  $\leq$  5.07, as verified with a calibrated pH meter (Thermo Scientific). The pH range studied covers natural tropospheric conditions of different environments.<sup>31-3</sup> The phenolic aldehydes are verified to be stable during the timescale of the experiment under the pH working conditions (Figures S1–S3, Supporting Information).

The concentration of phenolic aldehydes selected is justified by the reported molar ratio of this class of compounds from biomass burning that can range from 2 to 50 ppbv.<sup>34–36</sup> From the predicted partitioning in water by Henry's law, the selected concentrations represent realistic low phenolic aldehyde levels that can be encountered in tropospheric particles and atmospheric water in the vicinity of biomass burning pollution sites.<sup>9</sup> Similarly, tropospheric ozone molar ratios typically vary from 10 to 80 ppbv, depending on the geographical location, its proximity to pollution sources, and the season and time of the day.<sup>18,37–40</sup> Thus, the oxidation of phenolic aldehydes at low ozone levels (e.g.,  $\leq 66$  ppbv) was explored, but higher molar ratios (e.g., ca. 5100 ppbv) were also useful to reveal important oxidative processing steps.

The selected phenolic aldehyde solution at fixed pH was infused at a rate of 50  $\mu$ L min<sup>-1</sup> into a calibrated OESI-MS setup (Scheme 1) that has been customized from a Thermo Scientific MSQ Plus instrument.<sup>20,21,24,25</sup> The operating conditions for OESI-MS measurements were set to a nebulizer voltage of -1.9 kV, a cone voltage of 40 V, a nebulizer pressure of 70 psi N<sub>2</sub>(g), and a drying gas temperature of 250 °C. In

# Scheme 1. Online Electrospray Ionization Mass Spectrometry (OESI-MS) Reactor Diagram



this instrument (Scheme 1), the pneumatically aerosolized solutions forming micrometer-sized droplets at atmospheric pressures were encountered by a 0.2 L min<sup>-1</sup> flow of  $O_3(g)$  reacting with a contact time of a microsecond to generate the oxidized species detected in a microsecond as anions at specific mass to charge ratios (m/z).

Ozone was generated using a spark discharge ozone generator (Ozone Solutions) fed with  $O_2(g)$  (Scott-Gross, UHP) and diluted using  $N_2(g)$  (Scott-Gross, UHP), and the resulting concentration was monitored by a spectrophotometer (Thermo Scientific Evolution Array UV–Visible). The molar ratio of  $O_3$  reported in the figures corresponds to the 61 times final dilution value obtained from diluting with the  $N_2(g)$  nebulizing gas (12.0 L min<sup>-1</sup>).<sup>24</sup> The reported mass spectrometry data were acquired in the negative mode at fixed time intervals (i.e., time  $\geq 1$  min) after background subtraction of the solvent. Following the previous protocols, all identified species are indicated by their m/z values, as shown in the text, figures, and schemes.<sup>20,21,24</sup>

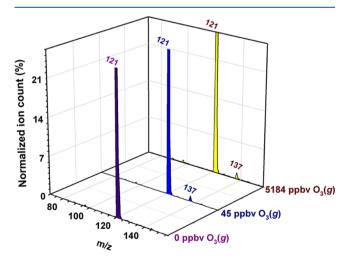
The OESI-MS experiments specifically involve interfacial oxidation reactions as depicted by four steps: (1) the oxidation reaction takes place in the early stage of aerosolization, right upon the uptake of  $O_3(g)$  molecules on the interface of aqueous microdroplets containing the aldehydic phenol. (2) The reacted first-generation droplets undergo evaporation accompanied by fission in smaller droplets that experience charge crowding on the surface, with repulsion overcoming the surface tension. (3) The production of the anions detected takes place from the latest progeny droplets that are the direct precursor to form the ions that reach the detector. Thus, the ions are sputtered off the surface as the charge builds up, making the experiment surface sensitive.<sup>20,21,24,25,41,42</sup> Solvent and gas-phase molecules are pumped away in the intermediate vacuum stage between the OESI probe and analyzer. (4) The ionized species are attracted toward the lower pressure orifice of the entrance cone, from where the ions travel through the entrance capillary at a lower pressure to a quadrupole mass analyzer (accurate to 0.17 amu) for detection at specific  $m/z^{-1}$ ratios. The OESI-MS flow-through reactor for probing fast oxidations at the air-water interface has been comprehensively described.<sup>24,42</sup> In addition, OESI-MS demonstrated that the interfacial  $pK_a$  of acetic and pyruvic acids reduced by 2.8 and 1.8 pH units, respectively, during the partitioning of gas-phase

molecules on the surface of water.<sup>41</sup> This setup has advanced the understanding of how oxidations occur on the surface of water.<sup>20,21,24,25</sup> Moreover, OESI-MS has been validated to be surface-sensitive by detecting the same products during in situ studies under a microsecond contact time and after offline analysis of surface reactions lasting a few hours.<sup>20,43</sup> In conclusion, although oxidations in the liquid bulk of these water-soluble compounds should proceed by similar (or the same) reactions reported below, they are not sampled using this OESI-MS method.

The redox potential of phenolic aldehydes was determined at pH 4.98, 6.01, 7.04, 8.03, 9.04, and 10.04 using cyclic voltammetry with an electrochemical workstation (CHI-660A, CH Instruments, Austin, TX) in a three-electrode configuration cell. The working electrode was a glassy carbon electrode (GCE) with a diameter of 3.0 mm. The GCE was polished with alumina powder (0.05  $\mu$ m) on a microcloth polishing pad, then washed thoroughly with deionized water, and finally sonicated for 10 s to remove any excess alumina powder. The other two electrodes were Ag/AgCl as a reference electrode and platinum as a counter electrode. Analyte solutions of 1.0 mM concentration were prepared in 1.0 M KCl (MP Biomedicals LLC,  $\geq$ 99%) from the stock solutions mentioned above. The solutions were pH-adjusted with 0.010 M HCl or 0.010 M, 0.100 M, and 1.000 M NaOH solutions, measured with the pH meter, and transferred to a 50.0 mL class A volumetric flask for final dilution in the supporting electrolyte solution. The potential was determined from -50to 1200 mV against the reference electrode at a 50 mV s<sup>-1</sup> scan rate. The voltammogram of the supporting electrolyte (e.g., at pH 8.00 as a background solvent) was verified to not overlap with the analytes. Duplicate measurements for each analyte were performed at room temperature to calculate their mean and standard deviation. The detailed processing of cyclic voltammetry data is provided in the Supporting Information.

## RESULTS AND DISCUSSION

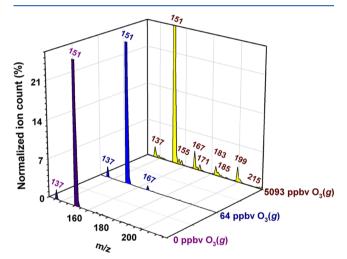
**OESI-MS Analysis.** Figure 1 shows the OESI-MS spectra of the aerosolized aqueous solution of 4-HBA ( $pK_{a,4-HBA} = 7.61$  at



**Figure 1.** OESI-MS spectra of 50  $\mu$ M aqueous solution of 4-HBA at pH 8.0 exposed to a 0.2 L min<sup>-1</sup> flow of gas with or without O<sub>3</sub>(g), as indicated in the labels. Ion count values (*I*) are normalized percentages relative to the most intense peak in each mass spectrum at m/z 121,  $I_{121}$ .

298 K)<sup>44</sup> in the absence and presence of 45 and 5184 ppbv  $O_3(g)$ . When no  $O_3(g)$  is present in the impinging gas encountering the microdroplets  $[0 \text{ ppby } O_3(g)]$ , the only species observed in the mass spectrum at m/z 121 (Figure 1) corresponds to the anion of 4-HBA generated from the loss of a phenolic proton. Therefore, 4-HBA is stable, unless  $O_3(g)$  is added to the reactor. For example, Figure 1 also shows the oxidation of the carbonyl group of 4-HBA on the surface of water at a low  $O_3(g)$  level (45 ppbv), with the peak of a product at m/z 137 assigned to 4-hydroxybenzoic acid observed in the mass spectrum.<sup>45</sup> The normalized ion count of this product,  $I_{137}$ , slightly increases with exposure to 5184 ppbv  $O_3(g)$ , as shown in Figure 1, indicating that the efficiency for its formation is limited by the low reactivity of 4-HBA rather than by the availability of  $O_3(g)$ . Alternatively, an isomer with m/z 137 could have been assigned to the monoanion of 3,4-dihydroxybenzaldehyde (a hydroxylation product).<sup>20,21</sup> However, the absence of heavier products strongly suggests the dominant oxidation to 4-hydroxybenzoic acid, with a stronger deactivating electron-withdrawing group of the aromatic ring than 4-HBA, which inhibits any further reactivity in the microseconds timescale.

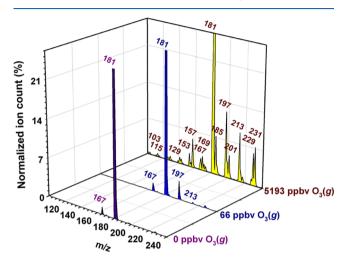
Figure 2 shows the reactivity of VL ( $pK_{a,VL} = 7.38$  at 298 K)<sup>46</sup> on the surface of water when exposed or not to O<sub>3</sub>(g). In



**Figure 2.** OESI-MS spectra of 50  $\mu$ M aqueous solution of VL at pH 8.0 exposed to a 0.2 L min<sup>-1</sup> flow of gas with or without O<sub>3</sub>(g), as indicated in the labels. Ion count values are normalized percentages relative to the most intense peak in each mass spectrum,  $I_{151}$ .

the absence of ozone, the reagent peak of VL is stable, as observed for the abundant anion at m/z 151 with a minor fragment ion at m/z 137 from the loss of 14 amu characteristic of MP breakup by collision-induced dissociation (CID). During exposure to 64 ppbv  $O_3(g)$ , a new product peak is observed at m/z 167, which can be assigned to either isomer 5hydroxyvanillin or vanillic acid. When VL is exposed to 5093 ppbv  $O_3(g)$ , the peak at m/z 167 increases considerably, and several new products can be noticed in the mass spectrum. For example, the peaks at m/z 183 are assigned to 2,5dihydroxyvanillin or 4-formylmuconic acid monomethyl ester. The peak at m/z 199 is due to 2,5,6-trihydroxyvanillin, 4-carboxymuconic acid monomethyl ester, or 4-formyl-5hydroxymuconic acid monomethyl ester, while the peak at m/z 215 is assigned to 2,5,6-trihydroxyvanillic acid or to 2carboxy-5-hydroxymuconic acid monomethyl ester. The CID decarboxylation of the species with m/z 215 yields the peak observed at m/z 171. Furthermore, the small peak at m/z 185 can be explained by the generation of 4-formyl-2-hydroxymuconic acid or an isomer. Decarboxylation of 4-carboxymuconic acid methyl ester with m/z 199 can generate the peak observed at m/z 155 assigned to muconic acid methyl ester.

Figure 3 displays the result of the oxidation of SA  $(pK_{a,SA} = 7.34 \text{ at } 298 \text{ K})^{47}$  on the surface of water exposed to a flow of



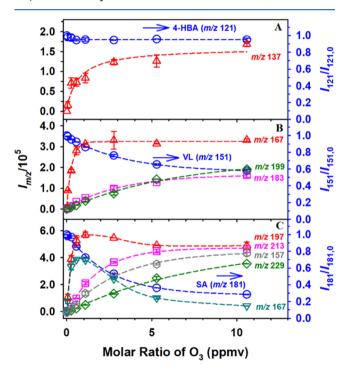
**Figure 3.** OESI-MS spectra of 50  $\mu$ M aqueous solution of syringaldehyde at pH 8.0 exposed to a 0.2 L min<sup>-1</sup> flow of gas with or without O<sub>3</sub>(g), as indicated in the labels. Ion count values are normalized percentages relative to the most intense peak in each mass spectrum,  $I_{181}$ .

0.2 L min<sup>-1</sup> with 0, 66, and 5152 ppbv  $O_3(g)$ . In the absence of  $O_3(g)$ , only the anion of SA at m/z 181 is registered in the mass spectrum together with a small CID fragment loss of 14 amu characteristic of MPs at m/z 167. New products appeared in the mass spectrum upon the addition of 66 and 5152 ppbv  $O_3(g)$ . In more detail, the oxidation product peak with m/z197 can be assigned to 2-hydroxysyringaldehyde or less likely to syringic acid. The peak at m/z 213 could correspond to the formation of 4-formyl-2-methoxymuconic acid methyl ester and 2,6-dihydroxysyringaldehyde or 2-hydroxysyringic acid. The product peak at m/z 229 can be assigned to 2,6dihydroxysyringic acid and 4-formyl-2-methoxy-5-hydroxymuconic acid methyl ester. Alternative products could be generated depending on the position for hydroxyl radical (HO<sup>•</sup>) attack or the specific aromatic bond that is broken during oxidation by O<sub>3</sub>. However, the initial oxidation by HO<sup>•</sup> of the aldehyde side chain is less favorable in the aqueous phase than the direct attack to ring positions activated by electron-donating groups.48

Importantly, the peak at m/z 167 (Figure 3) can also be assigned to 5-hydroxyvanillin in the presence of ozone, when HO<sup>•</sup> radicals substitute  $-OCH_3$  that is eliminated as methanol through oxidative demethoxylation.<sup>48</sup> The normalized ion count of this peak in the absence of ozone is 0.2% and increases to 0.9 and 2.2% for 66 and 5152 ppbv O<sub>3</sub>(g), respectively. In the case of replacing both methoxides, gallaldehyde is formed at m/z 153, which by additional substitution of -H by -OH generates 1,2,3,4-tetrahydroxybenzaldehyde at m/z 169. Alternatively, hydroxylation of gallaldehyde (m/z 153) produces gallic acid (m/z 169), which after ring cleavage by O<sub>3</sub> is converted into 3-carboxy-5hydroxymuconic acid (m/z 201). The last compound is a  $\beta$ dicarboxylic acid that easily decarboxylates<sup>49</sup> into 2-hydroxymuconic acid (m/z 157). Polihydroxylation of the aromatic ring of SA forms pentahydroxybenzoic acid also at m/z 201, a minor contribution to its total ion count. Gallaldehyde also serves as an important precursor to generate 3-formyl-5hydroxymuconic acid (m/z 185), when direct ozonolysis breaks the aromatic ring.

The decarboxylative fragmentation by CID of syringic acid was experimentally confirmed to contribute a product at m/z 153 with an ion count of only 1.6% relative to that of the parent peak. Therefore, it would be expected that the peak at m/z 229 (e.g., 2,6-dihydroxysyringic acid) must not be the source for m/z 185. Other peaks observed in the mass spectrum shown in Figure 3 originate from the ring fragmentation pathway of freshly formed hydroxyl-substituted phenolic aldehydes.<sup>20,21</sup> The list of such products includes 4-formyl-2-,3,5-trihydroxymuconic acid monomethyl ester (m/z 231), maleic acid methyl ester (m/z 103).

For the fixed  $\tau_c = 1 \ \mu s$  in the setup, the experiments below explored a progressive increment of  $[O_3(g)]$  from 45 to 66 ppbv up to 5 ppmv (Figure 4), to provide useful information on the interfacial reactions as if they occur during longer exposure timescales. Figure 4 shows how the ion count for the major oxidation products and the three reactants, as shown in



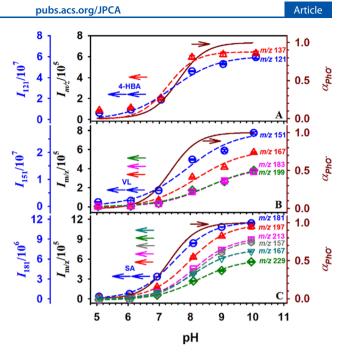
**Figure 4.** Normalized ion count (right axes) at pH 8.0 of (A) 4-HBA  $(m/z \ 121)$ , (B) VL  $(m/z \ 151)$ , and (C) SA  $(m/z \ 181)$  relative to their values before O<sub>3</sub>(g) addition,  $I_{m/z,0}I_{m/z,0r}$  for molar ratios of O<sub>3</sub> from 55.0 ppbv to 10.6 ppmv during experiments at pH 8.0. The respective (left axes) ion count,  $I_{m/z,0}$  of oxidation products is shown for (A) 4-hydroxybenzoic acid  $(m/z \ 137)$ ; (B) 5-hydroxyvanillin  $(m/z \ 167)$ , 2,5-dihydroxyvanillin  $(m/z \ 183)$ , and 2,5,6-trihydroxyvanillin  $(m/z \ 199)$ ; and (C) 2-hydroxysyringaldehyde  $(m/z \ 197)$ , 2,6-dihydroxysyringaldehyde  $(m/z \ 167)$ , and 2-hydroxysyringic acid  $(m/z \ 157)$ .

Figures 1–3, varies with increasing molar ratios of O<sub>3</sub>. The ion counts for the three phenolic aldehyde reactants relative to their value in the absence of O<sub>3</sub>(g),  $I_{m/z'}I_{m/z,0}$ , decrease exponentially for  $I_{181} > I_{151} > I_{121}$ , as shown in Figure 4, indicating the oxidation reactivity trend growths with the number of –OCH<sub>3</sub> substituents in the order 4-HBA < VL < SA. Additional electron-donating –OCH<sub>3</sub> groups decrease the redox potential, making the oxidation of the aromatic molecules thermodynamically more favorable.<sup>50</sup> This predicted reactivity enhancement agrees well with the measured electron transfer potentials for 4-HBA, VL, and SA upon reacting with O<sub>3</sub>(g), which are discussed later.

Complete dissolution of  $O_3(g)$  occurs on the outermost nanometer thin interfacial slab of the microdroplets. Diffusioncontrolled limitations can be neglected in this fast flow-through setup based on the behavior of the parent ions for increasing  $[O_3(g)]$ . The phenolic aldehydes display ion counts  $(I_{121}/I_{121,0},$  $I_{151}/I_{151,0}$ , and  $I_{181}/I_{181,0}$ ) that approach a steady state for increasing  $[O_3(g)]$ , as shown in Figure 4. Therefore, the phenolic aldehydes continuously diffuse from the core of the microdroplets to the nanometer-sized interface in nanoseconds, which is faster than  $\tau_c = 1 \ \mu s$ , and continuously replenish the reagent after it gets oxidized.<sup>20,21,24</sup>

Panel (A) of Figure 4 for 4-HBA (m/z 121) shows that  $I_{137}$ for syringic acid, the only hydroxylated product, increases at higher O<sub>3</sub>(g) levels. Similarly, in panels (B) and (C) of Figure 4,  $I_{167}$  and  $I_{197}$  representing the respective first hydroxylation products of VL  $(m/z \ 151)$  and SA  $(m/z \ 181)$  reach a maximum at ~1 ppmv  $O_3(g)$ . At higher  $O_3(g)$  levels, both  $I_{167}$ and  $I_{197}$  level off or start to decay at ~2.5 ppmv O<sub>3</sub>(g) because of secondary oxidation, eventually reaching a steady-state condition. For example, an increase in second and third oxidation products is registered for VL by  $I_{183}$  and  $I_{199}$  (panel B) and for SA by  $I_{213}$  and  $I_{229}$  (panel C). In addition, the fast development of  $I_{157}$  on higher exposure of SA to  $O_3(g)$  (panel C) indicates that the production of 2-hydroxymuconic acid from the ring fragmentation after oxidative demethoxylation<sup>48</sup> is favored. The comparable values for  $I_{197}$ ,  $I_{213}$ , and  $I_{157}$  from SA oxidation suggest that hydroxylation and ring fragmentation are both competitive oxidation channels. Similarly, the appearance of  $I_{167}$  reaches a maximum at ~1.0 ppmv O<sub>3</sub>(g) (panel C) and then decays as an intermediate in further reactions.

Figure 5 displays the change in  $I_{m/z}$  values for the same species shown in Figure 4 during OESI-MS oxidation experiments of 50.0 µM (A) 4-HBA, (B) VL, or (C) SA solutions for the pH range from 5.0 to 10.0 at constant 5016 ppbv  $O_3(g)$ . The pH range from 5.0 to 10.0 can be used in practical applications to cover the speciation of the three phenolic aldehydes studied, from fully protonated to fully dissociated species. In the context of atmospheric measurements, the experiments at pH 5.0 fall within the upper end of the pH occurrences for fine aerosol  $(0.8 \le pH \le 5.5)$ .<sup>51</sup> The experiments between pH 5.0 and 7.0 also represent well the center of the range occurring in cloud water (2.6  $\leq$  pH  $\leq$ 7.2).<sup>51</sup> Finally, the experiments between pH 7.0 and 10.0 can represent aerosol with a diameter of  $<2.5 \ \mu m \ (PM_{2.5})$  present over agricultural regions,<sup>52,53</sup> where anthropogenic biomass burning emissions often coincide with increased ammonia emissions from fertilizers and livestock.<sup>31,54</sup> Therefore, the experimental pH range outlines a set of true atmospheric conditions.



**Figure 5.** Ion counts,  $I_{m/z}$ , vs pH during experiments with 5016 ppbv O<sub>3</sub>(g) for (A) 4-HBA (m/z 121) and 4-hydroxybenzoic acid (m/z 137); (B) VL (m/z 151), 5-hydroxyvanillin (m/z 167), 2,5-dihydroxyvanillin (m/z 183), and 2,5,6-trihydroxyvanillin (m/z 199); and (C) SA (m/z 181), 2-hydroxysyringaldehdye (m/z 197), 2,6-dihydroxysyringaldehyde (m/z 213), 2,6-dihydroxysyringaldehyde (m/z 167), and 2-hydroxymuconic acid (m/z 157). The dashed lines display sigmoid fittings of the data. The right axes show (brown solid line) the calculated fraction of dissociation for the reactant phenolic aldehydes versus pH.

As expected, the ion count for all species, as shown in Figure 5, increases for higher pH, following a dependence controlled by the acid/base equilibrium of the phenolic group coming from the bulk solvent in this pH range.<sup>41,55,56</sup> Importantly, the data profiles for the three reactants, as shown in Figure 5, are fitted with sigmoid curves that resemble (within experimental error) the speciation of the phenolic aldehyde reactants. Furthermore, the profiles for the products, as shown in Figure 5, have inflexion points that are progressively shifted toward higher pH values (with the exception of  $I_{137}$ ) relative to the reactants. Because (1) the loss of 4-HBA after exposure to  $O_3(g)$ , as shown in panel (A) of Figure 5, is low (typically ~5%), (2) there is no consecutive reactions of the products at m/z 137, and (3) the inflexion point of the sigmoid curve versus pH for the product is shifted to the left of the reactant at m/z 121 (and  $pK_{a'4-HBA}$ ), it is possible to conclude that the more acidic product is 4-hydroxybenzoic acid. Indeed, the progressive shifting of the inflexion points of products toward higher pH values, as shown in panels (B) and (C) of Figure 5, suggests that consecutive hydroxylation of the aromatic ring occurs first for VL and SA before the carbonyl is converted into a carboxylic acid and fragmentation is possible for the products at m/z 157 to be observed.

The strong pH dependence for the oxidations, as shown in Figure 5, is in agreement with the related bulk ozonolysis studies of VL and SA in a stopped-flow reactor, which showed that rates increase exponentially before reaching a plateau in the pH range 7-8.<sup>57</sup> The redox potential of phenolic compounds decreases linearly with increasing pH,<sup>50</sup> favoring electron transfer between phenolic aldehydes and ozone and

pubs.acs.org/JPCA

Article

Table 1. Physical Constants of 4-HDA, VL, and SA	le 1. Physical Constants of 4-	HBA, VL,	and SA
--	--------------------------------	----------	--------

name	MW (amu)	$pK_a^a$	pН	$lpha_{ m PhOH}$	$lpha_{ m PhO}$ -	$E_{\rm red,1} (V)^{b}$	$E_{\rm red,2}~(V)^{c}$	$\Delta E_1 (\mathbf{V})^d$	$\Delta E_2 (V)^e$	$\Delta G_1 \; (\text{kJ mol}^{-1})^f$	$\Delta G_2 \ (\text{kJ mol}^{-1})^f$
4-HBA	122.12	7.61	4.98	0.998	0.002	1.136		-0.126		12.16	
			6.01	0.975	0.025	1.133		-0.123		11.87	
			7.04	0.788	0.212	1.119	0.884	-0.109	0.126	10.52	-12.16
			8.03	0.275	0.725	1.091	0.856	-0.081	0.154	7.82	-14.86
			9.04	0.036	0.964	1.086	0.843	-0.076	0.167	7.33	-16.11
			10.04	0.004	0.996	1.097	0.823	-0.087	0.187	8.39	-18.04
VL	152.15	7.38	4.98	0.996	0.004	0.902		0.108		-10.42	
			6.01	0.959	0.041	0.899		0.111		-10.71	
			7.04	0.686	0.314	0.883	0.669	0.127	0.341	-12.25	-32.90
			8.03	0.183	0.817	0.869	0.684	0.141	0.326	-13.60	-31.45
			9.04	0.021	0.979	0.871	0.672	0.139	0.338	-13.41	-32.61
			10.04	0.002	0.998	0.875	0.652	0.135	0.358	-13.03	-34.54
SA	182.17	7.34	4.98	0.996	0.004	0.848		0.162		-15.63	
			6.01	0.955	0.045	0.848		0.162		-15.63	
			7.04	0.666	0.334	0.847	0.633	0.163	0.377	-15.73	-36.37
			8.03	0.170	0.830	0.872	0.623	0.138	0.387	-13.31	-37.34
			9.04	0.020	0.980	0.886	0.621	0.124	0.389	-11.96	-37.53
			10.04	0.002	0.998	0.89	0.618	0.12	0.392	-11.58	-37.82
		1 C 1 - 1									

<sup>*a*</sup>Experimental values. <sup>44,46,47 b</sup>For PhO<sup>•</sup>/PhO<sup>-</sup> (reaction R1 in Scheme 2). <sup>*c*</sup>PhOH<sup>•+</sup>/PhOH (reaction R2 in Scheme 2) measured with Ag/AgCl at the given pH (Table S1, Supporting Information) and converted to standard hydrogen electrode (SHE) using the equation  $E_{red}$  (SHE) =  $E_{red}$  (Ag/AgCl at. KCl) + 0.197.<sup>64</sup>  $^{d}\Delta E_1 = E_{O_3/O_3^{\bullet^-}} - E_{PhO^{\bullet^+/PhOH}}$ .  $^{c}\Delta E_2 = E_{O_3/O_3^{\bullet^-}} - E_{PhO^{\bullet}/PhO^{-}}$ , with  $E_{O_3/O_3^{\bullet^-}} = 1.01 \text{ V}$ .  $^{65} f \Delta G = -nF\Delta E$  with n = 1 and  $F = 96.485 \text{ kJ mol}^{-1} \text{ V}^{-1}$ .

resulting in an enhanced reactivity at higher pH. In addition, competing photochemical processes should proceed with the participation of the reactive enol tautomer at high pH (when pH > p $K_a$  values, as shown in Table 1).<sup>58</sup>

The results mentioned above for the oxidation experiments on the surface of water indicate that the reactivity of phenolic aldehydes increases in the order 4-HBA < VL < SA. The comparative reactivity trend recorded by OESI-MS can be explained by the electron density donation effect added to 4-HBA by one or two extra methoxy ( $-OCH_3$ ) substituents. The increase in electron density of the aromatic ring by each additional  $-OCH_3$  group makes the reactant more susceptible toward electrophilic attack by ozone.<sup>59–61</sup> The two reactivity channels observed correspond to oxidation pathways from in situ generated HO<sup>•</sup> and direct ring fragmentation by O<sub>3</sub>.<sup>20,21</sup> These pathways are discussed in detail below.

**Oxidative Functionalization of the Aromatic Rings by HO**<sup>•</sup>. Scheme 2 presents an integrated reaction mechanism for the oxidation of the three phenolic aldehydes studied by in situ generated HO<sup>•</sup> radicals.<sup>20</sup> All reactions mentioned in the reaction schemes below are numbered consecutively. A key step for in situ production of HO<sup>•</sup> is the electron transfer between phenolic aldehydes and dissolved O<sub>3</sub> (reactions R1 and R2 in Scheme 2). An ozonide radical anion  $(O_3^{\bullet-})$  is formed in reaction R1 together with the phenoxyl radical, which can also be formed from the phenol radical cation undergoing ultrafast deprotonation.<sup>20,21,62,63</sup>

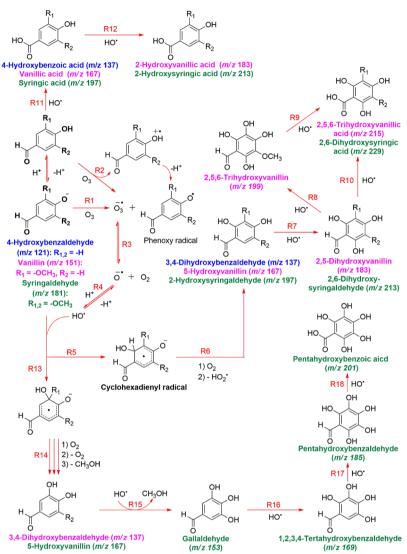
As a working example, the determined first and second redox potentials of SA at pH 8.03 (Supporting Information),  $E_{\rm red,1} =$ 0.872 V and  $E_{\rm red,2} = 0.623$  V (Table 1), respectively, and that of ozone,  $E_{\rm O_3} = 1.01$  V (at pH 7.0), indicate thermodynamically favorable  $\Delta E_2 = +0.385$  V and  $\Delta E_1 = +0.138$  V for electron transfer reactions R1 and R2, respectively. For the same example at pH 8.03, Table 1 also shows the calculated free energy change for reactions R1 and R2 of SA with ozone,  $\Delta G_2 = -13.4$  kJ mol<sup>-1</sup> and  $\Delta G_1 = -37.4$  kJ mol<sup>-1</sup>, respectively. Overall, Table 1 lists the experimental  $\Delta E$  and  $\Delta G$  values for the pH range from 5 to 10 of 4-HBA, VL, and SA, together with the physical constants and dissociation fractions of the three compounds at a given pH. A comparison of  $\Delta G_1$  and  $\Delta G_2$  values for each of the three compounds shows a larger tendency to undergo electron transfer with ozone for the phenoxide anion (reaction R1) rather than for the cation radical (reaction R2) at pH  $\geq$  7.0. Instead, at pH  $\leq$  6.0, as the fraction of phenol  $\alpha_{\text{PhOH}} \rightarrow 1$  for VL and SA, the only possibility left for electron transfer of these compounds implies the participation of their cation radicals (reaction R2 in Scheme 2), which is thermodynamically prevented for 4-HBA.

Equilibrium R3 with  $K_{eq} = 5.5 \times 10^{-7}$  M at 20 °C indicates that  $O_3^{\bullet-}$  coexists with the strong base  $O^{\bullet-}$  (p $K_a = 11.8$ ) which, under relevant environmental (and the experimental conditions here), is converted immediately by reaction R4 into its conjugated acid HO<sup>•.61,66</sup> This in situ generated HO<sup>•</sup> can play a crucial role in the degradation of organic compounds on atmospheric interfaces and provide useful information about the reactivity of the oxidizer, independent of its source and mechanism of formation.  $^{20,21,63,67,68}$  Once generated, HO $^{\bullet}$ attacks (i.e., in reaction R5) the activated positions (rich in electron density from the donating substituents -OH and/or  $-OCH_3$ ), producing a cyclohexadienyl radical that adds up molecular oxygen  $(O_2)$  through reaction R6.<sup>60,61</sup> The cyclohexadienyl peroxy radical generated eliminates hydroperoxyl radicals  $(HO_2^{\bullet})$  as part of reaction R6, producing a firstgeneration hydroxylated product after rearrangement. Although less likely in our system due to the low probability of detecting the encounter among two radicals, the phenoxyl radical has been proposed to also react directly with HO<sup>•</sup> to produce the corresponding hydroxylated product.<sup>69</sup>

The ortho HO<sup>•</sup> attack to the three phenolic aldehyde molecular probes creates cyclohexadienyl radicals in reaction R5 (Scheme 2). The cyclohexadienyl radicals react with  $O_2$  at a rate controlled by diffusion (e.g., for MPs  $2 \times 10^9 \text{ M}^{-1} \text{ s}^{-1})^{68}$  and eliminate a hydroperoxyl radical in reaction R6 to form hydroxylated products. This kind of functionalization reaction

Article

Scheme 2. Proposed Oxidation Mechanism of (Blue Font) 4-HBA, (Pink Font) VL, and (Green Font) SA by Hydroxyl Radicals on the Surface of Water<sup>*a,b*</sup>



<sup>a</sup>Reaction numbers are given in red font. <sup>b</sup>Observed products and m/z values are given in the same color of the precursor.

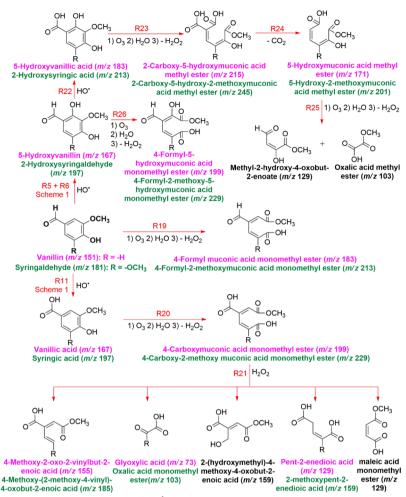
was observed on the surface of water for a series of substituted catechols.<sup>20,21</sup> For VL (precursor and products in purple font in Scheme 2), the consecutive hydroxylation by reactions R5 to R10 produces 5-hydroxyvanillin  $(m/z \ 167)$ , 2,5-dihydroxyvanillin  $(m/z \ 183)$ , 2,5,6-trihydroxyvanillin  $(m/z \ 199)$ , and finally 2,5,6-trihydroxyvaleric acid (m/z 215). Similarly, for SA (precursor and products in green font in Scheme 2), hydroxylation reactions produce 2-hydroxysyringaldehyde (m/z 197), 2,6-dihydroxysyringaldehyde (m/z 213), and 2,6dihydroxysyringic acid (m/z 229). The related ring hydroxylation product of 4-HBA (precursor and product in blue font in Scheme 2) would be 3,4-HBA (m/z 137), which is depicted in Scheme 2, but not believed to be efficiently formed in the microsecond contact time. Instead, >C=O side chain oxidation into -COOH by reaction R11 converts 4-HBA, VL, and SA into 4-hydroxybenzoic acid  $(m/z \ 137)$ , vanillic acid  $(m/z \ 167)$ , and syringic acid  $(m/z \ 197)$ , respectively.<sup>45</sup> In addition, 5-hydroxyvanillin and 2-hydroxysyringaldehyde can be converted (reaction R12) into 2-hydroxyvanillic acid (m/z)183) and 2-hydroxysyringic acid  $(m/z \ 213)$ , respectively.

MPs can also undergo oxidative demethoxylation reaction R13 by HO<sup>•</sup> attack to form *ipso* adducts. There is a low probability in this study for the ipso adducts to eliminate CH<sub>3</sub>OH directly to produce semiquinone radicals, a reaction that would require anoxic conditions.<sup>48,68</sup> Instead, the ipso adduct radicals undergo a complex multistep process (not shown in Scheme 2), symbolized by reaction R14, to incorporate O2 forming peroxyl radicals, lose O2 to form cyclohexadienyl products, and eliminate CH<sub>3</sub>OH to produce 3,4-dihydroxybenzaldehyde (m/z 137), 5-hydroxyvanillin (m/z167), and consecutively, by an analogous reaction R15, gallaldehyde (m/z 153). Once gallaldehyde is produced, its further hydroxylation generates 1,2,3,4-tetrahydroxybenzladehyde  $(m/z \ 169)$ , pentahydroxybenzaldehyde  $(m/z \ 185)$ , and pentahydroxybenzoic acid  $(m/z \ 201)$  by reactions R16, R17, and R18, respectively.<sup>20,21</sup>

**Fragmentation of the Aromatic Rings by O<sub>3</sub>.** Scheme 3 shows the proposed ring-opening fragmentation reactions for VL and SA. VL, SA, and their hydroxylation products react with  $O_3(g)$  forming primary ozonides, which then rearrange as Criegee intermediates<sup>70,71</sup> and react with water to form

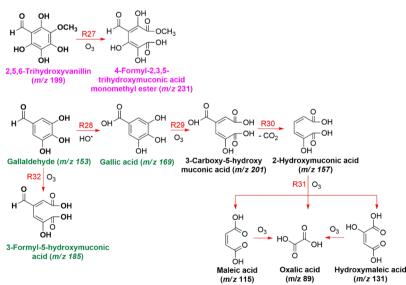
Article

Scheme 3. Proposed Ring-Opening Oxidation Reactions of VL, SA, and Some of their Hydroxylation Products by Ozone on the Surface of Water<sup>a,b</sup>



<sup>a</sup>Reaction numbers are given in red font. <sup>b</sup>Observed products and m/z values are given in the same color of the precursor.

Scheme 4. Proposed Ring-Opening Oxidation Reactions of the Selected Hydroxylation Products by Ozone on the Surface of  $Water^{a,b}$ 



<sup>*a*</sup>Reaction numbers are given in red font. <sup>*b*</sup>Observed products are indicated by their m/z values.

#### The Journal of Physical Chemistry A

hydroperoxide intermediates that decompose, eliminating  $H_2O_2^{72}$  into dicarboxylic acid monomethyl esters, as displayed in Scheme 3. For example, reaction R19 (Scheme 3) shows that VL and SA generate 4-formyl muconic acid monomethyl ester (m/z 183) and 4-formyl-2-methoxymuconic acid monomethyl ester (m/z 213), respectively. The previous process can occur also for vanillic and syringic acids (products of reaction R11 in Scheme 2) to form 4-carboxymuconic acid monomethyl ester (m/z 199) and 4-carboxy-2-methoxymuconic acid monomethyl ester (m/z 229) in reaction R20 (Scheme 3), respectively.

The previous products derived from VL and SA can undergo Baeyer–Villiger oxidation<sup>73</sup> with in situ formed  $H_2O_2$  in reaction R21 (Scheme 3) to produce 4-methoxy-2-oxo-2vinylbut-2-enoic acid (m/z 155) and 4-methoxy-(2-methoxy-4vinyl)-4-oxobut-2-enoic acid (m/z 185), pent-2-enedioic acid (m/z 129) and 2-methoxypent-2-enedioic acid (m/z 159), glyoxylic acid (m/z 73) and oxalic acid monomethyl ester (m/z103), respectively, among other common products such as 2-(hydroxymethyl)-4-methoxy-4-oxobut-2-enoic acid (m/z 159), maleic acid monomethyl ester (m/z 129), and formic acid (m/z 45).

Scheme 3 also includes examples of additional reactions for the products 5-hydroxyvanillic acid  $(m/z \ 183)$  and 2hydroxysyringic acid  $(m/z \ 213)$  from the hydroxylation sequence R5 + R6 + R22, respectively forming 2-carboxy-5hydroxymuconic acid methyl ester (m/z 215) and 2-carboxy-5hydroxy-2-methoxymuconic acid methyl ester  $(m/z \ 245)$  after aromatic ring cleavage. The previous  $\alpha_{,\beta}$ -unsaturated carboxylic acid monomethyl esters could undergo decarboxylation reaction R24, producing 5-hydroxymuconic acid methyl ester  $(m/z \ 171)$  and 5-hydroxy-2-methoxymuconic acid methyl ester  $(m/z \ 201)$ . Direct ozonolysis in reaction R25 (Scheme 3) could also explain the production of methyl-2-hydroxy-4oxobut-2-enoate  $(m/z \ 129)$  and oxalic acid methyl ester  $(m/z \ 129)$ 103). For instance, direct ozonolysis of 5-hydroxyvanillin (m/z)167) and 2-hydroxysyringaldehyde  $(m/z \ 197)$  in reaction R26 (Scheme 3) results in the production of 4-formyl-5hydroxymuconic acid monomethyl ester  $(m/z \ 199)$  and 4formyl-2-methoxy-5-hydroxymuconic acid monomethyl ester (m/z 229), respectively. Because no MS peaks other than m/z137 are observed for the oxidation of 4-HBA (m/z 121), this molecule did not undergo ring-opening ozonolysis at the microsecond contact time of the experiments.

Scheme 4 displays some final examples of ring cleavage products<sup>74</sup> from 2,5,6-trihydroxyvanillin (m/z 199) and gallaldehyde (m/z 153) generated in reactions R8 and R15 of Scheme 2, respectively. The former generates 4-formyl-2,3,5-trihydroxymuconic acid monomethyl ester (m/z 231) in reaction R27 (Scheme 4), and the latter after hydroxylation into gallic acid (reaction R28) produces in reaction R29 3-carboxy-5-hydroxymuconic acid (m/z 201). Decarboxylation in reaction R30 results in 2-hydroxymuconic acid (m/z 157), which can be further oxidized by reaction R31 into smaller species such as oxalic acid (m/z 89), maleic acid (m/z 115), and hydroxymaleic acid (m/z 131). The ring cleavage of gallaldehyde in reaction R32 explains the formation of 3-formyl-5-hydroxymuconic acid (m/z 185).

#### CONCLUSIONS AND ATMOSPHERIC IMPLICATIONS

This study reveals how interfacial reactions of two MPs among three phenolic aldehydes present in biomass burning emissions pubs.acs.org/JPCA

Article

contribute to increase the oxygenated fraction of organic compounds in brown carbon species found in SOA. The observed oxidation of phenolic aldehydes on the surface of water proceeds mainly through hydroxylation (Scheme 2) as well as by aromatic ring cleavage (Schemes 3 and 4) in the pH range investigated.<sup>20,21</sup> The key step to initiate hydroxylation reactions on the surface of water involves electron transfer reactions R1 and R2. These results demonstrate that  $O_3(g)$ levels as low as 45-66 ppbv can break the aromatic ring and also induce the formation of HO<sup>•</sup> radicals, as determined by the products detected in our atmospheric pressure ionization technique. Cyclic voltammetry measurements support the interpretation that at pH  $\leq$  10.8, the formed conjugate base  $O^{\bullet-}$  (pK<sub>a</sub> = 11.8),<sup>66</sup> from the favorable electron transfers from the acid/base pair, determined in Table 1 by  $\Delta G_1$  and  $\Delta G_2$ , becomes fully protonated. Furthermore, the results at pH 5 for the fully undissociated species should be valid down to pH 1, as for both cases, the undissociated fraction  $\alpha_{PhOH} \rightarrow 1$ .

OESI-MS experiments revealed the oxidative pathways for three phenolic aldehydes, and the careful interpretation of the data sets enabled inferring the reactive centers for proposing consecutive HO<sup>•</sup>- and O<sub>3</sub>-driven reactions. The generation of hydroxyl-substituted aromatics generally preceded the formation of aromatic and aliphatic carboxylic acids. Future works should compare the reaction products (1) between bulk and surface studies and (2) at longer exposures with those from direct photochemistry that generate intermediates with humiclike fluorescence properties that can be structurally related to the formation of dimer and trimer species by self-reaction of phenoxyl radicals or cross-reaction of phenoxyl and ketyl radicals.<sup>30</sup>

Given the availability of information for VL, this molecule can be used to compare the competitive loss pathways of phenolic carbonyls among phases and reaction pathways. The aqueous photodegradation of VL under simulated sunlight irradiation proceeds with a lifetime of  $\tau_{VL+h\nu(aq)} = 1.1-2.2$ h.<sup>29,30</sup> The rate constants of VL reacting with HO<sup>•</sup> in water at pH 5 and 9 are  $k_{\rm VL+HO} \cdot (_{aq})$ ,  $s = 3.3 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$  and  $k_{\rm VL+HO} \cdot (_{aq})$ ,  $g = 6.6 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$ , respectively.<sup>75</sup> To estimate the lifetime of VL against loss by HO<sup>•</sup> on the surface of water and in aqueous particles, the concentration of this radical is constrained by a low limit value  $[HO^{\bullet}(aq)] = 7.5 \times 10^{-15}$ radicals  $L^{-1}$ .<sup>76</sup> Therefore, the lifetimes of VL reacting with HO<sup>•</sup> in water at pH 5 and 9 are  $\tau_{VL+HO}$  (aq),5 = 7.8 h and  $\tau_{\rm VL+HO} \cdot_{(aq),9}$  = 3.9 h, respectively. Instead, the corresponding lifetime of VL on the surface of water is affected by the larger in situ produced HO<sup>•</sup> on the surface of water. From a previous air-water interface study with substituted catechols, and after scaling down the oxidizer level to 50 ppbv, an upper limit for the in situ generated  $[HO^{\bullet}]_{interface} = 2.0 \ (\pm 0.5) \times 10^{-12}$  radicals  $L^{-1}$  is obtained. In consequence, the interfacial oxidation of VL should proceed with much shorter lifetimes  $\tau_{\rm VL+HO}$  (interface).5 = 105 s and  $\tau_{\rm VL+HO}$  (interface).9 = 53 s, resulting in the major loss channel. Considering the average rate constant for the reaction of MPs with HO<sup>•</sup> in the gas phase  $(k_{\text{MPs+HO}}) = 6 \times 10^{-11} \text{ cm}^3 \text{ molecules}^{-1} \text{ s}^{-1}, 5^9 \text{ for an average}$ tropospheric  $[HO^{\bullet}] = 1.6 \times 10^6$  radicals cm<sup>-3</sup>,<sup>77</sup> the residence time in the gas phase for MPs is  $\tau_{\text{MPs+HO}} \cdot_{(g)} = 2.9$  h, which is much larger than at the interface but comparable to that in water. In contrast, the atmospheric lifetime of the related MP 4-propylguaiacol reacting with 40 ppbv  $O_3(g)$  on the surface of dry NaCl is  $\tau_{MPs+O_3(ads.)} = 28 h^{74}$  which is even ~10 times

longer than  $\tau_{MPs+HO^{\bullet}(g)}$ . Thus, the HO<sup>•</sup>-driven reactivity observed here at the air-water interface, despite the oxidizer origin, is expected to contribute significantly to the daytime loss of MPs on the surface of atmospheric aerosols and waters.

New heterogeneous processes, which have not been considered in atmospheric models before, show that ubiquitous MPs can generate precursors for SOA formation. The quickly produced water-soluble organic compounds by hydroxylation reactions are more likely to partition into the particle phase, where they can be further processed into aged species with different optical properties.<sup>20,21,43,78</sup> In addition, the large O/C ratio of the unsaturated dicarboxylic acids and related compounds produced from ozonolysis reactions suggests that the aged material is less volatile and more water soluble than its corresponding MP precursors.<sup>13,43,79</sup> This combined effect may significantly alter the light scattering and absorption properties of oxidized SOA.<sup>12,19,25</sup> Because the residence time of typical biomass burning aerosols against deposition is 1 week, future surface oxidation studies during longer timescales and variable environmental conditions should be useful to reveal the magnitude of absorption property changes.<sup>12,43</sup>

#### ASSOCIATED CONTENT

#### **3** Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.jpca.0c05944.

Control experiments, cyclic voltammograms at variable pH, cyclic voltammetry processing to obtain their second derivative, methodology for inflection potential determination, and experimental redox potentials versus Ag/AgCl reference electrode (PDF)

#### AUTHOR INFORMATION

#### **Corresponding Author**

Marcelo I. Guzman – Department of Chemistry, University of Kentucky, Lexington, Kentucky 40506, United States;
orcid.org/0000-0002-6730-7766; Phone: (859)323-2892; Email: marcelo.guzman@uky.edu

#### Author

Md. Sohel Rana – Department of Chemistry, University of Kentucky, Lexington, Kentucky 40506, United States

Complete contact information is available at: https://pubs.acs.org/10.1021/acs.jpca.0c05944

#### Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENTS

M.I.G. acknowledges funding from the U.S.A. National Science Foundation under award 1903744.

# REFERENCES

(1) Ito, A.; Penner, J. E. Historical emissions of carbonaceous aerosols from biomass and fossil fuel burning for the period 1870–2000. *Global Biogeochem. Cycles* **2005**, *19*, GB2028.

(2) Bond, T. C.; Streets, D. G.; Yarber, K. F.; Nelson, S. M.; Woo, J.-H.; Klimont, Z. A technology-based global inventory of black and organic carbon emissions from combustion. *J. Geophys. Res.: Atmos.* **2004**, *109*, D14203.

(3) Hawthorne, S. B.; Krieger, M. S.; Miller, D. J.; Mathiason, M. B. Collection and quantitation of methoxylated phenol tracers for

atmospheric pollution from residential wood stoves. *Environ. Sci. Technol.* **1989**, *23*, 470–475.

(4) Hawthorne, S. B.; Miller, D. J.; Langenfeld, J. J.; Krieger, M. S. PM-10 high-volume collection and quantitation of semi- and nonvolatile phenols, methoxylated phenols, alkanes, and polycyclic aromatic hydrocarbons from winter urban air and their relationship to wood smoke emissions. *Environ. Sci. Technol.* **1992**, *26*, 2251–2262.

(5) Myers-Pigg, A. N.; Griffin, R. J.; Louchouarn, P.; Norwood, M. J.; Sterne, A.; Cevik, B. K. Signatures of biomass burning aerosols in the plume of a saltmarsh wildfire in South Texas. *Environ. Sci. Technol.* **2016**, *50*, 9308–9314.

(6) Schauer, J. J.; Kleeman, M. J.; Cass, G. R.; Simoneit, B. R. T. Measurement of emissions from air pollution sources. 3.  $C_1-C_{29}$  organic compounds from fireplace combustion of wood. *Environ. Sci. Technol.* **2001**, *35*, 1716–1728.

(7) Bruns, E. A.; El Haddad, I.; Slowik, J. G.; Kilic, D.; Klein, F.; Baltensperger, U.; Prévôt, A. S. H. Identification of significant precursor gases of secondary organic aerosols from residential wood combustion. *Sci. Rep.* **2016**, *6*, 27881.

(8) Lauraguais, A.; Coeur-Tourneur, C.; Cassez, A.; Deboudt, K.; Fourmentin, M.; Choël, M. Atmospheric reactivity of hydroxyl radicals with guaiacol (2-methoxyphenol), a biomass burning emitted compound: Secondary organic aerosol formation and gas-phase oxidation products. *Atmos. Environ.* **2014**, *86*, 155–163.

(9) Sagebiel, J. C.; Seiber, J. N. Studies on the occurrence and distribution of wood smoke marker compounds in foggy atmospheres. *Environ. Toxicol. Chem.* **1993**, *12*, 813–822.

(10) Yu, L.; Smith, J.; Laskin, A.; Anastasio, C.; Laskin, J.; Zhang, Q. Chemical characterization of SOA formed from aqueous-phase reactions of phenols with the triplet excited state of carbonyl and hydroxyl radical. *Atmos. Chem. Phys.* **2014**, *14*, 13801–13816.

(11) Feng, Y.; Ramanathan, V.; Kotamarthi, V. R. Brown carbon: a significant atmospheric absorber of solar radiation? *Atmos. Chem. Phys.* 2013, 13, 8607–8621.

(12) Laskin, A.; Laskin, J.; Nizkorodov, S. A. Chemistry of atmospheric brown carbon. *Chem. Rev.* **2015**, *115*, 4335–4382.

(13) Eugene, A. J.; Xia, S.-S.; Guzman, M. I. Aqueous photochemistry of glyoxylic acid. J. Phys. Chem. A 2016, 120, 3817–3826.

(14) Guzmán, M. I.; Colussi, A. J.; Hoffmann, M. R. Photoinduced oligomerization of aqueous pyruvic acid. J. Phys. Chem. A 2006, 110, 3619–3626.

(15) Xia, S.-S.; Eugene, A. J.; Guzman, M. I. Cross photoreaction of glyoxylic and pyruvic acids in model aqueous aerosol. *J. Phys. Chem. A* **2018**, *122*, 6457–6466.

(16) Hallquist, M.; Wenger, J. C.; Baltensperger, U.; Rudich, Y.; Simpson, D.; Claeys, M.; Dommen, J.; Donahue, N. M.; George, C.; Goldstein, A. H.; et al. The formation, properties and impact of secondary organic aerosol: current and emerging issues. *Atmos. Chem. Phys.* **2009**, *9*, 5155–5236.

(17) Shrivastava, M.; Cappa, C. D.; Fan, J.; Goldstein, A. H.; Guenther, A. B.; Jimenez, J. L.; Kuang, C.; Laskin, A.; Martin, S. T.; Ng, N. L.; et al. Recent advances in understanding secondary organic aerosol: Implications for global climate forcing. *Rev. Geophys.* 2017, *55*, 509–559.

(18) IPCC, Climate change 2013: The physical science basis. Contribution of working group I to the fifth assessment report of the intergovernmental panel on climate change; Cambridge University Press: Cambridge, United Kingdom and New York, NY, USA, 2013; p 1535.

(19) von Schneidemesser, E.; Monks, P. S.; Allan, J. D.; Bruhwiler, L.; Forster, P.; Fowler, D.; Lauer, A.; Morgan, W. T.; Paasonen, P.; Righi, M.; et al. Chemistry and the linkages between air quality and climate change. *Chem. Rev.* **2015**, *115*, 3856–3897.

(20) Pillar, E. A.; Camm, R. C.; Guzman, M. I. Catechol oxidation by ozone and hydroxyl radicals at the air-water interface. *Environ. Sci. Technol.* **2014**, *48*, 14352–14360.

(21) Pillar, E. A.; Guzman, M. I. Oxidation of substituted catechols at the air-water interface: Production of carboxylic acids, quinones, and polyphenols. *Environ. Sci. Technol.* **2017**, *51*, 4951–4959.

#### The Journal of Physical Chemistry A

(22) US-EPA, Estimation Programs Interface Suite<sup>TM</sup> for Microsoft® Windows, v 4.11; United States Environmental Protection Agency: Washington, DC, USA, 2012.

(23) Huang, D. D.; Zhang, Q.; Cheung, H. H. Y.; Yu, L.; Zhou, S.; Anastasio, C.; Smith, J. D.; Chan, C. K. Formation and evolution of aqSOA from aqueous-phase reactions of phenolic carbonyls: Comparison between ammonium sulfate and ammonium nitrate solutions. *Environ. Sci. Technol.* **2018**, *52*, 9215–9224.

(24) Pillar, E. A.; Guzman, M. I.; Rodriguez, J. M. Conversion of iodide to hypoiodous acid and iodine in aqueous microdroplets exposed to ozone. *Environ. Sci. Technol.* **2013**, *47*, 10971–10979.

(25) Pillar-Little, E.; Guzman, M. An overview of dynamic heterogeneous oxidations in the troposphere. *Environments* **2018**, *5*, 104.

(26) Andreae, M. O.; Talbot, R. W.; Andreae, T. W.; Harriss, R. C. Formic and acetic acid over the central Amazon region, Brazil: 1. Dry season. J. Geophys. Res.: Atmos. 1988, 93, 1616–1624.

(27) Hu, Q.; Xie, Z.; Wang, X.; Kang, H.; Zhang, Y.; Ding, X.; Zhang, P. Monocarboxylic and dicarboxylic acids over oceans from the East China Sea to the Arctic Ocean: Roles of ocean emissions, continental input and secondary formation. *Sci. Total Environ.* **2018**, 640–641, 284–292.

(28) Kawamura, K.; Tachibana, E.; Okuzawa, K.; Aggarwal, S. G.; Kanaya, Y.; Wang, Z. F. High abundances of water-soluble dicarboxylic acids, ketocarboxylic acids and  $\alpha$ -dicarbonyls in the mountaintop aerosols over the North China Plain during wheat burning season. *Atmos. Chem. Phys.* **2013**, *13*, 8285–8302.

(29) Smith, J. D.; Kinney, H.; Anastasio, C. Phenolic carbonyls undergo rapid aqueous photodegradation to form low-volatility, light-absorbing products. *Atmos. Environ.* **2016**, *126*, 36–44.

(30) Vione, D.; Albinet, A.; Barsotti, F.; Mekic, M.; Jiang, B.; Minero, C.; Brigante, M.; Gligorovski, S. Formation of substances with humic-like fluorescence properties, upon photoinduced oligomerization of typical phenolic compounds emitted by biomass burning. *Atmos. Environ.* **2019**, *206*, 197–207.

(31) Andreae, M. O.; Merlet, P. Emission of trace gases and aerosols from biomass burning. *Global Biogeochem. Cycles* **2001**, *15*, 955–966.

(32) Alves, C. A.; Gonçalves, C.; Pio, C. A.; Mirante, F.; Caseiro, A.; Tarelho, L.; Freitas, M. C.; Viegas, D. X. Smoke emissions from biomass burning in a Mediterranean shrubland. *Atmos. Environ.* **2010**, *44*, 3024–3033.

(33) Keene, W. C.; Sander, R.; Pszenny, A. A. P.; Vogt, R.; Crutzen, P. J.; Galloway, J. N. Aerosol pH in the marine boundary layer. *J. Aerosol Sci.* **1998**, *29*, 339–356.

(34) Veres, P.; Roberts, J. M.; Burling, I. R.; Warneke, C.; de Gouw, J.; Yokelson, R. J. Measurements of gas-phase inorganic and organic acids from biomass fires by negative-ion proton-transfer chemical-ionization mass spectrometry. *J. Geophys. Res.* **2010**, *115*, D23302.

(35) Stockwell, C. E.; Veres, P. R.; Williams, J.; Yokelson, R. J. Characterization of biomass burning emissions from cooking fires, peat, crop residue, and other fuels with high-resolution proton-transfer-reaction time-of-flight mass spectrometry. *Atmos. Chem. Phys.* **2015**, *15*, 845–865.

(36) Gilman, J. B.; Lerner, B. M.; Kuster, W. C.; Goldan, P. D.; Warneke, C.; Veres, P. R.; Roberts, J. M.; de Gouw, J. A.; Burling, I. R.; Yokelson, R. J. Biomass burning emissions and potential air quality impacts of volatile organic compounds and other trace gases from fuels common in the US. *Atmos. Chem. Phys.* **2015**, *15*, 13915–13938.

(37) Lefohn, A. S.; Shadwick, D.; Oltmans, S. J. Characterizing changes in surface ozone levels in metropolitan and rural areas in the United States for 1980–2008 and 1994–2008. *Atmos. Environ.* **2010**, *44*, 5199–5210.

(38) Parrish, D. D.; Law, K. S.; Staehelin, J.; Derwent, R.; Cooper, O. R.; Tanimoto, H.; Volz-Thomas, A.; Gilge, S.; Scheel, H.-E.; Steinbacher, M.; et al. Long-term changes in lower tropospheric baseline ozone concentrations at northern mid-latitudes. *Atmos. Chem. Phys.* **2012**, *12*, 11485–11504.

(39) Cooper, O. R.; Gao, R.-S.; Tarasick, D.; Leblanc, T.; Sweeney, C. Long-term ozone trends at rural ozone monitoring sites across the United States, 1990–2010. *J. Geophys. Res.* **2012**, *117*, D22307.

(40) Oltmans, S. J.; Lefohn, A. S.; Shadwick, D.; Harris, J. M.; Scheel, H. E.; Galbally, I.; Tarasick, D. W.; Johnson, B. J.; Brunke, E.-G.; Claude, H.; et al. Recent tropospheric ozone changes – A pattern dominated by slow or no growth. *Atmos. Environ.* **2013**, *67*, 331–351.

(41) Eugene, A. J.; Pillar, E. A.; Colussi, A. J.; Guzman, M. I. Enhanced acidity of acetic and pyruvic acids on the surface of water. *Langmuir* **2018**, *34*, 9307–9313.

(42) Guzman, M. I.; Athalye, R. R.; Rodriguez, J. M. Concentration effects and ion properties controlling the fractionation of halides during aerosol formation. *J. Phys. Chem. A* **2012**, *116*, 5428–5435.

(43) Pillar, E. A.; Zhou, R.; Guzman, M. I. Heterogeneous oxidation of catechol. J. Phys. Chem. A 2015, 119, 10349–10359.

(44) Serjeant, E. P.; Dempsey, B. Ionisation Constants of Organic Ccids in Aqueous Solution; Pergamon Press, 1979; p 988.

(45) Net, S.; Alvarez, E. G.; Gligorovski, S.; Wortham, H. Heterogeneous reactions of ozone with methoxyphenols, in presence and absence of light. *Atmos. Environ.* **2011**, *45*, 3007–3014.

(46) Robinson, R. A.; Kiang, A. K. The ionization constants of vanillin and two of its isomers. *Trans. Faraday Soc.* **1955**, *51*, 1398–1402.

(47) Ragnar, M.; Lindgren, C. T.; Nilvebrant, N.-O. pK<sub>a</sub>-values of guaiacyl and syringyl phenols related to lignin. *J. Wood Chem. Technol.* **2000**, *20*, 277–305.

(48) Steenken, S.; O'Neill, P. Oxidative demethoxylation of methoxylated phenols and hydroxybenzoic acids by the hydroxyl radical. An in situ electron spin resonance, conductometric pulse radiolysis and product analysis study. *J. Phys. Chem.* **1977**, *81*, 505–508.

(49) Smith, J. Organic Chemistry; McGraw-Hill Education: New York, 2010.

(50) Aracri, E.; Tzanov, T.; Vidal, T. Use of cyclic voltammetry as an effective tool for selecting efficient enhancers for oxidative bioprocesses: Importance of pH. *Ind. Eng. Chem. Res.* **2013**, *52*, 1455–1463.

(51) Pye, H. O. T.; Nenes, A.; Alexander, B.; Ault, A. P.; Barth, M. C.; Clegg, S. L.; Collett, J. L., Jr.; Fahey, K. M.; Hennigan, C. J.; Herrmann, H.; et al. The acidity of atmospheric particles and clouds. *Atmos. Chem. Phys.* **2020**, *20*, 4809–4888.

(52) Parmar, R. S.; Satsangi, G. S.; Kumari, M.; Lakhani, A.; Srivastava, S. S.; Prakash, S. Study of size distribution of atmospheric aerosol at Agra. *Atmos. Environ.* **2001**, *35*, 693–702.

(53) Paulot, F.; Jacob, D. J. Hidden cost of U.S. agricultural exports: Particulate matter from ammonia emissions. *Environ. Sci. Technol.* **2014**, *48*, 903–908.

(54) Alves, C. A.; Gonçalves, C.; Pio, C. A.; Mirante, F.; Caseiro, A.; Tarelho, L.; Freitas, M. C.; Viegas, D. X. Smoke emissions from biomass burning in a Mediterranean shrubland. *Atmos. Environ.* **2010**, *44*, 3024–3033.

(55) Lin, P.-C.; Wu, Z.-H.; Chen, M.-S.; Li, Y.-L.; Chen, W.-R.; Huang, T.-P.; Lee, Y.-Y.; Wang, C. C. Interfacial solvation and surface pH of phenol and dihydroxybenzene aqueous nanoaerosols unveiled by Aerosol VUV Photoelectron Spectroscopy. *J. Phys. Chem. B* 2017, *121*, 1054–1067.

(56) Rao, Y.; Subir, M.; McArthur, E. A.; Turro, N. J.; Eisenthal, K. B. Organic ions at the air/water interface. *Chem. Phys. Lett.* **2009**, 477, 241–244.

(57) Ragnar, M.; Eriksson, T.; Reitberger, T. Radical Formation in Ozone Reactions with Lignin and Carbohydrate Model Compounds. *Holzforschung* **1999**, *53*, 292.

(58) Zhou, W.; Mekic, M.; Liu, J.; Loisel, G.; Jin, B.; Vione, D.; Gligorovski, S. Ionic strength effects on the photochemical degradation of acetosyringone in atmospheric deliquescent aerosol particles. *Atmos. Environ.* **2019**, *198*, 83–88.

(59) Lauraguais, A.; Bejan, I.; Barnes, I.; Wiesen, P.; Coeur, C. Rate coefficients for the gas-phase reactions of hydroxyl radicals with a

#### The Journal of Physical Chemistry A

series of methoxylated aromatic compounds. J. Phys. Chem. A 2015, 119, 6179–6187.

(60) Sandhiya, L.; Kolandaivel, P.; Senthilkumar, K. Mechanism and kinetics of the atmospheric oxidative degradation of dimethylphenol isomers initiated by OH radical. *J. Phys. Chem. A* **2013**, *117*, 4611–4626.

(61) von Gunten, U. Ozonation of drinking water: Part I. Oxidation kinetics and product formation. *Water Res.* **2003**, *37*, 1443–1467.

(62) Flyunt, R.; Leitzke, A.; Mark, G.; Mvula, E.; Reisz, E.; Schick, R.; von Sonntag, C. Determination of  ${}^{\bullet}OH$ ,  $O_2^{\bullet}$ , and hydroperoxide yields in ozone reactions in aqueous solution. *J. Phys. Chem. B* **2003**, 107, 7242–7253.

(63) Mvula, E.; von Sonntag, C. Ozonolysis of phenols in aqueous solution. Org. Biomol. Chem. 2003, 1, 1749–1756.

(64) Bard, A. J.; Faulkner, L. R. Electrochemical Methods: Fundamentals and Applications, 2nd ed.; John Wiley & Sons, Inc.: Phoenix, 2001.

(65) Koppenol, W. H. The reduction potential of the couple  $O_3/O_3^{\bullet}$ . FEBS Lett. **1982**, 140, 169–172.

(66) Elliot, A. J.; McCracken, D. R. Effect of temperature on Oreactions and equilibria: A pulse radiolysis study. *Int. J. Radiat. Appl. Instrum., Part C* **1989**, 33, 69–74.

(67) Anglada, J. M.; Martins-Costa, M.; Francisco, J. S.; Ruiz-López, M. F. Interconnection of reactive oxygen species chemistry across the interfaces of atmospheric, environmental, and biological processes. *Acc. Chem. Res.* **2015**, *48*, 575–583.

(68) Mvula, E.; Naumov, S.; von Sonntag, C. Ozonolysis of lignin models in aqueous solution: Anisole, 1,2-dimethoxybenzene, 1,4-dimethoxybenzene, and 1,3,5-trimethoxybenzene. *Environ. Sci. Technol.* **2009**, 43, 6275–6282.

(69) Net, S.; Nieto-Gligorovski, L.; Gligorovski, S.; Temime-Rousell, B.; Barbati, S.; Lazarou, Y. G.; Wortham, H. Heterogeneous lightinduced ozone processing on the organic coatings in the atmosphere. *Atmos. Environ.* **2009**, *43*, 1683–1692.

(70) Criegee, R. Mechanism of ozonolysis. Angew. Chem., Int. Ed. 1975, 14, 745–752.

(71) Herrmann, H.; Schaefer, T.; Tilgner, A.; Styler, S. A.; Weller, C.; Teich, M.; Otto, T. Tropospheric aqueous-phase chemistry: Kinetics, mechanisms, and its coupling to a changing gas phase. *Chem. Rev.* **2015**, *115*, 4259–4334.

(72) Chen, X.; Sun, Y.; Qi, Y.; Liu, L.; Xu, F.; Zhao, Y. Mechanistic and kinetic investigations on the ozonolysis of biomass burning products: Guaiacol, syringol and creosol. *Int. J. Mol. Sci.* **2019**, *20*, 4492.

(73) Ten Brink, G.-J.; Arends, I. W. C. E.; Sheldon, R. A. The Baeyer– Villiger reaction: New developments toward greener procedures. *Chem. Rev.* **2004**, *104*, 4105–4124.

(74) O'Neill, E. M.; Kawam, A. Z.; Van Ry, D. A.; Hinrichs, R. Z. Ozonolysis of surface-adsorbed methoxyphenols: kinetics of aromatic ring cleavage vs. alkene side-chain oxidation. *Atmos. Chem. Phys.* **2014**, *14*, 47–60.

(75) Mahal, H. S.; Badheka, L. P.; Mukherjee, T. Radical scavenging properties of a flavouring agent–Vanillin. *Res. Chem. Intermed.* **2001**, 27, 595–604.

(76) Arakaki, T.; Anastasio, C.; Kuroki, Y.; Nakajima, H.; Okada, K.; Kotani, Y.; Handa, D.; Azechi, S.; Kimura, T.; Tsuhako, A.; et al. A general scavenging rate constant for reaction of hydroxyl radical with organic carbon in atmospheric waters. *Environ. Sci. Technol.* **2013**, *47*, 8196–8203.

(77) Seinfeld, J. H.; Pandis, S. N. Atmospheric Chemistry and Physics: From Air Pollution to Climate Change, 2nd ed.; Wiley, 2006; p 1232.

(78) Tomaz, S.; Cui, T.; Chen, Y.; Sexton, K. G.; Roberts, J. M.; Warneke, C.; Yokelson, R. J.; Surratt, J. D.; Turpin, B. J. Photochemical cloud processing of primary wildfire emissions as a potential source of secondary organic aerosol. *Environ. Sci. Technol.* **2018**, 52, 11027.

(79) Ehn, M.; Thornton, J. A.; Kleist, E.; Sipilä, M.; Junninen, H.; Pullinen, I.; Springer, M.; Rubach, F.; Tillmann, R.; Lee, B.; et al. A large source of low-volatility secondary organic aerosol. *Nature* **2014**, *506*, 476.