

University of Kentucky

UKnowledge

---

Chemistry Faculty Publications

Chemistry

---

5-5-2018

## Mammalian Cell-Derived Vesicles for the Isolation of Organelle Specific Transmembrane Proteins to Conduct Single Molecule Studies

Faruk H. Moonschi

*University of Kentucky*, faruk.moonschi@uky.edu

Ashley M. Fox-Loe

*University of Kentucky*, ashley.loe@uky.edu

Xu Fu

*University of Kentucky*, xufu0626@uky.edu

Christopher I. Richards

*University of Kentucky*, chris.richards@uky.edu

Follow this and additional works at: [https://uknowledge.uky.edu/chemistry\\_facpub](https://uknowledge.uky.edu/chemistry_facpub)



Part of the [Cell and Developmental Biology Commons](#), and the [Chemistry Commons](#)

**Right click to open a feedback form in a new tab to let us know how this document benefits you.**

---

### Repository Citation

Moonschi, Faruk H.; Fox-Loe, Ashley M.; Fu, Xu; and Richards, Christopher I., "Mammalian Cell-Derived Vesicles for the Isolation of Organelle Specific Transmembrane Proteins to Conduct Single Molecule Studies" (2018). *Chemistry Faculty Publications*. 149.

[https://uknowledge.uky.edu/chemistry\\_facpub/149](https://uknowledge.uky.edu/chemistry_facpub/149)

This Article is brought to you for free and open access by the Chemistry at UKnowledge. It has been accepted for inclusion in Chemistry Faculty Publications by an authorized administrator of UKnowledge. For more information, please contact [UKnowledge@lsv.uky.edu](mailto:UKnowledge@lsv.uky.edu).

---

## Mammalian Cell-Derived Vesicles for the Isolation of Organelle Specific Transmembrane Proteins to Conduct Single Molecule Studies

Digital Object Identifier (DOI)

<https://doi.org/10.21769/BioProtoc.2825>

### Notes/Citation Information

Published in *Bio-protocol*, v. 8, issue 9, p. 1-10.

Copyright © 2018 The Authors; exclusive licensee Bio-protocol LLC.

The publisher has granted the permission for posting the article here.

## Mammalian Cell-derived Vesicles for the Isolation of Organelle Specific Transmembrane Proteins to Conduct Single Molecule Studies

Faruk H. Moonschi, Ashley M. Fox-Loe, Xu Fu and Chris I. Richards\*

Department of Chemistry, University of Kentucky, Lexington, Kentucky, USA

\*For correspondence: [chris.richards@uky.edu](mailto:chris.richards@uky.edu)

**[Abstract]** Cell-derived vesicles facilitate the isolation of transmembrane proteins in their physiological membrane maintaining their structural and functional integrity. These vesicles can be generated from different cellular organelles producing, housing, or transporting the proteins. Combined with single-molecule imaging, isolated organelle specific vesicles can be employed to study the trafficking and assembly of the embedded proteins. Here we present a method for organelle specific single molecule imaging via isolation of ER and plasma membrane vesicles from HEK293T cells by employing OptiPrep gradients and nitrogen cavitation. The isolation was validated through Western blotting, and the isolated vesicles were used to perform single molecule studies of oligomeric receptor assembly.

**Keywords:** Single molecule, Vesicles, Stoichiometry, ER and plasma membrane protein separation, Nicotinic receptor, OptiPrep, Photobleaching, Protein trafficking

**[Background]** A large number of transmembrane proteins are formed through the assembly of multiple subunits leading to complicated oligomeric structures that can often exist in multiple stoichiometries. Understanding how changes in assembly alter trafficking and localization within different organelles is essential to determining a protein's physiological role and the connection to diseases associated with maturation and transport. Single molecule approaches can provide a better understanding of the assembly of oligomeric proteins by directly measuring their stoichiometry (Ulbrich and Isacoff, 2007; Richards *et al.*, 2012). This approach avoids ensemble averaging which provides the average state of all the stoichiometries (Walter and Bustamante, 2014). Single molecule studies have recently been employed to understand the structural and functional properties of macromolecules including conformational dynamics (Tan *et al.*, 2014), ion channel gating (Wang *et al.*, 2016), ligand-receptor interaction (Moonschi *et al.*, 2015), and stoichiometric assembly (Ulbrich and Isacoff, 2007; Moonschi *et al.*, 2015). To conduct single molecule studies, it is imperative to isolate single receptors in a supporting bilayer. Separation from the cellular environment is often necessary because the endogenous concentration of membrane proteins in mammalian cells is typically much higher than conditions compatible with single molecule studies (Richards *et al.*, 2012). Additionally, live cell single molecule studies suffer from a number of disadvantages including high levels of auto-fluorescence, limited fluorophore brightness and the mobility of membrane proteins on the cell surface (Andersson *et al.*, 1998; Lippincott-Schwartz *et al.*, 1999). Isolation strategies using artificial bilayers such as liposomes are also problematic as they include an intermediate step where the protein is stabilized into a detergent solution which poses a threat to the structural integrity of the transmembrane protein. Membrane-derived

vesicles enable the protein to remain embedded in its physiological membrane maintaining its structural and functional integrity. These nanoscale vesicles have been employed to study stoichiometric assembly of multimeric proteins, to probe ligand receptor interactions, and to understand the effect of ligands on the assembly of nicotinic receptors (Fox *et al.*, 2015; Moonschi *et al.*, 2015; Fox-Loe *et al.*, 2017).

Transmembrane proteins are synthesized and assembled in the endoplasmic reticulum and then trafficked to the plasma membrane. Oligomeric proteins with multiple non-identical subunits can often be assembled with different subunit stoichiometries potentially leading to different trafficking and functional properties (Grady *et al.*, 2010). For instance,  $\alpha 4\beta 2$  nicotinic receptors are pentameric receptor with two possible stoichiometric assemblies:  $(\alpha 4)_2(\beta 2)_3$  and  $(\alpha 4)_3(\beta 2)_2$ . It has been hypothesized that cellular machinery preferentially traffics the high sensitivity isoform  $(\alpha 4)_2(\beta 2)_3$  over the other isoform from the ER to the plasma membrane. It has also been hypothesized that nicotine alters the assembly of this receptor from the low sensitivity to high sensitivity isoform in the ER leading to higher levels of the preferentially trafficked assembly in both the ER and plasma membrane (Lester *et al.*, 2009; Henderson and Lester, 2015). Single molecule studies of proteins from specific organelles enable the correlation of changes in structural assembly of the nicotinic receptors to changes in trafficking and assembly.

Here we present a method which can isolate single transmembrane proteins into the ER and plasma membrane derived vesicles using nitrogen cavitation and an OptiPrep gradient. The ER and plasma membrane derived vesicles exhibit different densities because they contain different phospholipids and associated proteins. ER originated vesicles are much denser than those obtained from the plasma membrane. The OptiPrep gradient was selected because of its superior ability to maintain isosmotic pressure independent of the density of the gradient used to isolate cellular organelles and subcellular vesicles (Graham *et al.*, 1994). We applied this method to study ligand induced changes in the assembly of nicotinic receptors in the ER as well as the plasma membrane and validated our method with Western blotting and single molecule step-wise photobleaching. We believe this method can be applied for virtually any type of transmembrane proteins to conduct single molecule studies and to understand organelle-specific structural and functional properties.

## **Materials and Reagents**

1. Cell culture flasks (Greiner Bio One International, catalog number: 658175)
2. 50 ml centrifuge tubes (VWR, catalog number: 89039-658)
3. Ultra-Clear ultracentrifuge tubes (Beckman Coulter, catalog number: 344061)
4. Four 5-ml Serological pipettes (VWR, catalog number: 89130-896)
5. One 9-inch-flint-glass Pasteur pipette (VWR, catalog number: 14672-380)
6. 15 ml centrifuge tubes (VWR, catalog number: 89039-666)
7. 1.5-ml tubes
8. Wine cork
9. Razor blades (VWR, catalog number: 55411-050)

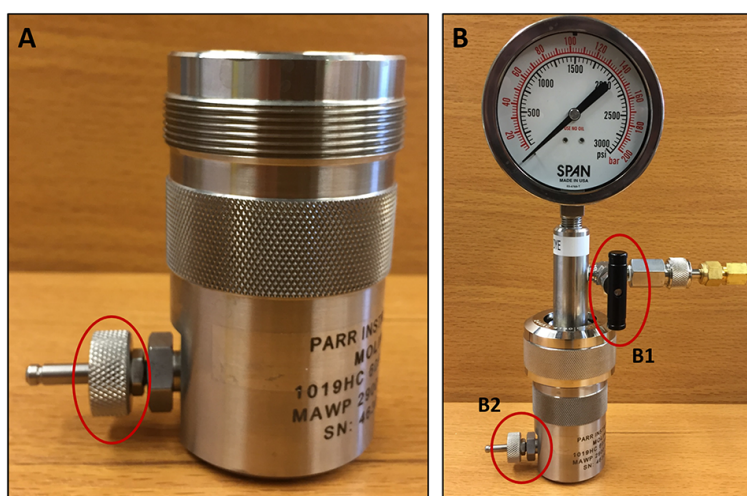
10. Two ice buckets (VWR, catalog number: 10146-202)
11. HEK293T cells (Sigma-Aldrich, catalog number: 85120602-1VL)
12. Deionized water
13. PBS buffer (VWR, catalog number: 97062-948)
14. Versene solution (Thermo Fisher Scientific, Gibco™, catalog number: 15040066)
15. Nitrogen gas
16. OptiPrep (60% solution, Sigma-Aldrich, catalog number: D1556-250ML)
17. Anti-calnexin (Abcam, catalog number: ab92573)
18. Anti-sodium potassium ATPase (Abcam, catalog number: ab76020)
19. Mouse anti-Rabbit secondary antibody (Santa Cruz Biotechnology, catalog number: sc-2357)
20. Clarity™ Western ECL Substrate (Bio-Rad Laboratories, catalog number: 1705060)
21. Tris-HCl (Sigma-Aldrich, Roche Diagnostics, catalog number: 10812846001)
22. Sodium chloride (NaCl) (Fisher Scientific, catalog number: BP358-1)
23. Magnesium chloride hexahydrate (MgCl<sub>2</sub>·6H<sub>2</sub>O) (Fisher Scientific, catalog number: BP214-500)
24. Calcium chloride (CaCl<sub>2</sub>) (Fisher Scientific, catalog number: C614-500)
25. Protease inhibitor mini tablet (Thermo Fisher Scientific, Thermo Scientific™, catalog number: A32955)
26. Sucrose (VWR, catalog number: 97063-788)
27. HEPES (Fisher Scientific, catalog number: BP310-500)
28. Hypotonic protease inhibitor solution (see Recipes)
29. Sucrose buffer (see Recipes)

## **Equipment**

1. One 1 ml pipettor (Gilson, catalog number: F123602)
2. One pipet controller (VWR, catalog number: 613-4442)
3. One pair of scissors
4. A set of stand (a stand and a clamp)
5. Incubator
6. Biosafety cabinet (Class II, Type A2)
7. Centrifuge (Beckman Coulter, model: Allerga® X-22R; Rotor: Beckman Coulter, model: SX4250)
8. Ultracentrifuge (Beckman Coulter, model: L-60)
9. Swing bucket rotor (Beckman Coulter, model: SW 28)
10. Fixed angle rotor (Beckman Coulter, model: 70 Ti)
11. Belly dancer or orbital shaker
12. Nitrogen cavitation chamber/vessel (Parr Instrument Company, catalog number: 4639)
13. Variable-Flow Peristaltic Pumps (Fisherbrand, Thermo Fisher)

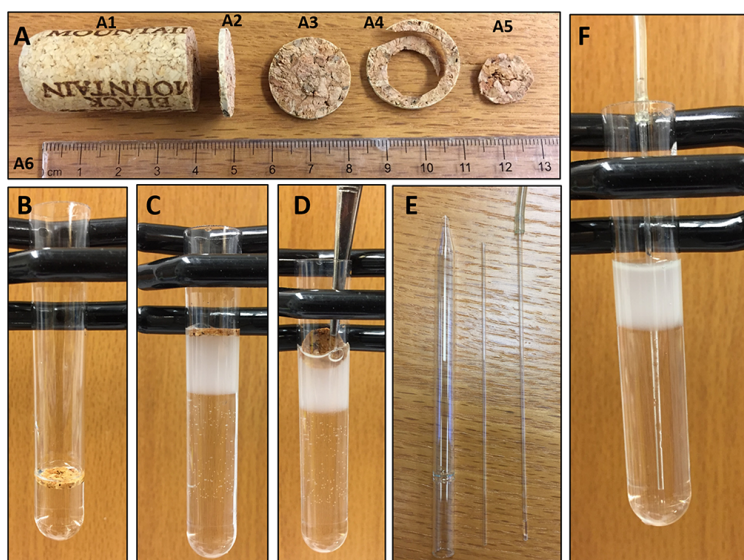
## Procedure

1. Take two flasks (250 ml, 75 cm<sup>2</sup>) of confluent HEK293T cells (about 12 million cells per flask) that are expressing a fluorescently labeled membrane protein.
2. Aspirate the media from the cells and rinse the cells with 5 ml 1x PBS buffer.
3. Add 5 ml Versene solution and incubate at 37 °C for 5 min to detach the cells from the flasks.
4. Collect the cell slurry in a 15 ml centrifuge tube.
5. Centrifuge the slurry at 300 x g for 5 min at room temperature.
6. Discard the supernatant (Versene solution) from the tube by careful aspiration and add 3 ml hypotonic protease inhibitor solution (Recipe 1) in the centrifuge tube. Resuspend the cells using pipetting in and out or swirling.
7. Place the tube in an ice bucket and shake the bucket on an orbital shaker for 10 min to facilitate swelling of the cells.
8. Transfer the cell slurry from the centrifuge tube into a pre-chilled (on ice) nitrogen cavitation chamber/vessel. Attach the vessel head and flow in nitrogen gas up to 600 psi. Incubate the cavitation chamber under pressure on ice for 20 min. Slowly open the valve to simultaneously release the pressure and collect the cell lysate into a 15 ml centrifuge tube (Figure 1). Keep the tube in ice until the cell lysate is added on top of the OptiPrep gradient.



**Figure 1. Instrumental setup used for nitrogen cavitation.** A. Take the precooled nitrogen cavitation chamber, close the valve (red circle) and add the cell slurry into the chamber. B. Attach the vessel head and open valve B1 to flow nitrogen gas inside the chamber and close valve B1 once the pressure reaches 600 psi. Place this chamber into ice for 5 min. Take the chamber out from the ice and slowly open valve B2 to simultaneously release the pressure and to collect the cell lysate into a 15 ml centrifuge tube.

9. Make the OptiPrep gradient in an Ultra-Clear ultracentrifuge tube as follows:
  - a. Cut a wine cork into ~2 mm thick circular slice using a razor blade (Figure 2A). Cut along the perimeter of the cork slice using a pair of scissors so that it can easily fit into the Ultra-clear ultracentrifuge tube. Wash the cork slice with water to remove debris. Leave it in sterile water until adding it to the ultracentrifuge tube.



**Figure 2. Pictorial presentation of the collection steps for ER and plasma membrane vesicles.** A. A wine cork (A1) is cut into a slice of about 2 mm thickness (A2). This slice (A3) is cut along the circumference to discard the outer area (A4) and keep a small circular slice (A5) which can easily fit into an Ultra-Clear ultracentrifuge tube. A scale (A6) is placed on the image for comparison of sizes. B. The small slice of cork is inserted into the tube and floats on the solution. C. As additional solution (20%, and 10% OptiPrep) is added the cork rises to the top. D. The cork slice is removed using a forceps without disturbing the gradient. E. A Pasteur pipette is broken to obtain the narrow tube which is inserted in the inlet tubing of a peristaltic pump. F. The resultant inlet tubing of the pump is inserted at the bottom-center of the Ultra-Clear tube containing the gradient solution.

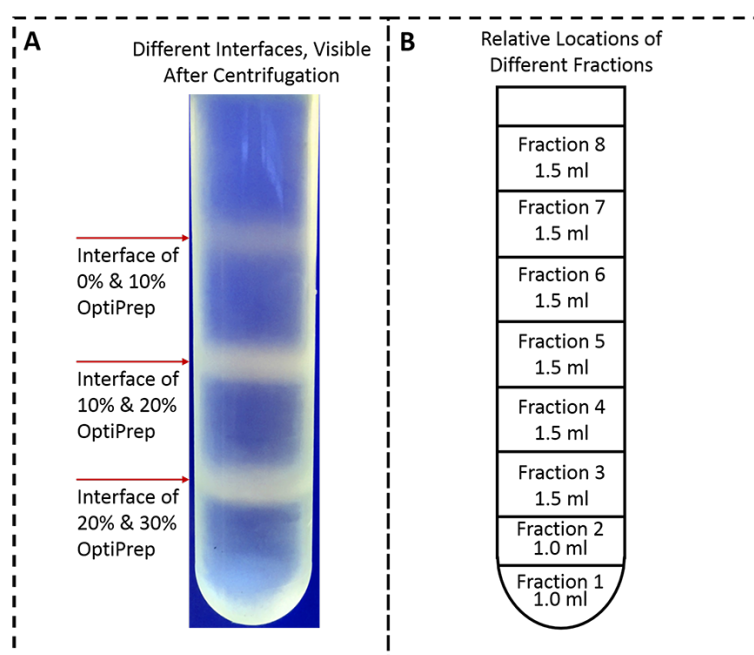
- b. Prepare 30%, 20%, and 10% OptiPrep solutions by diluting the original 60% OptiPrep solution with sucrose buffer (Recipe 2) into three 50 ml tubes. Calculations associated with the preparation of 30 ml of each solution can be found in Table 1.

**Table 1. Preparation of 30, 20 and 10% OptiPrep solution from 60% OptiPrep solution with sucrose buffer.** The original OptiPrep solution obtained from the supplier contains 60% OptiPrep.

To prepare	Volume of OptiPrep solution (60%)	Volume of sucrose buffer	Total volume
30% OptiPrep	15 ml	15 ml	30 ml
20% OptiPrep	10 ml	20 ml	30 ml
10% OptiPrep	5 ml	25 ml	30 ml

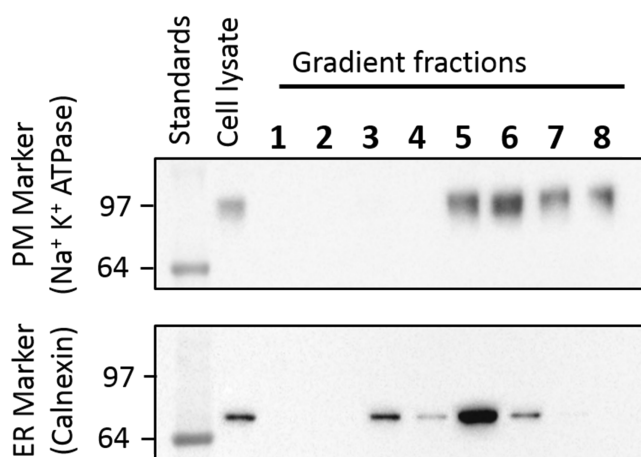
- c. In an Ultra-Clear ultracentrifuge tube, using a 5-ml Serological pipette and a pipette controller, add 3 ml 30% OptiPrep solution, then add the cork slice on top of the solution using a forceps (Figure 2B). Now add dropwise 3 ml 20% OptiPrep, then 3 ml 10% OptiPrep and finally 3 ml cell lysate on top of the cork slice. With the dropwise addition of solution in the tube, the cork slice stays floating on top of the solution and thus helps to avoid mixing of the added solution with the solution already present in the tube (Figure 2C).
- Note: Three distinct interfaces should be visible indicating 4 layers of solutions of different densities. The interfaces are clearly visible when the tube is placed against a light source.*
- d. Carefully remove the wine cork using a forceps without disturbing the gradient (Figure 2D).
10. Centrifuge the ultracentrifuge tube containing OptiPrep gradient in a swing bucket rotor at 25,000 rpm (112,000 x g) for 90 min at 4 °C.
11. After the centrifugation, three layers of vesicles should be observable: between cell lysate and 10% OptiPrep, 10% and 20% OptiPrep, and 20% and 30% OptiPrep (as shown in Figure 3).





**Figure 3. Presence of vesicles between the interfaces of OptiPrep solutions and the relative positions of different fractions collected.** A. Three layers of vesicles are visible: on the interface of 0% and 10% OptiPrep, 10% and 20% OptiPrep and 20% and 30% OptiPrep. This image was taken with a mobile phone camera. B. By inserting the inlet tubing of a peristaltic pump in the bottom-center of the tube, collect fractions 1 and 2 in series, 1.0-ml each, then fractions 3-8 in a series, 1.5 ml each.

12. Use a peristaltic pump to collect ER and PM fractions as follows:
  - a. Place the Ultra-Clear centrifuge tube vertically using a stand and a clamp.
  - b. Break the glass Pasteur pipette at the junction of the small-diameter tube and large diameter tube and insert the small-diameter tube inside the inlet tubing of the peristaltic pump (Figure 2E).
  - c. Carefully, insert the inlet tube of the peristaltic pump in the ultracentrifuge tube (fixed with a clamp and stand) until it touches the bottom-center of the centrifuge tube (Figure 2F). It is necessary to make sure that the gradient is not disturbed by the insertion process.
  - d. Start the peristaltic pump to collect different fractions keeping the interfaces in the same fraction (Figure 3B). To do so, collect fractions 1 and 2, 1 ml each and fractions 3 to 8, 1.5 ml each.
  - e. According to our Western blot results with anti-calnexin as an ER marker and anti-sodium potassium ATPase as a plasma membrane marker, fractions 3 and 4 contain only the ER and fractions 7 and 8 include only the plasma membrane vesicles (Figure 4). However, we have selected fractions 3 and 7 as the ER and plasma membrane vesicles respectively for our experiments.



**Figure 4. Western blot of the different fractions collected from OptiPrep gradient to isolate single receptors into the ER and plasma membrane specific vesicles.** Anti-sodium potassium ATPase antibody (1:200 dilution) was used to bind with the plasma membrane marker, sodium potassium ATPase, in the OptiPrep gradient (upper panel). An anti-calnexin antibody (1:200 dilution) was employed to bind with the ER marker, calnexin, in the gradients (lower panel). An HRP conjugated mouse anti-Rabbit secondary antibody (1:5,000 dilution) and Clarity™ Western ECL Substrate on a ChemiDoc imaging system (Bio-Rad) were utilized to locate the ER and Plasma membrane markers on the gels. The plasma membrane marker was found in the cell lysate and fractions 5 to 8 and the ER marker was located in the cell lysate and fractions 3 to 6. Therefore, fractions 3 and 4 contain only the ER vesicles; fractions 7 and 8 contain only the plasma membrane vesicles. We selected fraction 3 for ER vesicles and fraction 7 for the plasma membrane vesicles.

13. Dilute the ER and the plasma membrane fractions with 1x PBS buffer to 1:3 and centrifuge at 30,000 rpm (100,000 x g) for 1 h at 4 °C with the fixed angle rotor.
14. Resuspend the pellets each in 600 µl 1x PBS buffer or sucrose buffer to obtain ER and plasma membrane derived vesicles. We usually aliquot the vesicle solution into two or three 1.5-ml tubes and store at -80 °C. We use the vesicles within 7 days.

*Note: If a fraction contains no vesicles or a very small amount of vesicles, there will be no visible pellet on the surface of the tube after centrifugation. In this case, we collected the sample from the area where the vesicles might have present. It can be noted that a pellet is formed on the surface of the tube furthest from the center of the rotor and the possible location of the pellet can easily be assessed based on the location of the visible pellets in the other tubes.*

At this point, a fraction of the ER and plasma membrane derived vesicles will contain the protein of interest. For single molecule fluorescence studies, it is necessary to start with cells expressing the target protein conjugated with a fluorescent protein. Single receptor containing vesicles can be employed to study the stoichiometric assembly of the receptors by step-wise photobleaching technique, to probe ligand-receptor interaction using fluorescence correlation spectroscopy and

to understand dynamics and conformational changes of the receptor using single molecule Forster resonance energy transfer (smFRET).

### **Recipes**

1. Hypotonic protease inhibitor solution  
10 mM Tris-HCl  
10 mM NaCl  
1.5 mM MgCl<sub>2</sub>  
0.2 mM CaCl<sub>2</sub>  
Adjust pH to 7.4  
One protease inhibitor tablet per 10 ml buffer
2. Sucrose buffer  
250 mM sucrose  
10 mM HEPES  
Adjust pH to 7.4

### **Acknowledgments**

This work was supported in part by the NIDA training grant DA016176 (AMF) and NIDA DA038817. This work was adapted from Fox-Loe *et al.*, 2017, *J Biol Chem* 292(51): 21159-21169. We declare no conflict of interest or competitive interest related to this publication.

### **References**

1. Andersson, H., Baechi, T., Hoechl, M. and Richter, C. (1998). [Autofluorescence of living cells.](#) *J Microsc* 191(Pt 1): 1-7.
2. Grady, S. R., Drenan, R. M., Breining, S. R., Yohannes, D., Wageman, C. R., Fedorov, N. B., McKinney, S., Whiteaker, P., Bencherif, M., Lester, H. A. and Marks, M. J. (2010). [Structural differences determine the relative selectivity of nicotinic compounds for native  \$\alpha 4\beta 2^{\*}\$ -,  \$\alpha 6\beta 2^{\*}\$ -,  \$\alpha 3\beta 4^{\*}\$ - and  \$\alpha 7\$ -nicotine acetylcholine receptors.](#) *Neuropharmacology* 58(7): 1054-1066.
3. Graham, J., Ford, T. and Rickwood, D. (1994). [The preparation of subcellular organelles from mouse liver in self-generated gradients of iodixanol.](#) *Anal Biochem* 220(2): 367-373.
4. Fox, A. M., Moonschi, F. H. and Richards, C. I. (2015). [The nicotine metabolite, cotinine, alters the assembly and trafficking of a subset of nicotinic acetylcholine receptors.](#) *J Biol Chem* 290(40): 24403-24412.
5. Fox-Loe, A. M., Moonschi, F. H. and Richards, C. I. (2017). [Organelle-specific single-molecule imaging of  \$\alpha 4\beta 2\$  nicotinic receptors reveals the effect of nicotine on receptor assembly and cell-surface trafficking.](#) *J Biol Chem* 292(51): 21159-21169.

6. Henderson, B. J. and Lester, H. A. (2015). [Inside-out neuropharmacology of nicotinic drugs.](#) *Neuropharmacology* 96(Pt B): 178-193.
7. Lester, H. A., Xiao, C., Srinivasan, R., Son, C. D., Miwa, J., Pantoja, R., Banghart, M. R., Dougherty, D. A., Goate, A. M. and Wang, J. C. (2009). [Nicotine is a selective pharmacological chaperone of acetylcholine receptor number and stoichiometry. Implications for drug discovery.](#) *AAPS J* 11(1): 167-177.
8. Lippincott-Schwartz, J., Presley, J. F., Zaal, K. J., Hirschberg, K., Miller, C. D. and Ellenberg, J. (1999). [Chapter 16: Monitoring the dynamics and mobility of membrane proteins tagged with green fluorescent protein.](#) In: Sullivan, K. F. and Kay, S. A. (Eds.). *Methods in Cell Biology. Academic Press* 261-281.
9. Moonschi, F. H., Effinger, A. K., Zhang, X., Martin, W. E., Fox, A. M., Heidary, D. K., DeRouchey, J. E. and Richards, C. I. (2015). [Cell-derived vesicles for single-molecule imaging of membrane proteins.](#) *Angew Chem Int Ed Engl* 54(2): 481-484.
10. Richards, C. I., Luong, K., Srinivasan, R., Turner, S. W., Dougherty, D. A., Korlach, J. and Lester, H. A. (2012). [Live-cell imaging of single receptor composition using zero-mode waveguide nanostructures.](#) *Nano Lett* 12(7): 3690-3694.
11. Tan, Y. W., Hanson, J. A., Chu, J. W. and Yang, H. (2014). [Confocal single-molecule FRET for protein conformational dynamics.](#) *Methods Mol Biol* 1084: 51-62
12. Ulbrich, M. H. and Isacoff, E. Y. (2007). [Subunit counting in membrane-bound proteins.](#) *Nat Methods* 4(4): 319-321.
13. Wang, S., Vafabakhsh, R., Borschel, W. F., Ha, T. and Nichols, C. G. (2016) [Structural dynamics of potassium-channel gating revealed by single-molecule FRET.](#) *Nat Struct Mol Biol* 23: 31
14. Walter, N. G. and Bustamante, C. (2014) [Introduction to single molecule imaging and mechanics: seeing and touching molecules one at a time.](#) *Chem Rev* 114(6): 3069-71.