2015

A Therapeutic Approach for Senile Dementias: Neuroangiogenesis

Charles T. Ambrose
University of Kentucky, cambros@uky.edu

Click here to let us know how access to this document benefits you.

Follow this and additional works at: https://uknowledge.uky.edu/microbio_facpub

Part of the Geriatrics Commons, Molecular Genetics Commons, and the Neurology Commons

Repository Citation
Ambrose, Charles T., "A Therapeutic Approach for Senile Dementias: Neuroangiogenesis" (2015). Microbiology, Immunology, and Molecular Genetics Faculty Publications. 110.
https://uknowledge.uky.edu/microbio_facpub/110

This Article is brought to you for free and open access by the Microbiology, Immunology, and Molecular Genetics at UKnowledge. It has been accepted for inclusion in Microbiology, Immunology, and Molecular Genetics Faculty Publications by an authorized administrator of UKnowledge. For more information, please contact UKnowledge@lsv.uky.edu.
Alzheimer’s disease (AD) and related senile dementias (SDs) represent a growing medical and economic crisis in this country. Apart from cautioning persons about risk factors, no practical, effective therapy is currently available. Much of the recent research in AD has been based on the amyloid cascade theory. Another approach assumes a vascular basis for the senile dementias. This paper presents evidence from a score of studies that cerebral capillary density (CCD) declines during old age in animals and people and also in AD. Neuroangiogenic (NAG) factors initiate and maintain capillaries in the brain. Thus a waning level of these factors and the ensuing declining CCD would lead to local areas of reduced oxygen and glucose and result in impaired synaptic and neuronal function. The NAG hypothesis developed here proposes that the age-linked decline in CCD is a terminal condition in SDs, including many cases of AD. This age-linked decline is independent of any other of the various pathologies proposed as causing AD and listed in Table 1. Waning NAG factors would render the SDs a deficiency condition, somewhat like falling androgen levels in aging males. A logical corollary of this hypothesis is that chronic replacement therapy with recombinant forms of NAG factors may arrest the age-linked decline in CCD and prevent further loss of memory and mental deterioration. A transnasal route of therapy seems the most practical one for general use in the large aging populations.

X. KEY WORDS:
neuroangiogenesis, cerebral capillary density, senile dementia, Alzheimer’s disease, cognitive impairment
A THERAPEUTIC APPROACH FOR SENILE DEMENTIAS:

NEUROANGIOGENESIS

1. INTRODUCTION

The absence of an effective treatment for Alzheimer’s disease (AD) and other senile dementias continues notwithstanding the many thousands of studies on these medical conditions during the past half century. The diverse range of these investigations is shown in Table 1. Recently, Jack C. de la Torre employed “evidence based meta-analysis from standardized control studies” to determine whether the cause of AD could reasonably be ascribed to any of seven “more cited hypothetical proposals” [1]. The seven are indicated with an asterisk in the Table 1. Koch’s postulates were used as the framework for his assessments. De la Torre judged none of the seven completely satisfactory, although he found “partial positive support” for oxidative stress and the various vascular hypotheses. Dominating investigations over twenty years has been the amyloid cascade theory of Hardy and Higgins [2, 3]. Numerous clinical trials based on it and other proposed etiologies of AD have been carried out, but “none of which has yet achieved robust slowing of cognitive decline” [4]. A recent paper on AD suggested the obvious that “New therapeutic approaches are desperately needed” [5].

In medicine, the usual response to an unresolved clinical problem is to say that “more research is required.” However, an examination of the literature on AD, other senile dementias, and aging may itself be productive. Buried there and in unrelated reports on anatomy and
pharmacology are abundant data showing that a reduction in the cerebral capillary bed occurs in rats and mice during old age and commonly begins in people during late adulthood. This reduction has also been found in subjects with AD, most of whom, of course, were aged. Various changes in the capillaries discussed later have led investigator to suggest that a defective cerebral capillary bed could cause synaptic and neuronal dysfunction and ultimately produce abnormalities in memory, behavior, and general cognition.

The thesis of this essay is that the age-linked reduction in capillary numbers in various areas of the brain contributes to senile dementias, including possibly also AD. This idea is further developed in the neuroangiogenic (NAG) hypothesis, which proposes that the reduced cerebral capillary density (CCD) is due to the age-linked declining levels of vascular trophic factors in these areas. This hypothesis thus points to an untried therapeutic approach – restoring levels by administering recombinant neuroangiogenic factors.

Figures cited for the recent and anticipated incidence of AD in the United States are 5.4 million in 2011, 7.7 million in 2030, and 16 million in 2050 [6]. This growing crisis involves more than this one eponymic disease (AD) and includes the group termed vascular dementias (VaD) [6]. They overlap so much in clinical features and neuropathology that “a rigid distinction between AD and VaD is no longer tenable” [7]. Many others concur [8-11]. In this essay, both will often be considered collectively as the senile dementias (SDs) in order to emphasize a common vascular pathology. The generic term senile dementia has been viewed as a symptom complex of sixty disorders, of which 50-60% are AD, 10-20% VaD, and the rest include brain tumors, paresis of neurosyphilis, AIDS, toxins, and metabolic disorders. Again, in this essay, the term “senile dementia” includes AD and VaD but none of the rest.
The NAG hypothesis is not a rival to various theories about the etiology of AD, such as amyloidopathy, neurotransmitter deficits, multiple microinfarcts, and others listed in Table 1. This hypothesis proposes that in AD, an age-linked reduced cerebral capillary circulation may occur secondarily (or be in addition) to other brain pathology. Also, in many elderly persons devoid of any significant evidence of AD, a defective microcirculation alone may be a primary cause of synaptic dysfunction and/or neuronal loss and the ensuing cognitive decline.

This essay focuses first on the cerebral microcirculation and points out an error accounting for some discordant capillary density data in the literature. A restatement of the NAG hypothesis is followed by information on age-linked levels of angiotrophic factors. The concept of neurological/cognitive reserve is introduced and may explain a clinical-histological paradox in AD. The concluding sections center on the potential treatment of SDs with NAG factors.

2. CEREBRAL VASCULAR STUDIES

In 1984 Arnold B. Scheibel wrote that in persons with “presenile and senile dementia of the Alzheimer type … the diminished cerebral blood flow [CBF] is more or less proportional to the degree of mental impairment … loss of verbal ability … [and] verbal memory decrease” [12]. CBF and cerebral hypoxia have been implicated in SDs by many investigators [13-22]. A reduced CBF has been blamed for a lowered learning ability in older rats [23,24]. Such measurements reflect conditions of both the large afferent vessels largely outside the brain and cerebral capillaries in the brain parenchyma.

**Upstream.** Chronic impairment of the circulation may occur in arteries and arterioles altered by risk factors, such as atherosclerosis, hypertension, cardiac arrhythmias, carotid artery
disease, smoking, diabetes, etc. [22,25,26]. In 2004, de la Torre advanced a vascular hypothesis in which such risk factors sequentially induce brain hypoperfusion, a neuroglial energy crisis, and ultimately neurodegeneration beginning with mild cognitive impairment and ending with vascular dementia (VaD) and AD [27].

**Downstream.** Investigators considering a microvascular basis for AD have focused on the blood brain barrier or qualitative abnormalities in the cerebral capillaries [28-34]. Several groups proposed that amyloid deposits in small cerebral vessels could impair the delivery of essential nutrients to cerebral neurons [35,36]. Over the years Zlokovic has advanced multifactorial hypotheses, which invoked faulty clearance of amyloid β peptide (Aβ) across the blood-brain barrier, oxidative stress by neurotoxic Aβ peptides, and other processes -- all ultimately leading to synaptic and neuronal dysfunction [37].

As early as 1974, Hachinski et al. proposed that multiple cerebral infarcts produce mental deterioration in the elderly [38]. Three reviews in 2012 summarized many reports associating cognitive impairment with cerebral microinfarcts [39-41]. Their etiology has been variously ascribed to an embolic origin, intermittent focal ischemic episodes, or cerebral amyloid amyloidopathy [41]. Cullen et al. related amyloid plaque and tangle densities to microhemorrhages and cognitive loss [42].

Scheibel prefaced his article on aging and dementia cited above with “A neuron is no better than the vessel that nurtures it” [12]. Apart from the microhemorrhages above noted, cerebral microvascular abnormalities in AD have often been described in picturesque terms (see below, Section 3). Largely overlooked have been the age-linked decline in the number of cerebral capillaries. While amyloid deposits, microinfarcts, and microvessel distortions are
rather easy to observe and count, cerebral capillary density (CCD) has been until recently more
difficult to measure (see later, Section 4).

3. INCREASED CEREBRAL CAPILLARY ABNORMALITIES

*In aged persons.* In 1873, three decades before Alois Alzheimer described the first case
of AD, J. Batty Tuke had reported that the brains of many insane persons were full of deformed
blood-vessels showing “undue straightness, tortuosity, and kinking” [43]. Although their ages
were not stated, many of these institutionalized patients were likely elderly and some may have
been cases of AD. In 1967, a century after Tuke’s report, Hassler observed glomerular loop
formations, vascular bundles, and vascular wickerworks in the arteries of brain slices from the
frontal and occipital lobes [44]. In 231 autopsies selected at random, he found these changes to
be common the brains of persons over age 65 but uncommon in those younger than age 50. He
theorized that such “deformities are responsible for the slow cerebral circulation in senile
dementia.” In 1978 Ravens described similar vascular alterations in senile brains at the level of
small arteries, arterioles, and pre-capillaries [45]. Wilkinson et al. also ascribed the reduced
CBF to vascular resistance in the deformed capillaries [46].

*In AD.* In a 1993 article entitled “Can disturbed brain microcirculation cause
Alzheimer’s disease?,” de la Torre and Mussivand focused on cerebral capillaries distorted by
amyloid deposits and a thickened basement membrane [35]. They cited 13 reports of cerebral
vascular abnormalities found at autopsy in AD and other forms of dementia and reasoned that
any disturbed blood flow would reduce delivery of oxygen and glucose to cerebral neurons and
possibly lead to AD. A decade later, Bailey et al. described various distortions of microvessels
in AD brains -- string vessels, glomerular loop formation, and twisted or tortuous vessels -- and
reiterated the view that microvascular changes “may precede the cognitive decline and neuropathological changes of AD” [47].

Among the 13 reports mentioned above was one by Fisher et al., who had observed a striking increase in tortuous and looped arterioles and capillaries in brain sections of AD subjects versus controls [48]. The average counts for distorted capillaries in 6 control subjects (ages 23-90) and 16 subjects with AD (ages 75-92) in various areas of the brain were as follows for controls vs. aged: prefrontal cortex: 213 vs. 657, basal forebrain: 167 vs. 916, hippocampus: 230 vs. 867, and motor/sensory cortex: 218 vs. 908. The same study revealed a “statistically significant reduction in the vascular net density” (reviewed in the next section) and thus suggests a temporal connection between emerging capillary distortions and a later declining cerebral capillary density.

4. DECREASED CEREBRAL CAPILLARY DENSITY  (Tables 2 and 3)

*CCD in aged rats.* In the 1920s Edward Horne Craigie pursued studies on quantitative morphometry of rat brain capillaries. He measured the length of each capillary segment present in a fixed number of adjacent brain sections and interpreted the sum of the lengths found in this volume of brain tissue as an index of the cerebral capillary density. In aged rats he found decreased CCD in two areas of the cerebral cortex – *regio insularis* and *regio temporalis* [49]. For example, the CCD in the *regio insularis* of five-month old rats was 856 µ, while that of 13-month old rats was only 638 µ. In the *regio temporalis* the comparable values were 958 µ and 880 µ. See Table 2. (Here and below in comparisons of CCDs, the average figure for the older members of a set is in bold italicized type.)
Other early investigators counted capillaries by various systems which yielded numbers unique to their particular study and comparable only within it. Later studies have employed computerized image analysis [50-52]. For example, Amenta et al. used alkaline phosphatase histochemistry with an ASBA image analyzer (Leica, Cambridge, UK). They determined the sum of capillary lengths in 500 µm² sections in the frontal cortex of 12- and 27-month old male Wistar and found values 190 µm and 168 µm, respectively [53]. In this same investigation, the older rats had comparable capillary reductions in the occipital cortex and other brain areas.

Table 2 of this essay lists age-related reductions in CCD from eleven studies in rats and one each in mice and Macaque monkeys [34,45,49,54-63].

**CCD in aged persons.** Reduced CCDs have also been reported in aged persons. In Table 3, the top five sets of data (#14-18) illustrate the capillary density in human brains at different ages in different areas: frontal and temporal cortices, paraventricular nuclei of the hypothalamus, and the hippocampus [64-69]. For example, Mann et al. reported the “mean capillary measure” in the frontal cortex of mentally normal subjects. In six adults ages 26-58 the average value was 965.6 µm ±132, while in nine older subjects age 76-96 it was 782.6 µm ± 56 [64]. The other four reports in this table show a comparable reduction with aging.

Parenthetically, the microcirculation in the aged retina parallels that in the aged brain. Kuwabara and Cogan noted “a loss of endothelium … in the capillaries of the peripheral retina in persons past middle life (50 years)” [70].

**CCD in AD.** In Table 3, data in the bottom five sets of figures (#16-20) indicate decreased CCDs in the brains of persons with AD compared with other age-paired normal controls [47,66-69]. For example, Fisher et al. found in the basal forebrain of six normal subjects age 23-90 an average “vascular density index” of 86.8, while sixteen persons with Alzheimer’s
disease, age 76-92 yielded an index value of 42.7 [47]. Kitaguchi et al. measured the cerebral capillary density for six controls (age 74 ± 4) and eight AD subjects (age 79 ± 12) in two areas. The control and AD values were, respectively, 28 and 21 in the frontal cortex and 28 and 20 in the parietal cortex [69].

Kitaguchi’s study also reported that the average weight of the control group brains was 1244 g ± 57, while that of the AD brains was only 1020 g ±11 (p<0.05). Because of significant brain atrophy in the AD group, the authors concluded that the actual reduction in the capillary density “may be more severe” – i.e., less than the above AD values of 21 and 20.

5. POTENTIAL ERRORS IN CCD MEASUREMENTS

As suggested above and noted by others, cerebral atrophy/shrinkage may mask capillary loss and represents a source of error when comparing CCD measurements in young and old brains [45,66]. A simple formula, \( \text{CCD} = \frac{N}{V} \), will illustrate the issue. CCD is determined from a particular capillary count (N) within a fixed volume of brain tissue (V). If V is reduced as a result of shrinkage and N stays unchanged, then CCD will appear falsely high. Neglecting to consider brain atrophy may explain why some studies have reported no reduction in CCD with aging, “nonsignificant trends,” or even increased CCDs [71-73]. Cortical volume shrinkage has also been extensively discussed in papers concerning neuron numbers in normal aging and AD [74-77]. The parameters of such shrinkage are discussed below.

_Brain shrinkage in aged rats._ Smith reported the brain volume in rats of middle vs. old age. The average volume of the left hemisphere of one-year old rats is 226.6 cc and of two-year olds is 205.6 cc. (9% loss) [78]. Wilkinson et al. concluded that the age-related changes in the vasculature of the rat cerebral cortex reflect parenchymal atrophy and are “not uniformly
distributed throughout the cortex” [45]. Others have confirmed that the percentage shrinkage varies in different regions of the brain [75,76,79-82]. This variation may account for Craigie’s finding in aged rats a reduced average capillary density in two areas of the cerebral cortex but not in three others, where shrinkage may have occurred and yielded falsely elevated values [49].

**Brain shrinkage in aged persons.** Appel and Appel reported that the mean weight of 2080 (sic) non-lesioned human brains decreased by 11% between the ages 25 and 96 [83]. Smith determined that the volume of one hemisphere measures on average at age 30 years 281 cc and at age 60 years 255 cc. (9% loss) [84]. Jernigan et al. found volume loss between the ages of 30 and 90 to be 14% in the cerebral cortex, 26% in the cerebral white matter, and 35% in the hippocampus [85]. Age-related shrinkage of the hippocampus has been noted by others [86-88].

### 6. THE NEUROANGIOGENIC (NAG) HYPOTHESIS

In a 2010 paper, I proposed that the remarkable coordinated finger/hand/foot dexterity displayed by concert pianists was due to additional synaptic connections in the primary motor cortex supported by an increased local capillary density, which in turn was initiated and maintained there by local neuroangiogenesis [89]. I am not aware of any modern histological or imaging studies on the brains of highly proficient pianists. However, in right-handed string musicians “the cortical representation of the fingers of the left hand … [is] larger than that in controls” [90]. Also magnetic resonance imaging reveals that very proficient jugglers develop in their mid temporal area an expanded gray matter volume which persists enlarged so long as they maintain this skill [91]. Similarly, London taxicab drivers show an increased volume of gray matter in the posterior hippocampus during their years of continued mastery of the city’s 25,000 streets but not after their retirement [92]. These transiently expanded areas of gray matter likely
reflect the sequence of increased synaptic connections supported by local microcirculation
induced by an augmented neuroangiongenesis, as I proposed for the motor cortex of concert
pianists. There are supporting data for this sequence in animal studies, as reviewed elsewhere
[89]. Interest in this area alerted me to the research of Craigie and others showing a waning
CCD in aged animals. These led me logically to reports of older persons in whom the CCD had
declined – specifically, those with AD and senile dementia.

From this study of the neurological literature came the NAG hypothesis, which concerns
SDs in general. The hypothesis involves the two key ideas introduced earlier: 1) the age-related
decline in cerebral capillary density in certain areas of the brain and 2) reduced levels there of
neuroangiogenic factors. Thus SDs may be viewed as a deficiency disorder – analogous to low
testosterone in some older men, for whom androgen therapy is corrective. And as alluded
earlier, this hypothesis infers a specific therapy for SDs – i.e., exogenous NAG factors
administered chronically to aging persons [93,94].

The NAG hypothesis also suggests that an age-associated down-regulation of genes
coding for these factors varies in different parts of the brain and thus may direct where local
CCD decline first occurs and later continues. This determines the course and character of senile
dementia in a given person – for example, a weakened declarative memory (hippocampus, etc.)
or disturbed emotions (amygdala) or impaired executive functions (prefrontal cortex, temporal-
parietal association cortices), etc. Senile dementia commonly begins with memory loss and later
emotional and personality changes but not always in this order [95]. Brun and Gustafson
described a common pattern of “amnesia, aphasia, apaxia, spatial dysfunction, and agnosia” [96].
Motor and sensory functions are not commonly affected early in the course of AD unless infarcts
occur [97].
Relevant here is a view about cerebral capillaries which is the reverse of the role given them in the NAG hypothesis. Again, the hypothesis proposes that the age-linked reduction of these capillaries leads to a local deficiency of glucose and oxygen and to the subsequent neuronal impairment/death and synaptic dysfunction. A different view is that in AD the death of neurons (from various causes) would lessen the local demand for oxygen and glucose and account for a reduced microcirculation. Capillaries are formed and maintained by angiogenic factors. I do not understand the biological mechanism by which a lessened demand could reduce neuroangiogenesis. In cases of senile dementia not involving AD (and neuronal death) but due to defective synapses there is reduced CCD. Also, capillary density declines with age in other tissues, as discussed later in Section 8. But this controversy over the fate of capillaries in the brain seems moot in light of the therapy proposed by the NAG hypothesis. Regardless of whether a waning CCD is a primary or secondary event in senile dementias, a reduced capillary density may be restored by the administration of exogenous, recombinant NAG factors.

7. ANGIOGENESIS IN GENERAL

In higher animals, angiogenic/endothelial trophic factors are responsible for vasculogenesis (new blood vessels from angioblasts) during early development and for angiogenesis (new capillaries from preexisting vessels) later in life. Details about the factors have been amply reviewed elsewhere [98-103]. Their individual names indicate their target or their cellular origin. A major one is the vascular endothelial growth factor (VEGF), which will be mentioned frequently later in this essay [104]. A review on angiogenesis by Felmeden et al. listed the major groups of growth factors involved as vascular (VEGF-B, -C, -D, -E), fibroblast (FGF-1, -2) and angiopoietin (-1 & -2), but other may contribute – e.g., placental growth factor
(VEGF-A), platelet derived growth factor (PDGF), transforming growth factor (TGFβ), insulin like growth factor (ILGF), and others [105]. Krupinski et al. identified the three most important trophic factors “especially relevant to brain angiogenesis” as the vascular endothelial growth factor (VEGF), endothelial cell growth factor-β (ECGFβ), and transforming growth factor-β (TGFβ) [106]. Other factors and their mechanism have been reviewed elsewhere [107].

As will be noted in a later section (#9), comparable measurements of NAG factors in the brains of mature vs old animals and adult vs. aged persons are not yet available. But direct evidence for angiogenesis declining with age has been found in muscles of rabbits and mice by Rivard et al. [108]. A summary of this work follows because of its relevance to the NAG hypothesis.

8. ANGIOGENESIS IN MUSCLES

Rivard et al. investigated “the hypothesis that angiogenesis is impaired as a function of age” [108]. Local ischemia in areas throughout the body stimulates angiogenic factors present to expand the capillary circulation there. The authors made one hind limb ischemic in young and old laboratory animals by resection of the femoral artery and at a later time measured the capillary density in the affected leg muscle. They used alkaline phosphatase to stain capillaries in muscle sections prepared from six young rabbits (age 6-8 months) and seven old rabbits (age 4-5 years) 10 days and 40 days after resection of one femoral artery. Collateral vessel development was reflected in angiographic scores, which on Day 10 and Day 40 were for the young 0.32 rising to 0.68 and in the old 0.43 rising to 0.48. In summary, on Day 10 scores “were similarly low” for both age groups (0.32 and 0.43) but by Day 40 had risen to 0.68 in the young rabbits but only 0.48 in the old. Also on Day 40, the average capillary count per mm² for the
young rabbits was 170 and for older ones was only 130, indicating a lower level of angiogenesis in the older animals. (The data cited above and below have been extrapolated from Figure 4E & 4F and 5G & 5H of the Rivard paper.)

In a comparable study in mice, unilateral hind limb ischemia was created in young and old animals. Capillaries of muscle sections were stained by an immunohistochemical method using CD31, an endothelial cell adhesion molecule. At 28 days after surgery, the average capillary density was 710 for the 12-week old mice and only 350 for the 2-year old ones. The above data show decreased capillaries in the muscles of both aged rabbits and mice vs. those in young animals. These findings in leg muscles parallel the data in Tables 2 and 3 concerning CCDs in the brains of older animals and aging people.

The authors also measured VEGF mRNA expression in ischemic hind limbs and found that it was reduced in the muscles of both old rabbits and old mice compared with levels observed in young controls. This reduction is that expected for NAG factors in the brains of some older persons with impaired cognition.

Finally, in this study by Rivard et al., 14 rabbits with unilateral hind limb ischemia (seven 6-8 months old and seven 4-5 years old) were injected with recombinant human vascular endothelial growth factor (rh VEGF) on postoperative day 10 via the iliac artery of the ischemic limb. The average capillary densities of young and old rabbits were determined on Day 10 (just before treatment) and on Day 40. Among the young rabbits the values were 170 (on Day 10) vs. 280 (on Day 40 = 64% increase). Among the old rabbits they were 130 (on Day 10) vs. 190 (on Day 40 = 46% increase). Thus, in both age groups the administration of VEGF increased the capillary density. The authors commented that “Advanced age … does not preclude
augmentation of collateral vessel development in response to exogenous angiogenic cytokines.”
Whether these findings can be extrapolated to human subjects is of prime interest in this essay.

9. **NEUROANGIOGENESIS**

Again, there are little direct data on NAG factors in the brain to parallel those figures reported above for VEGF mRNA expression in ischemic muscles of older rabbits and mice. But there is a general consensus about the presumed importance of such factors in the brain. For example, in 1986 Bär et al. wrote that the maintenance or repair of cerebral capillaries in rats “needs a continuous action of an angiogenetic stimulus” [109]. In 1992, Ferrara et al. suggested that the “presence of the growth factor may be required to maintain the differentiated state of those vessels [rat cerebral capillaries], which otherwise might undergo involution” [110]. And a decade later, when writing about angiogenesis in general, Carmeliet also concluded that “threshold levels of VEGF [are needed] for the survival and maintenance of quiescent vessels in healthy organs” [111].

*NAG in rats.* Several research groups have determined levels of VEGF mRNA during the immediate postnatal period in rats [112,113]. To my knowledge, levels during later ages have not been reported in rodents, except for the study in mice (and rabbits) by Rivard et al. noted above. Nonetheless, such levels can be inferred in one notable study comparing young vs. older rats challenged under different conditions. De la Torre developed rat models of chronic hypoperfusions in which visuospatial memory impairment and recovery from were measured by the Morrison water maze test [25]. Three different age groups of rats were studied for the speed of their recovery. Young rats recovered soonest, middle age rats more slowly, and old rats not at all. My interpretation of these findings is that old rats lacked an adequate restorative or
maintenance level of NAG factors, while middle age rats had a moderate level and younger ones possessed them in healthy abundance [93].

**NAG in people.** Unlike the ample data showing a declining CCD in aged person (Table 3), comparative levels of VEGF or other angiotrophic factors have not been reported in healthy mature subjects versus elderly persons. Kalaria et al. found “enhanced VEGF immunoreactivity in clusters of reactive astrocytes” in the neocortex of AD brains compared to those of elderly controls, but the authors attributed this “compensatory mechanism to counter insufficient vascularity or reduced perfusion apparent in AD” [114]. Yang et al. found that VEGF is co-localized with beta amyloid plaques of patients with AD, but they ascribed this to “vascular dysfunction in the progression of AD” [115]. Neither of these studies defines clearly the relative general levels of NAG factors in adults versus aged subjects.

10. **THE NAG HYPOTHESIS AND COGNITIVE RESERVE**

   A troubling issue for advocates of the amyloid cascade theory has been “the imperfect correlation between cognitive status and [amyloid-β] deposits in the brain” [116]. Some persons cognitively normal at death exhibit abundant accumulation of amyloid plaques and neurofibrillary tangles, while others with dementias show negligible cerebral amyloid involvement [117-122]. The former group represents a dilemma for the amyloid cascade hypothesis, while the latter cases might be simply diagnosed as VaD or “pre-AD.” The NAG hypothesis may explain this discrepancy based on the view that individuals may vary in their “cognitive reserve” – a term used previously by several investigators and now relevant to the NAG hypothesis, as discussed next [123-125].
Individuals with a putative greater neurological/cognitive reserve than others may tolerate neurological damage from amyloid other neuropathology without displaying evidence of cognitive impairment if they have a compensatory, superior microcirculation supporting that reserve. Thus “amyloid-laden” or microinfarcted subjects who are cognitively normal may have a high CCD level, adequate to support needed neurons, while “amyloid-poor” or infarct-free persons who are cognitively deficient may have a low, inadequate CCD level. In other words, if toxic amyloid deposits or some other cerebral pathology reduce neuronal and synaptic function to a borderline level, these conditions may not yet produce recognizable mental impairment in some persons who, however, could be tipped into dementia by the decline in their CCD as they age further. Again, the import of this aside is that a reduced CCD during aging may have a deleterious effect on brain function added to any primary pathology produced in the brain – e.g., by amyloid, etc. (Table 1).

11. NEURONAL LOSS VERSUS SYNAPTIC DYSFUNCTION

According to the NAG hypothesis, administering angiogenic factors to persons with SDs might forestall further mental deterioration. Treatment might also recover faded memories and revive lost executive functions if the defective cognition is due merely to synaptic dysfunction rather than neuronal loss. In a given person, cognitive impairment may result from either neuronal dysfunction/death, synaptic dysfunction, or both. Neuronal loss is currently believed to be generally irreversible, while synaptic connections appear to wax and wane with demand, as evidenced in the brains of some time jugglers and London taxi drivers. Neuronal loss is consistently found in subjects with AD but is an unsettled issue in papers concerning cognitively impaired aged persons having no evidence of AD [76,126,127]. Many such studies have failed
to find neuronal loss in the latter [75,128]. This suggests that cognitive decline in non-AD subjects may be due mainly to synaptic dysfunction. Thus, a beneficial outcome with NAG therapy seems more probable in memory-challenged aged persons lacking signs of AD than in persons with overt AD. However, some part of the impaired cognition in AD may also be due to reversible synaptic dysfunction.

12. THERAPEUTIC OPTIONS FOR SDs

Issa et al. found elevated levels of vascular endothelial growth factor (VEGF) in the penumbra of the infarcted area of human brain tissue after ischemia stroke. They proposed the factor as a potential therapeutic agent [129]. Numerous studies in rats and mice reported the positive effect of VEGF on surgically-induced focal cerebral ischemia [130-138]. The report by Rivard et al. discussed earlier is an example in rabbits and mice of enhanced muscle vascularity following VEGF treatment [108].

Using such factors to treat aged persons with AD has also been inferred or proposed previously by others. In 2004 Ward & LaManna wrote, “It had been suggested that controlling angiogenesis may also provide novel therapeutic approaches for treatment of [various] disorders,” such as AD, etc. [102]. Carmeliet & Jain remarked, “The revascularization of ischaemic tissues would benefit millions, but therapeutic angiogenesis is an unmet medical need” [103]. Wang et al. observed that “little is known about the restorative effect of VEGF for AD in vivo,” but based on their studies in [transgenic] mice showing improved cognitive function when injected with VEGF, the authors proposed “that VEGF should be pursued as a novel therapeutic agent for treatment of AD” [138]. This therapy would seem also applicable for all SDs.
Evidence that angiotrophic factors may benefit persons with AD came from a complex surgical procedure for enhancing their cerebral circulation. In 1996, Harry S. Goldsmith began transposing human omentum onto the brain surface of persons with AD [139]. An omental pedicled flap was lifted from the transverse colon and passaged subcutaneously up the abdomen, chest, and neck to the base of the skull. After being pulled through openings in it, the dura mater, and arachnoid membrane, the flap was laid directly on the parietal-temporal area of one hemisphere -- all the while maintaining the original circulation [140]. Previous studies in dogs and monkeys had shown that this procedure increased the cerebral blood flow through the collateral circulation which developed between the omental pedicle and the surface of the brain.

Research groups with Goldsmith and with William R. Shankle reported in numerous papers that some AD subjects showed marked improvement mentally and physically [140-142]. This procedure had also been found to benefit patients who suffered with transient ischemic attacks or were poststroke [143].

Post mortem examination of the brains of a dozen or so such AD cases given such grafts confirmed that collateral circulation had been established in the area underneath the pedicle and also “in zones unrelated to omental placement, such as the occipital area and the contralateral cerebral hemisphere” [143]. The question arises of how other areas of the brain (e.g., hippocampus, frontal cortex) remote from the above, benefited from the omental pedicle, as reflected in the general cognitive improvement? Besides increasing the local blood flow, Goldsmith et al. speculated that the pedicle graft might also provide the brain elsewhere with angiogenic and neurotrophic factors produced by omental adipocytes [143].

Zhang et al. measured levels of VEGF protein in 19 tissues and organs of rats [144]. A few are listed here: omentum at 884 pg/mg, inguinal adipose tissue at 87, pituitary at 106, brain
at 8, brain stem at 1.45. They also cultured rat omental adipocytes and vascular stromal cells and found the former to be the primary source of endothelial cell mitogen. A “major portion” of this was abolished by VEGF antibody, confirming VEGF as the prominent angiogenic factor produced by the omentum. If similar high levels of VEGF occur in the human omentum, this would explain the more remote effect in the brain of omental pedicles laid on its surface and would support Goldsmith’s speculation.

13. CAUTIONS CONCERNING NAG THERAPY

Recombinant forms of many trophic factors are now available for experimental and clinical use [145]. Recent research by Chopp et al. on strokes noted earlier suggests that several factors may be required for their optimal treatment [130]. The same consideration might also apply to a waning CCD in persons with senile dementias.

The synergistic effect of VEGF and basic FGF on angiogenesis was reported in both cultured endothelial cells and a rabbit ischemic hind limb model [146-149]. The “complementary actions of VEGF and angiopoietin-1 on blood vessel growth and leakage” have been discussed in three papers [150-152]. Zang et al. described how VEGF enhances angiogenesis but promotes blood-brain barrier leakage in the ischemic brain [134]. Recall that VEGF was originally termed vascular permeability factor. In mice, angiopoietin-1 protects the vasculature against plasma leakage [151]. The reports above justify emphasizing below the reservations expressed by others regarding therapy with NAG factors.

Carmeliet has stressed that vessel formation is a “complex process, requiring a finely tuned balance between numerous stimulating and inhibitory signals” [111]. Yancopoulos et al. wrote that several members of the VEGF, angiopoietin, and ephrin families “interact in a
complementary and coordinated manner to form functional vessels [capillaries] without leaks.”

The use of a single growth factor to reconstitute a vascular bed is “now viewed as somewhat naïve and even misguided” [152]. Three other groups expressed a similar concern [136,149,150]. And very recently Thau-Zuchman et al. stressed that treating traumatic brain injuries with VEGF along “could be a double edged sword,” for it might increase vascular permeability and exacerbate cerebral edema [153]. However, none of the animal studies using VEGF have reported unusual internal bleeding, but they were short term experiments.

14. ROUTES OF ADMINISTRATION

In earlier publications, I outlined various routes for introducing NAG factors into the brain [93,94]. The following is a distillation of that review. Parenterally administered NAG factors, as well as those given orally and absorbed into the circulation, would have access to receptors on the intraluminal surface of all capillaries in the body. These two routes have the obvious drawback in that the NAG factors would not be limited to the brain but would reach other parts of the body where they might produce undesirable angiogenesis. But blood borne NAG factors could become localized in the brain if they were first modified by either of following two methods – 1) being conjugated to an antibody specific for an antigen on the intraluminal surface of capillary endothelial cells or 2) coupled to magnetic nanoparticles, introduced into the circulation (or CSF), and attracted to the cerebral capillary bed and maintained there by a magnetic field maintained over the skullcap.

Studies on Parkinson’s disease and related neurological diseases have employed three other routes for administering drugs directly into the brain: indwelling catheters in the lateral ventricle for access to the CSF, multiple microcatheters penetrating the skull and reaching the
subarachnoid space over the cerebral hemisphere, and burr holes in the calvaria allowing access to the surface of the cerebral cortex for placing slow sustained release systems [93,154-156]. Any of these three avenues could be tested experimentally on a small number of subjects with AD, but none is practical as a “standard” method for millions of aged persons. However, the transnasal route introduces agents directly into the brain and seems an eminently suitable method for the frequent, regular delivery of NAG factors.

15. TRANSNASAL THERAPY

There are numerous summaries describing this delivery method [157-160]. Agents crossing the nasal epithelium may gain access to the brain by three routes: neural, lymphatic, or vascular [161]. The olfactory nerves (beneath the olfactory mucosa) offer either of two pathways – a intracellular/transcellular pathway (within receptor-mediated neurons) and a extracellular/paracellular pathway (via channels formed by ensheathing cells along the axons). Transport of drugs along the former is slow, while that along the latter is fast with some drugs reaching the brain within 30 min. [158,159]. This is an optimal path for neurotrophic factors but would seem less so for neuroangiogenic ones.

A second route, the lymphatics, involves CSF in perineural and perivascular spaces leading into the subarachnoid space and thence into the brain parenchyma. Angiogenic factors presumably act on the intraluminal surface of capillaries. If these factors entered the brain’s interstitial spaces by either of the above two routes, they might reach the interior of capillaries by the reverse of the usual outward passage across the BBB.

A third route involves small blood vessels in the lamina propria underlying the mucosa of the nasal vestibule, olfactory region, and respiratory region. Nasal arteries convey blood via
local capillaries to nasal veins which join the facial veins, innominate veins, etc. to complete its return to the heart. But **nasal veins** also connect to the perihypophyseal vascular complex. A cavernous sinus-carotid artery complex is the counterpart in man and rats to the carotid rete mirabile in pigs, sheep, and oxen. By this short cut, venous blood from the nose may enter the arterial system of the brain in a process termed “counter current exchange” [161]. Any NAG factors present would thus reach capillaries in the brain and could possibly promote angiogenesis on their intraluminal surface.

The feasibility of transnasal therapy in people is supported by many animal studies. For example, Shipley inserted gelfoam soaked in a wheat germ agglutinin-horseradish peroxidase solution into the nose of rats and later found this marker in neurons “in the midbrain and pons throughout the entire expanse of the olfactory cortex to the caudal pole of the cerebral hemisphere” [162]. Investigators with William H. Frey II have used the transnasal route for administering various neurotrophic factors and also insulin to mice and rats [163-166]. They administered recombinant human NGF by nose drops to anesthetized rats and used an enzyme-linked immunosorbent assay to localize the factor in the brain [167]. They found concentrations of NGF within an hour reached 3,400 picomoles in the olfactory bulb (lying on the cribriform plate) and 660-2200 picomoles in four “adjacent brain regions” and several subcortical regions – hippocampus and amygdala.

In an early paper by this group, the authors suggested that the intranasal route be used for “long-term treatment of Alzheimer’s disease” with “potent drugs, including peptides and proteins” [167]. Clinical trials in AD subjects are now underway with insulin administered transnasally [168]. For introducing NAG factors, a nasal spray would seem an obvious vehicle, but a recent article on this subject included the picture of an 18th century English gentleman
taking a pinch of snuff [94]. Perhaps in the future, people may regularly use nicotine-free snuff containing the appropriate cocktail of NAG factors to ward off or ameliorate senile dementias.

16. SUMMARY & CONCLUSIONS

This essay is a synthesis of data and ideas culled from the enormous literature concerning AD, amyloid, SDs, aging, and other areas. In 17 published studies, data on cerebral capillaries show reduced values in older animals and people (Tables 2 & 3). Among the numerous articles on AD which I have read, only five examined the CCD in various parts of the brain (Table 3, #16-20), but each such paper reported reduced values compared with control groups. These observations have not been accorded any general note by other investigators of AD. Nor has consideration been given to the idea that an age-linked decline in the CCD may be important in the terminal clinical course of persons with various SDs.

The NAG hypothesis presented here proposes that senile dementia, and in some instances AD, may be caused by a reduced CCD due to a genetically determined, age-linked decline of NAG factors. Capillary function may compromised by various other mechanisms (e.g., amyloid deposits, microinfarcts, etc.), but the age-linked reduced CCD occurs in addition to them. The resulting decreased delivery of oxygen and glucose to the brain parenchyma would undermine neuronal and synaptic function in the aged and in AD. A reduced CCD could impair cognition independent of any other of the various pathologies proposed as causing AD and listed in Table 1. In elderly persons (with or without AD), a waning CCD may be a terminal vascular condition, accounting for impaired cognition. The concept of cognitive reserve is a key consideration in the NAG hypothesis.
An age-linked decline of NAG factors renders SDs a deficiency disorder, theoretically amenable to replacement therapy. The administration of recombinant neuroangiogenic factors has been shown to augment capillary density following surgically induced focal cerebral ischemia in rats and hind limb ischemia in rabbits. The NAG hypothesis offers a rational basis for considering these factors in the treatment of persons with SDs. The most practical route for administering a safe, suitable cocktail of angiongenic factors over many years may be an intranasal one, as in a nasal spray or perhaps via a snuff-like preparation.

Finally, as indicated earlier, the NAG hypothesis is not a rival to other theories concerning the etiopathology of AD, which indeed may have multiple ones. Hickham’s dictum states that “Patients can have as many diseases as they damn well please” [169]. By analogy, some disease conditions like AD may have multiple causes. Indeed, certain cases of AD might possibly benefit from dual treatment – for the diminished capillary circulation and for some other underlying cerebral pathology.

END
ACKNOWLEDGEMENTS

I am very grateful for the valuable, insightful comments made by three reviewers of early versions of this paper. I am again indebted to the Medical Center Library of the University of Kentucky and in particular to its very helpful staff member, Mrs. Amanda Williams. Finally I acknowledge the support of I.S. Tray II.

FOOTNOTES:
[1] The Indian elephant pictured in Table 1 refers to the legend of the six blind Hindus who encountered one for the first time. Each described it according to the particular part of its body he examined. The legend parallels somewhat research on AD.

REFERENCES


[145] Cell Sciences, Inc., Canton, MA. See Info@cellscience.com and other sources.


**END**
**Table 1. Approaches pursued in the study of Alzheimer’s Disease** [1, Footnote 1]

1. Nerve growth factor.
2. Neurotoxic trace elements: Al, Hg, Fe, Cu
3. Neurotransmitter deficits: cholinergic, noradrenergic, etc.
4. Oxidative stress, oxygen free radicals,
5. Inflammation
6. Mitochondrial dysfunction
7. Autophagy, protein turnover
8. Neuronal cell cycle abnormalities
9. Calcium channel blockers
10. Cortical glucose utilization and transport across the blood brain barrier
11. Autoimmunity
12. Transmissible agent: prion, subviral entities, HSV type 1
13. Amyloid hypothesis: Aβ accumulation, BFT, amyloid aggregates, tau
14. Genetics
15. Vascular risk factors
16. Microinfarcts in the brain

* De la Torre’s seven “more cited hypothetical proposals” [1].
[Footnote 1] The Indian elephant pictured in Table 1 refers to the legend of the six blind Hindus who encountered one for the first time. Each described it according to the particular part of its body he examined. The legend parallels somewhat research on AD.
**TABLE 2.** Cerebral capillary density in **Rats, Mice, & Macaque monkeys**

<table>
<thead>
<tr>
<th>Author/s</th>
<th>Young</th>
<th>Adult</th>
<th>Aged</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>RATS</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1. Craigie, 1925 [49]... TABLE 7</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- sum of capil. lengths; av. of 5 laminae</td>
<td>d 0</td>
<td>5 mo</td>
<td>13 mo</td>
</tr>
<tr>
<td>-- <em>regio insularis:</em></td>
<td>262 µ</td>
<td>856 µ</td>
<td>638 µ</td>
</tr>
<tr>
<td>-- <em>regio temporalis:</em></td>
<td>231 µ</td>
<td>958 µ</td>
<td>880 µ</td>
</tr>
<tr>
<td>2. Klein &amp; Michel, 1977 [54]... Table III</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- number of vascular elements/Chalkley count</td>
<td>6-8 mo</td>
<td>25-27 mo</td>
<td></td>
</tr>
<tr>
<td>-- frontal &amp; occipital neocortex:</td>
<td>--</td>
<td>448</td>
<td>351</td>
</tr>
<tr>
<td>3. Knox &amp; Oliveira, 1980 [55]... Table 2</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- number of microvessels/core field</td>
<td>3 mo</td>
<td>24 mo</td>
<td></td>
</tr>
<tr>
<td>-- cerebral cortex:</td>
<td>--</td>
<td>143.3 µm ± 5.8</td>
<td>131.8 µm ± 5.7</td>
</tr>
<tr>
<td>4. Wilkinson et al., 1981[45]... Fig. 3 (est’d)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- number of vessel fragments/unit zone</td>
<td>25 mo</td>
<td>31 mo</td>
<td></td>
</tr>
<tr>
<td>-- cerebral cortex</td>
<td>420 ± 10</td>
<td>365 ± 23</td>
<td></td>
</tr>
<tr>
<td>5. Knox, 1982 [56]... Table 4</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- numerical capillary density</td>
<td>3 mo</td>
<td>25 mo</td>
<td></td>
</tr>
<tr>
<td>-- cerebral cortex:</td>
<td>--</td>
<td>73.95 ± 2.47</td>
<td>68.05 ± 4.84</td>
</tr>
<tr>
<td>6. Casey &amp; Feldman, 1985 [57]... Table 1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- mean volume density ratio</td>
<td>6 mo</td>
<td>33 mo</td>
<td></td>
</tr>
<tr>
<td>-- medial nucleus of trapezoid body</td>
<td>--</td>
<td>0.0404 ± 0.0023</td>
<td>0.0283 ± 0.0015</td>
</tr>
<tr>
<td>7. Buchweitz-Milton &amp; Weiss, 1987 [58] ... Table 2 &amp; 4</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- average total capillary length</td>
<td>8-10 mo</td>
<td>21-33 mo</td>
<td></td>
</tr>
<tr>
<td>-- cortex</td>
<td>830.1 + 115</td>
<td>576.6 + 36</td>
<td></td>
</tr>
<tr>
<td>-- all 6 brain areas</td>
<td>836.6 + 54</td>
<td>592.2 + 22.2</td>
<td></td>
</tr>
<tr>
<td>8. Jucker &amp; Meier-Ruge, 1989 [59] ... Table 1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- no. of capillaries/measurement field</td>
<td>18 mo</td>
<td>27½ mo</td>
<td></td>
</tr>
<tr>
<td>-- parietal cortex, region 39</td>
<td>38.6 + 1.6</td>
<td>27.4 + 2.6</td>
<td></td>
</tr>
<tr>
<td>-- hippocampus, CA1 region</td>
<td>32.0 + 1.2</td>
<td>24.4 + 1.7</td>
<td></td>
</tr>
<tr>
<td>9. Jucker et al., 1990 [60] ... Tables 1a &amp; 1b</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- number of capillaries/field</td>
<td>18 mo</td>
<td>27 mo</td>
<td></td>
</tr>
<tr>
<td>-- hippocampus, CA1</td>
<td>32.0 ± 1.2</td>
<td>24.4 ± 1.7</td>
<td></td>
</tr>
</tbody>
</table>
-- parietal cortex, area 39 -- 38.6 ± 1.6 27.4 ± 2.6

10. Amenta et al., 1995a [53] ... Table 3
   -- sum of capillary lengths/field 12 mo (8) 27 mo (8)
   -- frontal cortex -- 190 µm ± 8 168 µm ± 6.3
   -- occipital cortex 148 µm ± 4.3 130 µm ± 4.7
   -- Ammon’s horn 170 µm ± 4.6 145 µm ± 6.1

11. Amenta et al., 1995b [61] ... Table 4
   -- capillary profile number 12 mo (10) 18 mo (10)
   -- frontal cortex 122 ± 7 71 ± 4.3
   -- occipital cortex 130 ± 6 82 ± 5
   -- hippocampus 113 ± 6 58 ± 4

MICE

12. Biron et al., 2011 [34] ... text & Table 4A
   -- % laminin fluorescent area/unit area -- 5 mo (4) 18-24 mo (5)
   -- hippocampus & frontal cortex 0.2321 ± 0.0110 0.1882 ± 0.001

MACAQUE MONKEY

13. Burns et al. 1979 [62] ... Table 1
   -- mean cross-sectional area of entire capillary in µ² 4 yr (3) 20 yr (5)
   -- frontal cortex 31.6 ± 8.1 22.8 ± 2.2
   -- occipital cortex 36.6 ± 10.4 20.7 ± 3.5

Key for Figs. 2 & 3: Average age or range of ages in years above figures.
   Number in each group: ( ).  d = day, mo = month
### TABLE 3. Cerebral capillary density in People

<table>
<thead>
<tr>
<th>Author/s</th>
<th>Adult</th>
<th>Aged</th>
<th>AD</th>
</tr>
</thead>
<tbody>
<tr>
<td>14. Mann et al., 1986 [64]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- capillary length sum/unit volume, mm/mm(^3)</td>
<td>26-58 yr (6)</td>
<td>76-96 yr (9)</td>
<td></td>
</tr>
<tr>
<td>-- frontal cortex:</td>
<td>965.5</td>
<td>783.6</td>
<td>--</td>
</tr>
<tr>
<td>15. Abernathy et al., 1993 [65]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- capillary length sum/unit vol., mm/mm(^3)</td>
<td>30-50 yr (4)</td>
<td>60-85 yr (10)</td>
<td></td>
</tr>
<tr>
<td>-- paraventricular nuclei:</td>
<td>1560</td>
<td>1100</td>
<td>--</td>
</tr>
<tr>
<td>-- range</td>
<td>2300 -1100</td>
<td>1800 - 800</td>
<td></td>
</tr>
<tr>
<td>16. Bell &amp; Ball, 1981 [66]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- mean capillary density, mm/mm(^3)</td>
<td>av. 38 yr (5)</td>
<td>av. 74 yr (5)</td>
<td></td>
</tr>
<tr>
<td>-- entorhinal</td>
<td>148</td>
<td>124</td>
<td>111</td>
</tr>
<tr>
<td>-- overall of 6 zones</td>
<td>124</td>
<td>102</td>
<td>101</td>
</tr>
<tr>
<td>17. Bell &amp; Ball, 1990 [67]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- av. capillary density, mm/mm(^3)</td>
<td>12-54 yr (8)</td>
<td>67-95 (8)</td>
<td></td>
</tr>
<tr>
<td>-- visual cortex, lamina 1</td>
<td>141</td>
<td>99</td>
<td>97</td>
</tr>
<tr>
<td>-- visual cortex, av. of 6 laminae</td>
<td>251</td>
<td>212</td>
<td>206</td>
</tr>
<tr>
<td>18. Buée et al., 1994 [68]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- % capil. surface area of total cortical field area</td>
<td>49 yr (1)</td>
<td>av. 79 yr (3)</td>
<td></td>
</tr>
<tr>
<td>-- cortical areas:</td>
<td>26.32%</td>
<td>18.95%</td>
<td>16.50%</td>
</tr>
<tr>
<td>19. Fisher et al., 1990 [47]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- vascular density index</td>
<td>control: 23-90 yr (6)</td>
<td>AD: 76-92 yr (16)</td>
<td></td>
</tr>
<tr>
<td>-- prefrontal cotex:</td>
<td>94.6</td>
<td></td>
<td>75.4</td>
</tr>
<tr>
<td>-- basal forebrain:</td>
<td>86.8</td>
<td></td>
<td>42.7</td>
</tr>
<tr>
<td>-- hippocampus:</td>
<td>82.3</td>
<td></td>
<td>50.2</td>
</tr>
<tr>
<td>-- motor sensory cortex:</td>
<td>94.3</td>
<td></td>
<td>83.1</td>
</tr>
<tr>
<td>20. Kitaguchi et al., 2007 [69]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-- capillary density in test grid method</td>
<td>control: 73 ± 4 (6)</td>
<td>AD: 79 ± 12 (8)</td>
<td></td>
</tr>
<tr>
<td>-- frontal cortex</td>
<td>28</td>
<td>21</td>
<td></td>
</tr>
<tr>
<td>-- parietal cortex</td>
<td>28</td>
<td>20</td>
<td></td>
</tr>
</tbody>
</table>