

University of Kentucky

UKnowledge

Sanders-Brown Center on Aging Faculty
Publications

Aging

10-23-2008

Focal cerebral ischemia in the TNFalpha-transgenic rat

L. Creed Pettigrew

University of Kentucky, cpetti@uky.edu

Mark S. Kindy

Medical University of South Carolina

Stephen W. Scheff

University of Kentucky, sscheff@email.uky.edu

Joe E. Springer

University of Kentucky, joe.springer@uky.edu

Richard J. Kryscio

University of Kentucky, kryscio@email.uky.edu

See next page for additional authors

Follow this and additional works at: https://uknowledge.uky.edu/sbcoa_facpub



Part of the [Family, Life Course, and Society Commons](#), and the [Geriatrics Commons](#)

Right click to open a feedback form in a new tab to let us know how this document benefits you.

Repository Citation

Pettigrew, L. Creed; Kindy, Mark S.; Scheff, Stephen W.; Springer, Joe E.; Kryscio, Richard J.; Li, Yizhao; and Grass, David S., "Focal cerebral ischemia in the TNFalpha-transgenic rat" (2008). *Sanders-Brown Center on Aging Faculty Publications*. 25.

https://uknowledge.uky.edu/sbcoa_facpub/25

This Article is brought to you for free and open access by the Aging at UKnowledge. It has been accepted for inclusion in Sanders-Brown Center on Aging Faculty Publications by an authorized administrator of UKnowledge. For more information, please contact UKnowledge@lsv.uky.edu.

Focal cerebral ischemia in the TNFalpha-transgenic rat

Digital Object Identifier (DOI)

<http://dx.doi.org/10.1186/1742-2094-5-47>

Notes/Citation Information

Published in *Journal of Neuroinflammation*, v. 5, 47.

© 2008 Pettigrew et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Authors

L. Creed Pettigrew, Mark S. Kindy, Stephen W. Scheff, Joe E. Springer, Richard J. Kryscio, Yizhao Li, and David S. Grass

Research

Open Access

Focal cerebral ischemia in the TNF α -transgenic rat

L Creed Pettigrew^{*1,2,3}, Mark S Kindy^{4,5}, Stephen Scheff¹, Joe E Springer⁶,
Richard J Kryscio⁷, Yizhao Li⁸ and David S Grass⁹

Address: ¹Sanders-Brown Center on Aging, University of Kentucky, Lexington, Kentucky, USA, ²Department of Neurology, University of Kentucky, Lexington, Kentucky, USA, ³Veterans Administration (VA) Medical Center, Lexington, Kentucky, USA, ⁴Department of Neuroscience, Medical University of South Carolina, Charleston, South Carolina, USA, ⁵Ralph H. Johnson VA Medical Center, Charleston, South Carolina, USA, ⁶Department of Physical Medicine & Rehabilitation, University of Kentucky, Lexington, Kentucky, USA, ⁷Department of Statistics and School of Public Health, University of Kentucky, Lexington, Kentucky, USA, ⁸Jinan Great Wall Hospital, Jinan, Shandong, PR China and ⁹Xenogen Biosciences, Cranbury, New Jersey, USA

Email: L Creed Pettigrew* - cpetti@uky.edu; Mark S Kindy - kindyms@musc.edu; Stephen Scheff - sscheff@email.uky.edu; Joe E Springer - joe.springer@uky.edu; Richard J Kryscio - kryscio@email.uky.edu; Yizhao Li - swillia@uky.edu; David S Grass - David.Grass@xenogen.com

* Corresponding author

Published: 23 October 2008

Received: 16 January 2008

Journal of Neuroinflammation 2008, **5**:47 doi:10.1186/1742-2094-5-47

Accepted: 23 October 2008

This article is available from: <http://www.jneuroinflammation.com/content/5/1/47>

© 2008 Pettigrew et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: To determine if chronic elevation of the inflammatory cytokine, tumor necrosis factor- α (TNF α), will affect infarct volume or cortical perfusion after focal cerebral ischemia.

Methods: Transgenic (TNF α -Tg) rats overexpressing the murine TNF α gene in brain were prepared by injection of mouse DNA into rat oocytes. Brain levels of TNF α mRNA and protein were measured and compared between TNF α -Tg and non-transgenic (non-Tg) littermates. Mean infarct volume was calculated 24 hours or 7 days after one hour of reversible middle cerebral artery occlusion (MCAO). Cortical perfusion was monitored by laser-Doppler flowmetry (LDF) during MCAO. Cortical vascular density was quantified by stereology. Post-ischemic cell death was assessed by immunohistochemistry and regional measurement of caspase-3 activity or DNA fragmentation. Unpaired *t* tests or analysis of variance with post hoc tests were used for comparison of group means.

Results: In TNF α -Tg rat brain, the aggregate mouse and rat TNF α mRNA level was fourfold higher than in non-Tg littermates and the corresponding TNF α protein level was increased fivefold ($p \leq 0.01$). Infarct volume was greater in TNF α -Tg rats than in non-Tg controls at 24 hours ($p \leq 0.05$) and 7 days ($p \leq 0.01$). Within the first 10 minutes of MCAO, cortical perfusion measured by LDF was reduced in TNF α -Tg rats ($p \leq 0.05$). However, regional vascular density was equivalent between TNF α -Tg and non-Tg animals ($p = \text{NS}$). Neural cellular apoptosis was increased in transgenic animals as shown by elevated caspase-3 activity ($p \leq 0.05$) and DNA fragmentation ($p \leq 0.001$) at 24 hours.

Conclusion: Chronic elevation of TNF α protein in brain increases susceptibility to ischemic injury but has no effect on vascular density. TNF α -Tg animals are more susceptible to apoptotic cell death after MCAO than are non-Tg animals. We conclude that the TNF α -Tg rat is a valuable new tool for the study of cytokine-mediated ischemic brain injury.

Background

Tumor necrosis factor- α (TNF α) is a pleiotropic cytokine suspected to enhance or deter cellular survival through activation of receptor-mediated signal transduction. When present in supra-physiological levels after injury, it is known to modulate neural cell loss in cerebral ischemia [1], intracerebral hemorrhage [2], chronic cerebral oligemia [3], and trauma [4]. The level of TNF α in human brain becomes elevated after cerebral infarction [5] and appears sequentially in the infarct core and peri-infarct areas before expression in tissue within the unaffected hemisphere [6]. Elevated levels of TNF α have been observed consistently in serum [7-9] and in cerebrospinal fluid [8,9] after acute ischemic stroke. In animal models of cerebral ischemia, high levels of TNF α have been found after global [10,11] and focal [12] ischemic injury. Several investigators reported reduction of infarct volume through inhibition of TNF α [13-15], although Nawashiro and colleagues [16] showed that pretreatment of mice by intracisternal administration of TNF α reduced infarct volume paradoxically without an inhibitor.

Active, soluble TNF α is derived through proteolytic cleavage from a transmembrane precursor by TNF convertase (TACE), a member of the disintegrin and metalloproteinase (ADAM) group of enzymes. Among neural cell types, soluble TNF is secreted robustly by microglia and astrocytes [17,18] but is also expressed by neurons and steroid-producing cells of the adrenal *zona reticularis*. Once released, soluble TNF circulates as a homotrimer that will bind with one of two distinct receptors of 55-kDa (TNFR1) or 75-kDa (TNFR2) molecular weight. The complex interactions of both soluble and transmembrane TNF α with the two TNF receptors have been reviewed [19]. The first receptor, p55/TNFR1, may bind either form of TNF α before dissociating from an endogenous inhibitor and recruiting the TNF receptor-associated protein death domain (TRADD). Once complexed, p55/TNFR1-TRADD may either facilitate apoptosis by activation of caspase-8 or enhance cell survival through disinhibition of the nuclear transcription factor, NF κ B. The second receptor, p75/TNFR2, becomes active only upon binding transmembrane TNF α and will develop a low-affinity interaction with TRADD. The p75/TNFR2-TRADD construct is less stable than p55/TNFR1-TRADD and is considered to have a more focused role in cell survival by upregulating anti-apoptotic factors or preventing activation of caspase-8. Certain steps in the cascade, such as the activation of caspase-8 to induce apoptosis and the release of NF κ B to generate pro-survival elements, arise upon stimulation of the same receptor (p55/TNFR1) and may constitute a natural check-and-balance system that determines the ultimate fate of the cell.

Our previous work suggests that the apparent duality of effect attributed to TNF α is controlled by interaction between the cytokine ligand and p55/TNFR1 or p75/TNFR2. We demonstrated that inactivation of both receptors in double-knockout mice causes exacerbation of the excitotoxic effects of kainic acid in the CA₃ region of the hippocampus and expansion of infarct volume after middle cerebral artery occlusion (MCAO) [20]. We further highlighted the role of p55/TNFR1 as a gatekeeper molecule by showing that knock-out mice lacking only this receptor had significantly greater neuronal injury after kainic acid injection or MCAO [21].

In this paper, we describe the first application of focal cerebral ischemia in a unique transgenic rat overexpressing the murine TNF α gene. We present the results of a feasibility experiment testing the hypothesis that chronic elevation of TNF α in brain increases infarct volume after focal cerebral ischemia. We also report on comparison of TNF α -transgenic (TNF α -Tg) rats with non-transgenic (non-Tg) controls to determine if overexpression of the TNF α gene alters the vascular anatomy of the cerebral cortex, the circulatory response of the brain to ischemia, or post-ischemic cellular apoptosis. Our overall goal in performing this work was to determine how elevated levels of TNF α protein, such as may be observed in human brain after stroke, will modify cerebral ischemic injury.

Methods

Construction of the transgenic animal

The TNF α -Tg rat was constructed in the research facility of Xenogen Biosciences in Cranbury, New Jersey. Construction began by isolation of the murine TNF α gene from a 129 SV mouse genomic library as described previously [22]. The 2.8 kb EcoRI fragment containing the murine TNF α promoter and the entire coding region of the corresponding murine TNF α gene was ligated to the 0.77 kb EcoRI-PstI/SalI fragment containing the 3'-untranslated region and polyadenylation signal of the human β -globin gene [23]. The BamHI-SalI fragment was microinjected into fertilized eggs of Sprague-Dawley (S-D) rats using standard techniques [24]. TNF α -Tg males were bred to female, wild type S-D rats. Approximately half of the resulting offspring carried the murine TNF α gene, as confirmed by polymerase chain reaction (PCR) genotyping with primers specific for the murine TNF α gene, 5'-GGG GAA AGG TGG TAT CTC-3' (sense primer) and 5'-CGC TTT CTT GCT GTC CAA-3' (antisense primer) applied to total cellular DNA isolated from tail snip tissue.

Measurement of TNF α mRNA and protein

The Institutional Animal Care and Use Committee of the University of Kentucky approved all experiments performed with the TNF α -Tg rat. To analyze TNF α mRNA levels in TNF α -Tg and non-Tg brain, reverse transcriptase-

PCR (RT-PCR; Ambion, Austin, TX) was performed on total RNA isolated from one cerebral hemisphere. An enzyme-linked immunosorbent assay (ELISA) was used to detect aggregate mouse and rat TNF α in the remaining cerebral hemisphere (R & D Systems, Minneapolis, MN).

To determine the contribution of the native rat gene to the mRNA response, we screened for rat TNF α mRNA in different brain regions, external organs, and serum in a second set of animals. Samples were homogenized and total RNA was extracted with an RNeasy Mini Kit (QIAGEN Inc., Valencia, CA). The Quantikine rat TNF α probes and Calibrator Kit (#RN510) was then used with an mRNA base kit (#RN000) of R & D Systems to quantify rat TNF α mRNA. Parallel samples were homogenized in phosphate-buffered saline (PBS), centrifuged, and total rat and mouse TNF α was measured by ELISA (R & D Systems; Quantikine M catalog #MTA00).

To measure TNF α protein and mouse or rat mRNA in small quantities of brain tissue (= 50 mg) obtained from ischemic animals, we developed a micro-sample preparation technique and refined our protein ELISA and mRNA quantification methods. Tissue samples obtained from the border zone approximating the ischemic penumbra and the infarct core in the rat brain were homogenized in Mammalian Tissue Lysis Buffer (#C3228; Sigma Chemical Company, St. Louis, MO) consisting of a dialyzable mild detergent (bicine), 150 mM NaCl, and a cocktail of protease inhibitors (Roche). The homogenates were centrifuged at 12,000 \times g for 10 minutes at 4°C. Protein concentration was quantified with the BCA protein assay kit (#23225, Pierce Chemicals, Rockford, IL).

For measurement of TNF α protein in small tissue samples, we used an ELISA kit (eBioscience, San Diego, CA) containing a monoclonal antibody that can detect both mouse and rat TNF α protein in the range of 10 pg/ml – 10 ng/ml. To measure rat or mouse mRNA in small tissue samples, we extracted total RNA with the RNeasy Mini Kit and used the same Quantikine mRNA base kit described above. Quantikine rat TNF α mRNA probes and Calibrator Kit (#RN510) and Quantikine mouse TNF α mRNA probes and Calibrator Kit (#RN410) with Quantikine mRNA base kit (#RN000) of R & D Systems were used to quantify mouse and rat TNF α mRNA. After completion of this assay, mouse or rat TNF α mRNA was quantified in attomoles/ml/1 μ g total RNA and expressed as a ratio (mouse/rat TNF α mRNA).

Western blot analysis for TRADD protein

We measured expression of TRADD protein, derived upon interaction of active, soluble TNF α with p55/TNFR1, to demonstrate that TNF α protein retained biological activity in the ischemic brain of the TNF α -Tg rat. Micro-sam-

ples of brain tissue were homogenized and prepared as described for measurement of TNF α protein. The expression of TRADD protein was determined by western blot analysis using a monoclonal anti-TRADD antibody (BD Biosciences, USA). Aliquots of brain homogenate (30 μ g of protein) were loaded onto 10% SDS-PAGE gels for electrotransfer onto nitrocellulose membranes. After transfer, the membranes were blocked by incubation in 5% BSA in TRIS-buffered saline with 0.01% Tween-20 for one hour at room temperature and were washed three times (10 minutes each) with wash buffer (1 \times TBS, 0.05% Tween-20). The membranes were then incubated with primary antibody (anti-TRADD monoclonal antibody; #610572, BD Biosciences Pharmingen, San Diego, CA) at a dilution of 1:250 for one hour at room temperature. The nitrocellulose membrane was then washed and incubated with horseradish peroxidase-conjugated goat anti-mouse IgG (1:1,000; #34080, Cell Signaling Technology, Danvers, MA) for another hour at room temperature. After three additional washes in TBS/Tween-20, immunoreactive bands were developed using an ECL chemiluminescence substrate (#34080, Pierce). A densitometry image analysis system (Kodak 2000RT) was used for quantification of band intensities on X-ray film. To screen for gel loading errors, the western blots were stripped and reprobated with an antibody to β -actin, a constitutively expressed protein.

Immunohistochemical studies

For immunohistochemical studies confirming cell identity and intracellular expression of TNF α , we prepared 30- μ m coronal brain slices and applied methods described in our previous publications [25-27]. We used a rabbit polyclonal antibody to identify both rat and mouse TNF α (AMC3012, Biosource International, Camarillo, CA; 1:2000 dilution). We chose NeuN (MAB377, Chemicon International, Temecula, CA; mouse monoclonal; 1:1000 dilution) to immunolabel neuronal nuclear protein, anti-GFAP (anti-glial fibrillary acidic protein; MAB3402, Chemicon; mouse monoclonal; 1:1000 dilution) to identify astroglia, CD11b/OX-42 (MAB1405, Chemicon; mouse monoclonal; 1:1000 dilution) to label microglia and macrophages, and anti-APC/Ab-7 (anti-oligodendroglial cell body; OP80, Oncogene Research Products; mouse monoclonal; 1:1000 dilution) to identify oligodendrocytes. To identify what type of neural cell was expressing TNF α in the transgenic animal, co-localization studies were performed by incubating coronal brain sections with the anti-TNF α antibody and NeuN, anti-GFAP, CD11b/OX-42, or anti-APC/Ab-7 labeled with fluorophores.

Suture-occlusion model of focal cerebral ischemia

Because the murine TNF α gene was expressed within S-D rats, we chose the Zea Longa technique [28] for suture-occlusion of the MCA already validated in this animal

strain. Male transgenic and non-Tg rats of 250–350 gm body weight were subjected to one hour of suture-occlusion by our modification [29] of the original Zea Longa method. Each animal was fasted overnight in preparation for surgery and then anesthetized by IP injection of chloral hydrate (350 mg/kg) and xylazine (4 mg/kg). Rectal and temporalis muscle temperatures were maintained at 36.5–37.5 °C by external warming. A suture-occluder prepared by the method of Belayev and others [30] was advanced retrograde through the external carotid artery and into the internal carotid artery to occlude the MCA. After one hour, the suture-occluder was withdrawn. In each rat, the left femoral artery was reversibly cannulated for assessment of mean arterial blood pressure (MABP) and blood gases before, during, and after suture-occlusion.

Measurement of infarct volume

Infarct volume was measured 24 hours or 7 days after removal of the suture-occluder from the MCA in ischemic animals. Rats prepared for measurement of infarct volume were re-anesthetized by IP injection of the same combined doses of chloral hydrate and xylazine used as anesthesia for surgery to achieve focal cerebral ischemia. For brain sampled after 24 hours of post-ischemic reperfusion, each anesthetized animal was euthanized for staining of coronal brain sections with triphenyltetrazolium chloride (TTC) [31]. Total percent infarct volume was quantified using NIH Image v. 1.62 on a computer-assisted imaging system described in our previous work [26,29]. Infarct volume was corrected for ischemic edema by the method of Swanson and co-authors [32].

For measurement of infarct volume after 7 days of post-ischemic reperfusion, animals were re-anesthetized by IP injection of chloral hydrate/xylazine and perfused transcardially with 30 ml of cold, heparinized 0.1 M PBS (pH 7.4), followed by 60 ml of 4% paraformaldehyde in PBS. Each brain was removed *en bloc* and placed in 4% paraformaldehyde/0.1 M PBS at 4 °C for 24 hours. The fixed brain specimen was then washed externally with PBS × 3 (10 minutes each) at 4 °C. The brain specimen was blocked and immersed in 20% sucrose/0.1 M PBS at 4 °C for 24 hours. The fixed, sucrose-protected brain was mounted on a metal chuck with cryostat embedding medium. The mounted brain was frozen in powdered dry ice and brought to ambient temperature within the cryostat (-21 °C) before 80-µm coronal sections were cut to encompass the anticipated infarct volume. Every fifth section was mounted directly onto glass microscopic slides that were dried on a slide warmer at 40 °C. The dried sections were defatted in xylene overnight before rehydration through gradient alcohol solutions to water. The resulting sections were then stained in 5% cresyl violet acetate for 2 minutes, rinsed in water, dehydrated through gradient alcohol

solutions to xylene, and coverslipped. Total percent infarct volume was quantified using NIH Image v. 1.62 on the same computer-assisted imaging system used for quantifying 24-hour infarct volume, but without correction for tissue edema.

Assessment of neural cell death after focal cerebral ischemia

We employed both qualitative and quantitative methods to evaluate apoptotic cell death in the brains of TNF α -transgenic and non-Tg rats after ischemic injury. For qualitative determination of what cell type was undergoing caspase-3 activation after ischemia, we adapted the method of McEwen and Springer [33] from its original application in experimental spinal cord trauma to our model of suture-occlusion of the MCA. Twenty-four hours after reversal of ischemia by withdrawal of the suture-occluder, each rat was re-anesthetized by IP injection of chloral hydrate/xylazine and perfused transcardially with 20 ml of cold, heparinized 0.1 M PBS (pH 7.4), followed by 40 ml of 4% paraformaldehyde in PBS. The brain was removed *en bloc* and placed in 4% paraformaldehyde in PBS at 4 °C for one hour. The fixed brain specimen was then washed externally with PBS × 3 (10 minutes each) at 4 °C. The brain specimen was then blocked and immersed in 20% sucrose-PBS at 4 °C for 48 hours, before being frozen rapidly on dry ice and embedded in tissue freezing medium (Triangle Biomedical Sciences, Durham, NC). The frozen brain was then cut into 20-µm coronal sections on a cryostat (Microm Laborgerate, Walldorf, Germany). Consecutive sections taken from both TNF α -Tg and non-Tg rats were mounted in pairs on a series of charged glass slides (Superfrost Plus; Fisher Scientific, Pittsburgh, PA).

The sections were air-dried and stored at -20 °C before preparation for fluorescence immunohistochemistry, all procedures for which were performed at room temperature except as noted. All sections were air-dried for 30 minutes after removal from cold storage, washed repeatedly in PBS for one hour, and pre-incubated for 30 minutes in a blocking solution comprised of 5% normal goat serum (NGS) and 0.05% Triton X-100 in PBS. The blocking solution was removed and sections were incubated in a humidified chamber overnight at 4 °C in PBS that contained 5% NGS and one of the following sets of primary antibodies: (a) rabbit anti-human/mouse activated caspase-3 IgG (polyclonal, 1:1,000; R & D Systems, Minneapolis, MN), (b) mouse anti-neuronal nuclei (NeuN) IgG (monoclonal, 1:600; Chemicon International, Temecula, CA) to identify neurons, or (c) anti-glial fibrillary acidic protein (GFAP; monoclonal, 1:600; Sigma, St. Louis, MO) to localize astrocytes. To test for nonspecific staining by the secondary antibodies, additional slides were processed in a similar fashion with the primary antibodies excluded. All slides were then washed repeatedly in PBS

for one hour before being incubated in the dark for a second hour in PBS that contained 5% NGS and the following fluorescent secondary antibodies: Cy-3-conjugated goat anti-rabbit IgG (polyclonal, 1:1,600; Jackson ImmunoResearch Laboratories, West Grove, PA) and AlexaFluor 488 goat anti-mouse IgG (monoclonal, 1:600; Molecular Probes, Eugene, OR). The secondary antibody solution was removed and the slides were washed repeatedly in PBS for 10 minutes. Tissue sections were immediately coverslipped with ProLong Anti-Fade (Molecular Probes) and stored in the dark at 4°C to retard fading of the fluorescent labels. A Zeiss AxioPlan microscope (Zeiss; Oberkochen, Germany) was used to review the tissue sections after processing. Co-localization of immunoreactivity was performed with Photoshop (Adobe Systems; San Jose, CA) to superimpose paired images (40×) showing the fluorescent secondary antibodies identifying anti-active caspase-3 and the cell type of interest.

To determine if neural cell death was preceded by caspase-3 activation, we quantified the activity of this pro-apoptotic enzyme in ischemic brain. TNF α -Tg and non-Tg rats underwent MCAO by suture-occlusion for one hour. After 3 or 24 hours of post-ischemic reperfusion, each animal was re-anesthetized by IP injection of chloral hydrate and was perfused transcardially with 30 ml of 15% India ink (v/v) diluted in normal saline, as per our published method [27]. The brain was removed *en bloc*, chilled, and cut into 2-mm sections. Normally perfused brain was darkened by the ink, whereas unperfused tissue remained pale. The infarct core within each slice was dissected and snap-frozen, as was a thin rim of partially stained tissue at the interface between normal and unperfused cortex that approximated the ischemic penumbra. The resulting tissue samples were prepared by the micro-sample method described above and caspase-3 activity was measured with a caspase-3 substrate AC-DEVD-AFC Fluorometric Assay Kit (R & D Systems). All procedures were carried out according to the manufacturer's instructions. Equal amounts of protein samples were mixed with caspase-3 reaction buffer containing the fluorescent peptide AC-DEVD-AFC, pipetted in duplicate in a 96-well microtiter plate, and incubated at 37°C for 180 minutes. Caspase-3 mediated cleavage of AC-DECD-AFC into free AFC was measured using a fluorescent microplate reader (LS50B, Perkin Elmer) at a wavelength excitation of 400 nm and emission of 505 nm. Readings were obtained serially at 0, 30, 60, 120, and 180 minutes. The change in fluorescence (Δ fluorescence units) was calculated by subtracting the minimal ($t = 0$ minutes) from the maximal ($t = 180$ minutes) reading and the caspase-3 activity in each sample was calculated in Δ fluorescence units/minute/100 mg protein.

To confirm that intracellular caspase-3 activation preceded apoptotic cell death, we performed a quantitative assay for DNA fragmentation after focal cerebral ischemia. For this technique, we adapted the method of Shimizu and colleagues for measurement of DNA fragmentation in ischemic gerbil hippocampus [34]. TNF α -Tg and non-Tg rats underwent MCAO by suture-occlusion for one hour. After 3 or 24 hours of post-ischemic reperfusion, each animal was re-anesthetized by IP injection of chloral hydrate and was perfused transcardially with India ink. The brain was removed *en bloc*, chilled, cut into 2-mm sections, and sampled as described in the preceding paragraph. Frozen tissue samples were homogenized with a glass tissue grinder in isolation buffer comprised of 215 mM mannitol, 75 mM sucrose, 0.1% BSA, 1 mM EGTA (pH 7.2), and 20 mM HEPES (pH:7.2). Each homogenate was then centrifuged at 3,000 \times g for 5 min at 4°C to remove whole DNA within intact cellular nuclei. The resulting supernatant was recentrifuged at 13,000 \times g for 10 min at 4°C to discard a crude mitochondrial pellet. The second supernatant enriched for cytosolic, fragmented DNA was aliquoted and stored at -80°C. Protein concentration in the second supernatant was measured with the BCA protein assay kit (Pierce).

DNA fragmentation in the cytoplasmic supernatant was quantitatively assayed by antibody-mediated capture and detection of histone-associated DNA fragments (mono- or oligonucleosomes; Cell Death Detection ELISA Plus kit; Roche Molecular Biochemicals, Mannheim, Germany). Briefly, a 20- μ l aliquot of the cytoplasmic supernatant was used in the ELISA following the manufacturer's standard protocol. Optical density values of absorbance at 405 and 490 nm (reference wavelength) were determined with a microplate reader (SpectraMAX 340PC, Molecular Device) after incubation with a peroxidase substrate. Background values (incubation buffer alone) were subtracted from each sample. All samples were run in duplicate. The direct reading units of OD values were converted to mU (absorbance = 1000 mU). The results were shown as mU (arbitrary unit) per mg protein.

Measurement of regional cerebral perfusion and quantification of vascular density

We used laser-Doppler flowmetry (LDF) to measure regional cerebral perfusion in TNF α -Tg rats or non-Tg controls during focal cerebral ischemia and post-ischemic reperfusion. Following our previously reported method [29], each animal underwent craniotomy for placement of a 2-mm laser-Doppler probe over intact dura approximately 3 – 5 mm posterior to bregma and 4 mm lateral to midline. The probe was positioned over a cortical region affected by MCAO, from 4.2 mm anterior to 5.8 mm posterior in relation to bregma, avoiding large pial vessels to obtain reproducible, stable readings [35]. Baseline meas-

urements were taken every five minutes during 10 minutes of pre-ischemic baseline, 60 minutes of MCAO, and the first 30 minutes of post-ischemic reperfusion. Results were expressed as percent of pre-ischemic baseline.

For quantification of vascular density, we employed a stereological method used previously to study vascular changes in rodent hippocampal dentate gyrus [36]. Male TNF α -Tg and non-Tg rats (n = 3 per group) that were not subjected to MCAO were anesthetized by IP injection of sodium pentobarbital (55 mg/kg) and underwent thoracotomy to gain access to the heart. After cross-clamping of the descending aorta, each animal was infused with 20 ml undiluted Higgins black waterproof India ink #4115 (Sanford, Bellwood, IL). The infusate was delivered through a 20-gauge needle inserted directly into the beating left ventricle, using hand-applied pressure. The infused animal was decapitated; the brain was then removed *en bloc* and post-fixed in 4% paraformaldehyde for 48 hours. It was then cryoprotected in 20% sucrose (w/v) for three days under constant, mild agitation. Coronal sections of 50- μ m thickness were cut with a freezing microtome throughout the rostral-caudal extent of a specified region of the cortex extending from the septal area (inter-aural level 10.7) to the occipital region (inter-aural level -0.3) [37]. The Bioquant Image Analysis System (Nashville, TN) was used to analyze 12 equidistant sections taken from every animal. For each section, an unbiased method [38] was employed to select a region of interest (ROI; 5.26 mm²) within the cortex. With the Bioquant device, a thresholding routine was used to estimate the percent area of the ROI occupied by ink-labeled blood vessels. The vascular percent areas within the ROIs in the 12 coronal sections were averaged to derive a mean value for each animal.

Statistical analysis

All physiological measurements, levels of TNF α mRNA and protein in brain, and infarct volumes by experimental group were expressed as means \pm SD. Unpaired *t* tests were used for comparisons of group means for infarct volume, cortical vascular density, and selected physiological measurements (pre-ischemic whole blood glucose and hematocrit). All other physiological data, including measurements of cortical perfusion by LDF, were obtained serially in each rat and were compared by two-way (experimental group and sampling time) analysis of variance (ANOVA) with repeated measures. For comparison of TNF α mRNA or protein in different brain regions, external organs, and serum in TNF α -Tg and non-Tg animals, ANOVA was used with Fisher's protected least significant difference post hoc test that corrects for multiple comparisons. TRADD protein levels and mouse/rat TNF α mRNA ratios were also compared with ANOVA/Fisher's test. Statistical significance was considered at $p \leq 0.05$.

Results

Production and characterization of the TNF α -Tg rat

The murine TNF α promoter and the entire coding region of the corresponding murine TNF α gene (Figure 1, Panel a) were introduced into the TNF α -Tg rat. Weanling pups underwent tail snip on post-natal Day 21 to obtain tissue for isolation of total cellular DNA. The 501-kb fragment of the murine TNF α gene was present only in the transgenic animal, allowing confirmation of genotype by PCR (Figure 1, Panel b).

Cells that were immunoreactive for TNF α were found only in the brain of the TNF α -Tg rat. By histological examination, the neural anatomy of the TNF α -Tg rat appeared similar to that of non-Tg littermates (Figure 2, Panels a - j) when not subjected to ischemic injury. Co-localization

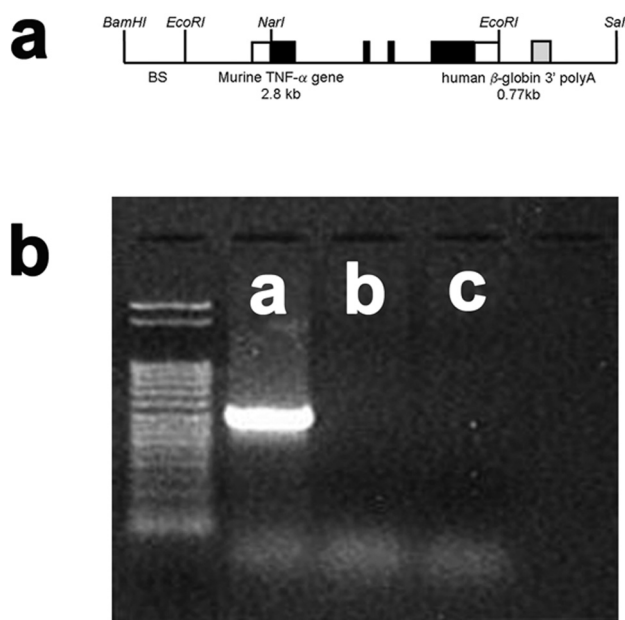


Figure 1
Structure of the murine TNF α transgenic construct and confirmation of genotype. The structure of the murine TNF α transgenic construct is illustrated in Panel a. An example of polymerase chain reaction (PCR) products stained with ethidium bromide to confirm the genotype of the TNF α -transgenic (TNF α -Tg) rat is shown in Panel b. Clear boxes in the drawing of the transgenic construct represent untranslated sequences from the mouse TNF α gene, black boxes indicate the coding region of the TNF α gene, and the gray box (including the remainder of the EcoRI to Sall fragment) represent the human β -globin 3' fragment. BS indicates pBluescript vector. Panel b shows PCR products in tail snip tissue taken from a TNF α -Tg rat (Lane a), a non-transgenic littermate (Lane b), and a Sprague-Dawley rat (Lane c). A 501 bp fragment of the murine TNF α gene was detected only in the transgenic rat.

studies (Figure 3, Panels a – c) showed overlap of signal between mouse TNF α protein and a neuronal marker, NeuN. This result identifying the rat neuron as the primary neural cell expressing TNF α protein was confirmed in three TNF α -Tg rats from three different litters. Every fifth coronal section was sampled in each of the three transgenic animal brains.

Murine TNF α mRNA and protein were overexpressed abundantly in the TNF α -Tg rat. In Figure 4, multiple panels show the results of assays for TNF α mRNA and protein in transgenic animals and non-Tg littermates that were not subjected to ischemic injury. The aggregate level of rat

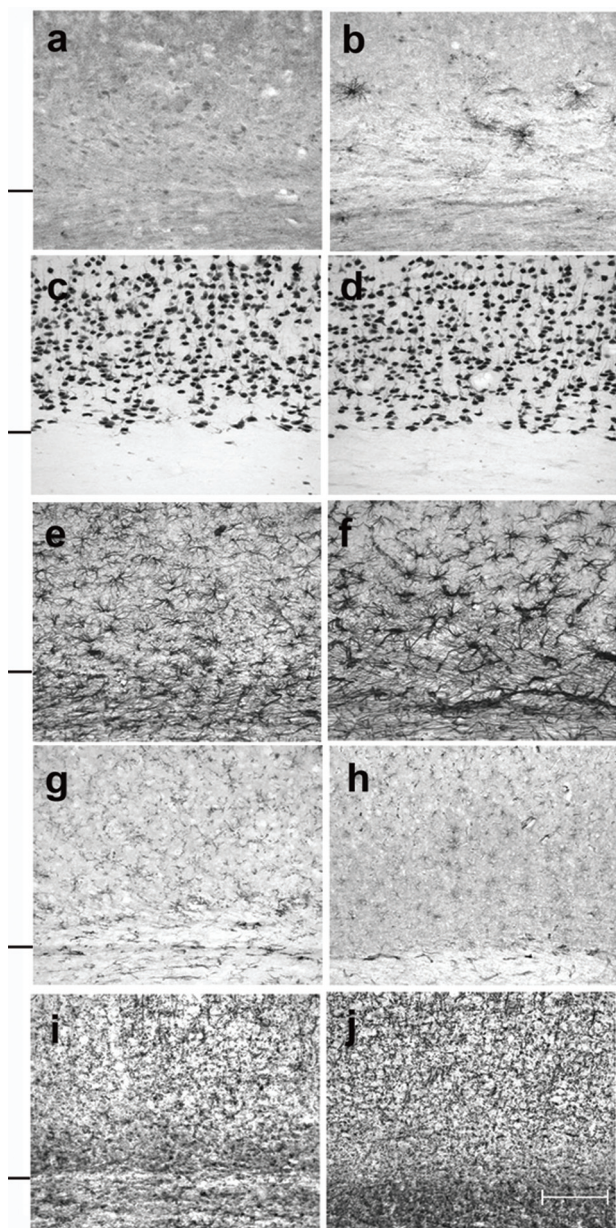


Figure 2

Figure 2

Cellular expression of TNF α protein in transgenic and non-transgenic rat brain.

This photographic collage shows cellular expression of TNF α protein and compares the brain anatomy of transgenic and non-transgenic (non-Tg) rats that were not subjected to ischemic injury. Coronal brain sections were incubated with primary antibodies identifying mouse or rat TNF α protein (Panels a and b), neurons (Panels c and d), astroglia (Panels e and f), microglia/macrophages (Panels g and h), or oligodendroglia (Panels i and j). All light microscopic images were taken at 20 \times magnification. Transgenic rat brain is shown in Panels b, d, f, h, and j. All remaining images are of brain in non-Tg littermates. Cells that were immunoreactive for TNF α appeared only in transgenic animals, as shown in Panel b. A hash mark drawn to the left of each pair of 20 \times images represents the interface between neocortex and internal capsule. Scale bar = 100 μ m

and mouse TNF α mRNA assayed by RT-PCR (Panel a) was fourfold higher in TNF α -Tg than in non-Tg rats ($n = 3$ per animal type; $p \leq 0.01$; unpaired t test). The mean level of rat TNF α mRNA in the cortex of the transgenic rat, 1.3 ± 0.5 attomoles/ μ g total RNA, was not significantly different from 0.8 ± 0.1 in the non-Tg animal and was no more remarkable in cerebellum and brainstem (data not shown). The aggregate level of rat and mouse TNF α protein in whole brain homogenate (Panel b) was fivefold higher in TNF α -Tg rats ($n = 3$ per animal type; $p \leq 0.01$; unpaired t test). In Panel c, we show the results of a second ELISA assaying for rat and mouse TNF α protein in three brain regions (cortex, cerebellum, and brainstem), a variety of organs outside the brain, and in randomly sampled serum in both TNF α -Tg and non-Tg animals ($n = 3$ per animal type). Cortex and brainstem in the TNF α -Tg rat had significantly higher levels ($p \leq 0.001$; ANOVA/Fisher's test). The level of aggregate TNF α protein in the cerebellum of the TNF α -Tg animal (44 ± 18 pg/100 μ g protein) was higher than in the non-Tg rat (7 ± 3) but did not reach statistical significance. It is noteworthy that the levels of aggregate TNF α were almost two-fold higher in the heart, kidney, and skeletal muscle of the transgenic animal in comparison to non-Tg rats, although these differences failed to reach statistical significance. We attributed the difference in quantity of aggregate TNF α protein shown in Panels b and c to the dilutional signal-to-noise loss in the brain homogenate.

Focal cerebral ischemia in the TNF α -Tg rat

We induced focal cerebral ischemia by unilateral MCA suture-occlusion in TNF α -Tg rats and non-Tg littermates. Infarct volumes were calculated after 24 hours or 7 days of post-ischemic reperfusion. The 24-hour infarct volumes were derived by computer-assisted planimetry of digitized, TTC-stained coronal brain slices; all data sets were

corrected for edematous tissue expansion. The 7-day infarct volumes were obtained by the same approach but were derived from examination of coronal brain sections stained with cresyl violet to identify Nissl substance. After 24 hours, TNF α -Tg rats had larger infarctions demonstrated by TTC staining of coronal brain slices (Figure 5, Panels a and b). In the TNF α -Tg rat ($n = 12$), mean infarct volume \pm SD was larger than in non-Tg littermate controls ($n = 17$; $p \leq 0.05$; unpaired t test; see Figure 5, Panel c). Table 1 shows all physiological data obtained on randomly sampled animals subjected to MCAO and measurement of infarct volume by TTC staining. There was no significant group-by-time interaction on two-way ANOVA of any variable. These results support our conclusion that between-group differences in infarct volume could not be

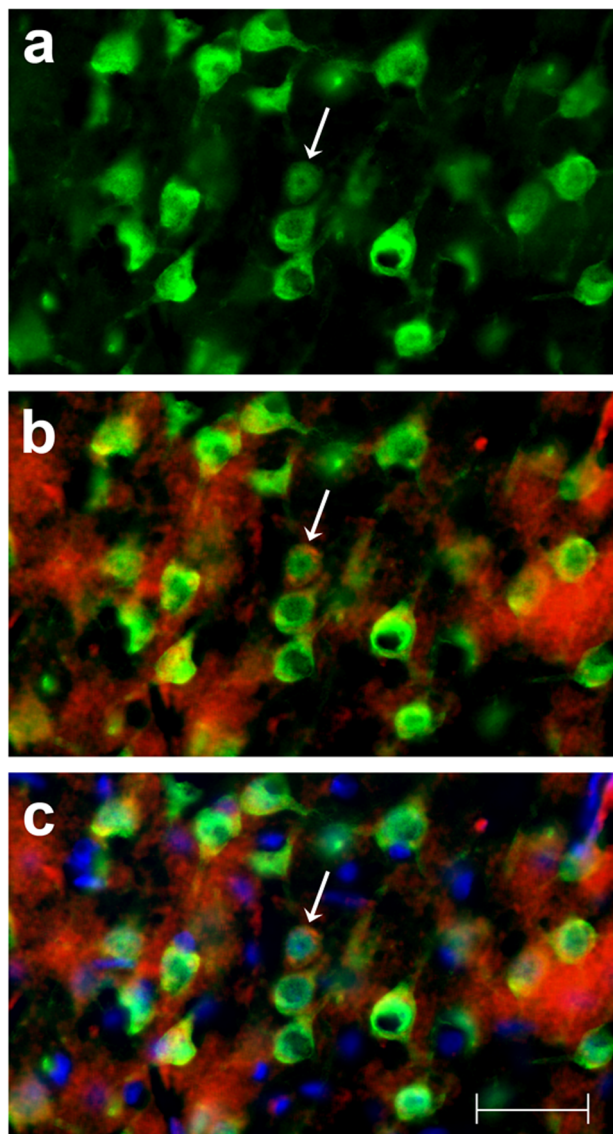


Figure 3

Figure 3
Identification of neural cell type containing TNF α protein. This photographic collage shows the result of co-localization studies identifying the neural cell type containing TNF α protein in the brain of the transgenic animal. All images were taken of TNF α -transgenic rat brain that had not been subjected to ischemic injury. Coronal brain sections were incubated with primary antibodies identifying mouse or rat TNF α protein and neurons. All images were made by fluorescence microscopy at 40 \times magnification. In Panel a, cortical neurons in TNF α -Tg rat brain are immuno-labeled with the anti-neuronal antibody NeuN tagged with a green fluorophore. In Panel b, co-localization images are merged to show that several NeuN-labeled neurons (green) express TNF α protein immuno-labeled with anti-mouse/rat TNF α antibody tagged with a red fluorophore. In Panel c, co-localization images are merged to show NeuN-labeled neurons (green), TNF α protein (red), and cellular nuclei labeled with Hoechst stain (blue). A representative neuron with cytoplasmic inclusions labeled to show expression of TNF α protein is identified in each Panel (arrow). In Panel c, nuclei of several non-neuronal cells that do not express TNF α protein are shown. Scale bar = 50 μ m

attributed to unique susceptibility of the TNF α -Tg rat to hypoxemia or hypercarbia during or after ischemia.

Recognizing that TNF α and other inflammatory cytokines rise to peak levels in rat brain within 24 – 48 hours after ischemic injury [39], we re-assessed infarct volume 7 days after reversal of MCAO to observe for a delayed effect on injury progression. We also wished to know if the difference between infarct volumes in TNF α -Tg and non-Tg rats observed at 24 hours proved resilient at 7 days. We found that TNF α -Tg rats had larger infarcts that persisted after 7 days of post-ischemic reperfusion (Figure 6, Panels a and b). Mean percent infarct volume remained higher in TNF α -Tg rats ($n = 10$) than in non-Tg littermates ($n = 11$; $p \leq 0.01$; unpaired t test; see Figure 6, Panel c). We performed between-group comparisons of mean pre-ischemic body weight, anticipating that TNF α -Tg rats may be affected by somatic wasting that could contribute to the observed difference in infarct volume at 7 days. The pre-ischemic mean weight in TNF α -Tg rats prepared for MCAO and measurement of infarct volume at 7 days was 287.2 ± 33.5 (SD) g, compared to 310.1 ± 21 in non-Tg littermates ($p = \text{NS}$).

To determine if ischemic stress altered the capacity for constitutive synthesis of TNF α protein in the brain of the transgenic animal, we examined regional levels of TNF α protein and mRNA in TNF α -Tg and non-Tg rat brain after 3 or 24 hours of post-ischemic reperfusion. Figure 7 is a composite bar graph showing mean levels of TNF α pro-

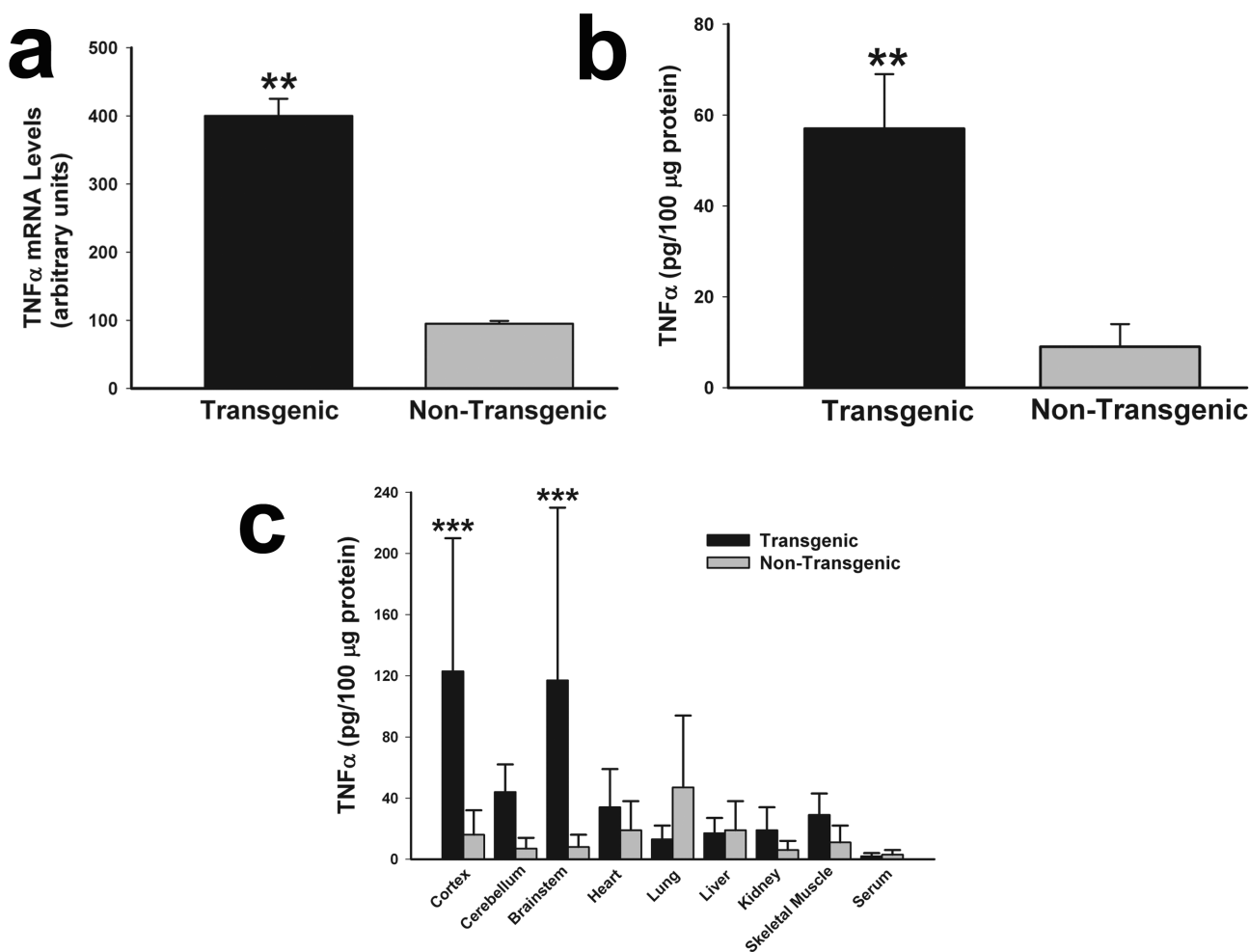


Figure 4
Organ-specific expression of TNFα mRNA and protein in transgenic and non-transgenic rats. Two bar graphs show aggregate rat and mouse TNFα mRNA (Panel a) or protein (Panel b) in brain homogenates prepared from transgenic animals and non-transgenic (non-Tg) littermates (n = 3 in each group). A third bar graph (Panel c) shows TNFα protein levels in three brain compartments, various external organs, and randomly obtained serum samples from a second set of transgenic and non-Tg animals (n = 3 in each group). No animal was subjected to ischemic brain injury. In Panel a, the mean ± SD level of TNFα mRNA was significantly higher in transgenic rats (p ≤ 0.01; unpaired t test). The mean TNFα protein level was also significantly elevated in transgenic animals, as shown in Panel b (p ≤ 0.01; unpaired t test). When TNFα protein was surveyed in multiple organs and in serum, its content in cortex and brainstem was at least fivefold higher than in non-Tg animals (p ≤ 0.001; ANOVA/Fisher's test). **p ≤ 0.01; ***p ≤ 0.001

tein (Panel a) and the corresponding mean ratios of mouse to rat TNFα mRNA (Panel b) in the border zone approximating the ischemic penumbra and in the developing infarct core of both TNFα-Tg and non-Tg rats. The results in Panel a confirm that TNFα protein in control tissue from the transgenic rat is present in almost twice the quantity found in non-Tg littermates, as would be expected from assay of TNFα in non-ischemic brain tissue obtained from both animals shown in Figure 3. After 3 hours of reperfusion in animals subjected to MCAO, mean TNFα levels within the border zone and the devel-

oping infarct core in the TNFα-Tg rat rose above those in the non-Tg animal by statistically insignificant margins. However, mean TNFα levels became significantly elevated within the border zone and infarct core after 24 hours of reperfusion in both TNFα-Tg and non-Tg rats, in comparison to animals of the respective type whether non-ischemic controls or ischemic rats 3 hours after reversal of MCAO (n = 3 - 4 per group; p ≤ 0.001 for all pairwise comparisons; ANOVA/Fisher's test). In addition, the mean TNFα level in the border zone adjacent to the infarct core in the TNFα-Tg rat at 24 hours was significantly

Table 1: Physiological Data.

Test Group	Glucose (mmol/L)	Hematocrit (%)	Core Temperature (°C) (temporalis muscle)	Measurements			
				Mean Arterial Blood Pressure (mm Hg)	Arterial Oxygen Pressure (mm Hg)	Arterial Carbon Dioxide Pressure (mmHg)	Arterial Blood pH (Units)
TNF α -Transgenic Rats (n = 7)							
Pre-ischemic Sample	4.29 \pm 0.38	47 \pm 1	36 \pm 0.4/36 \pm 0.4	99 \pm 10	73 \pm 6	47 \pm 4	7.30 \pm 0.02
Ischemic Sample	-	-	37 \pm 0.2/37 \pm 0.3	84 \pm 13	71 \pm 5	44 \pm 4	7.30 \pm 0.02
Post-ischemic Sample	-	-	37 \pm 0.3/37 \pm 0.5	84 \pm 9	70 \pm 3	43 \pm 4	7.31 \pm 0.02
Non-Transgenic Controls (n = 12)							
Pre-ischemic Sample	4.18 \pm 0.80	47 \pm 2	36 \pm 0.6/36 \pm 0.5	96 \pm 13	71 \pm 8	50 \pm 3	7.29 \pm 0.02
Ischemic Sample	-	-	37 \pm 0.3/36 \pm 0.3	92 \pm 12	69 \pm 8	48 \pm 3	7.29 \pm 0.02
Post-ischemic Sample	-	-	37 \pm 0.4/37 \pm 0.4	88 \pm 14	71 \pm 7	45 \pm 3	7.30 \pm 0.02

Arterial blood samples were obtained 15 minutes before insertion of the suture (pre-ischemic sample), immediately before its removal (ischemic sample), and after 15 minutes of reperfusion (post-ischemic sample).

higher than the comparable level in the non-Tg animal ($p \leq 0.01$; ANOVA/Fisher's test). Mean ratios of mouse and rat mRNA trended upward in all brain regions and at both sampling times in TNF α -Tg rats, compared to non-Tg littermates. However, this trend achieved statistical significance only between control animals that were not subjected to ischemic injury ($p \leq 0.01$; ANOVA/Fisher's test).

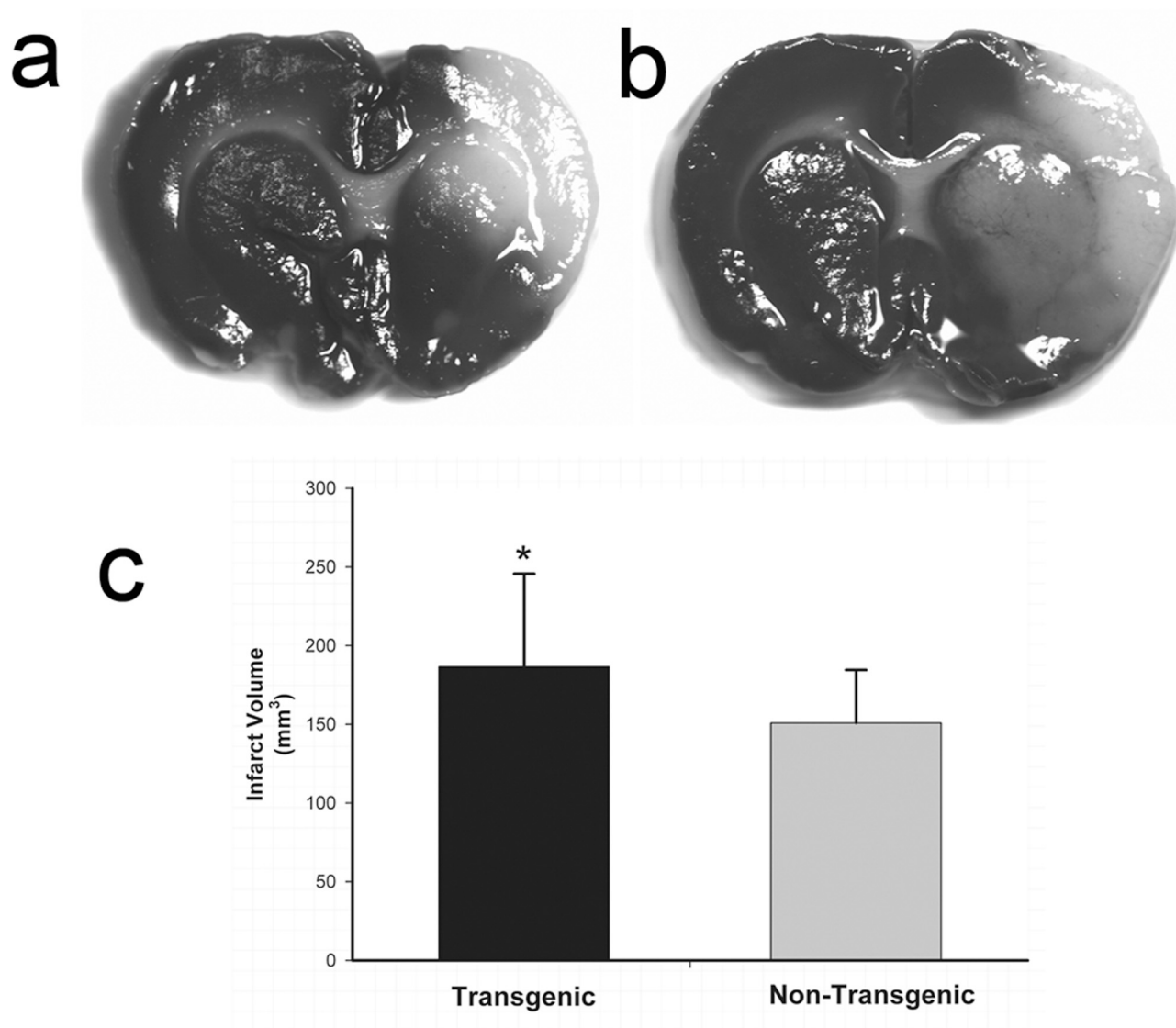
To confirm that TNF α protein synthesized in the brain of the transgenic animal is biologically active, we quantified expression of TRADD protein. Homogenates of brain sampled from TNF α -Tg and non-Tg rats were incubated with a primary antibody that recognizes rat TRADD only after it has been activated through binding of TNF α to p55/TNFR1, thereby implying biological activity of the cytokine. Regional expression of TRADD protein was uniformly higher in TNF α -Tg rat brain than in non-Tg controls by statistically significant margins (see Figure 8). After 3 hours of post-ischemic reperfusion, mean TRADD expression in TNF α -Tg brain was increased in the infarct core ($n = 5$ per animal type; $p \leq 0.05$; ANOVA/Fisher's test) and in the adjacent border zone approximating the ischemic penumbra ($p \leq 0.01$; ANOVA/Fisher's test). To demonstrate that chronically elevated levels of TNF α enhanced expression of TRADD protein without ischemic stress, we sampled cortical tissue from the normally-perfused hemisphere in ischemic animals and found that TRADD was nearly threefold higher in the TNF α -Tg rat ($p \leq 0.01$ compared to non-Tg animals; ANOVA/Fisher's

test). There were no significant differences between TRADD protein expression in the normally-perfused hemisphere and the ischemic border zone or infarct core in the TNF α -Tg rat ($p = NS$).

Apoptotic cell death after focal cerebral ischemia in the TNF α -Tg rat

We performed a qualitative assessment of apoptotic cell death, supported by quantitative assays of caspase-3 activation and DNA fragmentation in the ischemic brain of the TNF α -Tg and non-Tg rat. In Figure 9, co-localization studies in normal or ischemic striatum taken from transgenic and non-Tg animals prove that neurons show cytoplasmic caspase-3 activation while undergoing apoptotic death 24 hours after MCAO. Parallel studies performed to determine if caspase-3 activation was also observed in astroglia showed only rare astrocytes with co-localized GFAP and caspase-3 activation in ischemic striatum of both animals (data not shown). Comparison of Panel d to b in Figure 9 suggests, but does not prove, that degenerating neurons showing caspase-3 activation are more abundant in ischemic striatum within transgenic than in non-Tg brain. Apoptotic bodies were observed within degenerating neurons in ischemic TNF α -Tg rat brain (see Figure 10).

To pursue this observation implying that the TNF α -Tg rat may be susceptible to apoptotic neuronal death, we measured caspase-3 activity in the infarct core and in the border zone region approximating the ischemic penumbra in

**Figure 5**

Quantification of brain infarct volume in TNF α -transgenic and non-transgenic rats at 24 hours. Representative infarcts are shown in the brain of a non-transgenic (Panel a) and a TNF α -transgenic (TNF α -Tg) rat (Panel b). In Panel c, a bar graph shows mean percent infarct volume in both TNF α -Tg and non-Tg rats. All animals underwent suture-occlusion of the right middle cerebral artery for one hour followed by 24 hours of post-ischemic reperfusion. The 1-mm coronal brain sections in Panels a and b were stained for triphenyltetrazolium chloride. Panel a shows infarction (blanched tissue) confined to the right cortical mantle and a portion of the underlying striatum in a non-Tg rat. In the transgenic animal shown in Panel b, there is blanching of tissue in the right cortical mantle that indicates completed infarction and mottled discoloration throughout the entire ipsilateral striatum that is a maturing infarct. Panel c shows that mean \pm SD infarct volume in the transgenic group (n = 12) was significantly greater than that of the non-Tg group (n = 17; *p \leq 0.05; unpaired t test). All volume measurements were corrected for tissue edema.

both animals. In the TNF α -Tg rat, caspase-3 activity was significantly greater than in non-Tg controls within the border zone and in the developing infarct after 3 hours of reperfusion (see Figure 11; n = 3 – 4 per group; p \leq 0.05 for all pairwise comparisons; ANOVA/Fisher's test). At 24 hours, caspase-3 activity remained significantly higher

within the infarct core of the TNF α -Tg rat compared to controls (p \leq 0.05; ANOVA/Fisher's test). As a direct, quantitative measure of apoptotic cell death in ischemic brain, we assayed DNA fragmentation represented by histone-associated DNA complexes released into cellular cytoplasm. In the border zone approximating the

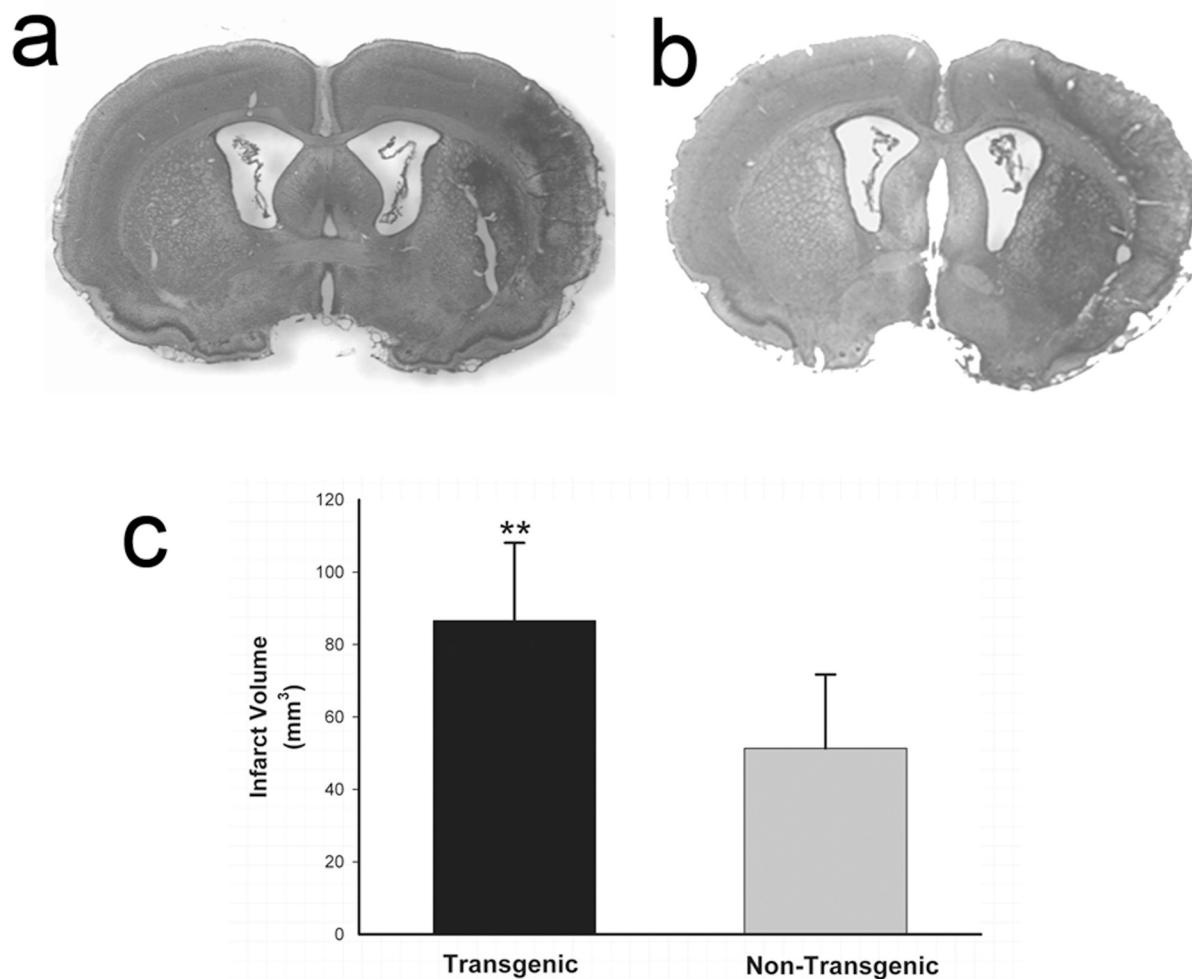


Figure 6

Quantification of brain infarct volume in TNF α -transgenic and non-transgenic rats at 7 days. Representative infarcts are shown in the brain of a non-transgenic (non-Tg; Panel a) and a TNF α -transgenic (TNF α -Tg) rat (Panel b). In Panel c, a bar graph shows mean \pm SD infarct volume in both TNF α -Tg and non-Tg rats. All animals underwent suture-occlusion of the right middle cerebral artery for one hour followed by 7 days of post-ischemic reperfusion. The 80- μ m coronal sections of brain shown in Panels a and b were stained with cresyl violet for Nissl substance. Panel a shows well-demarcated infarction confined to the right cortical mantle and a portion of the underlying striatum in the non-Tg rat. In the transgenic animal shown in Panel b, the infarct occupies a larger proportion of the right cortical mantle and almost all of the ipsilateral striatum. Panel c shows that mean infarct volume in the transgenic group (n = 10) was significantly greater than that of the non-Tg group (n = 11; **p \leq 0.01; unpaired t test).

ischemic penumbra in TNF α -Tg rat brain sampled at 24 hours, mean DNA fragmentation was increased by almost 200-fold compared to non-ischemic transgenic brain (see Figure 12; p \leq 0.0001), over 60-fold compared to border zone tissue in ischemic TNF α -Tg brain sampled at 3 hours (p \leq 0.001), and by 4-fold over that observed in non-Tg brain after 24 hours of post-ischemic reperfusion (p \leq 0.01; n = 6 - 7 per group; all between-group comparisons by ANOVA/Fisher's test). Within the developing infarct

core, DNA fragmentation was again increased by almost 200-fold compared to the non-ischemic transgenic brain (p \leq 0.0001) and 60-fold compared to infarct zone tissue sampled from TNF α -Tg rat brain at 3 hours (p \leq 0.001; all between-group comparisons by ANOVA/Fisher's test). There was no significant between-group difference in elevated mean levels of DNA fragmentation within the infarct core after 24 hours of post-ischemic reperfusion (p = NS). These quantitative results corroborate the findings of

the co-localization studies in Figures 9 and 10 showing that the TNF α -transgenic rat is highly susceptible to apoptotic neuronal death after ischemia.

Cerebral blood flow in the TNF α -Tg rat

After observing larger infarct volumes in the TNF α -Tg rat after 24 hours and 7 days of post-ischemic reperfusion, we were obligated to determine if the apparent susceptibility of the transgenic animal to ischemic injury was related to genetic modification of cerebral vascular reactivity or density. We used laser-Doppler flowmetry for serial measurement of cortical perfusion before, during, and immediately after MCAO to assess cerebral vascular reactivity in TNF α -Tg rats and non-Tg littermates. The multi-line plot in Figure 13 demonstrates that cortical perfusion within the infarct core of the TNF α -Tg rat fell below that of non-Tg controls shortly after MCAO. The TNF α -Tg rats had significantly lower mean LDF at 5 and 10 minutes of ischemia ($n = 19 - 21$ per animal type; $p \leq 0.01$ and $p \leq 0.05$, respectively; repeated measures ANOVA). There were no significant differences in LDF between the two groups of animals during the remaining 50 minutes of ischemia or 30 minutes of post-ischemic reperfusion.

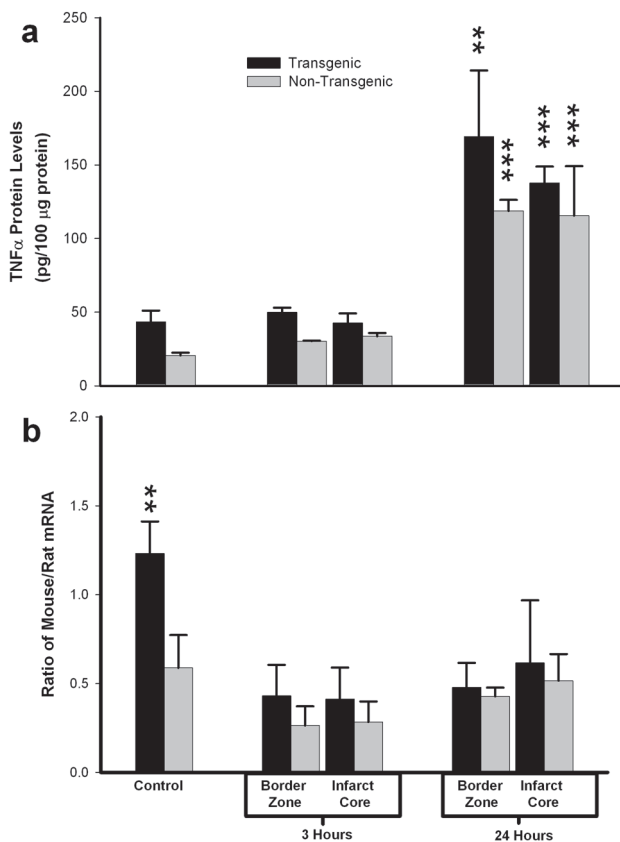


Figure 7

Figure 7

Regional expression of TNF α mRNA and protein in brain after focal ischemia. Two bar graphs show mean \pm SD levels of TNF α protein (Panel a) and the corresponding mean ratios of mouse to rat TNF α mRNA (Panel b) in transgenic and non-transgenic rat brain with or without ischemic brain injury. TNF α -transgenic (TNF α -Tg) and non-Tg rats underwent one hour of focal brain ischemia achieved by unilateral middle cerebral artery occlusion (MCAO). Tissue samples were obtained from the developing infarct core and the border zone approximating the ischemic penumbra in both groups of animals after 3 or 24 hours of post-ischemic reperfusion. As shown in Panel a, mean TNF α levels were significantly elevated within the border zone and infarct core after 24 hours of reperfusion in both TNF α -Tg and non-Tg rats, in comparison to animals of the respective type whether non-ischemic controls or ischemic rats sampled 3 hours after reversal of MCAO ($n = 3 - 4$ per group; $p \leq 0.001$ for all pairwise comparisons; ANOVA/Fisher's test). The mean TNF α level in the border zone adjacent to the infarct core in the transgenic rat at 24 hours was significantly higher than the comparable level in the non-Tg animal ($p \leq 0.01$; ANOVA/Fisher's test). Panel b of Figure 5 shows the mean ratios of mouse and rat TNF α mRNA corresponding to the TNF α protein levels from the same animals reported in Panel a. The mean ratio of mouse to rat TNF α mRNA was significantly higher in TNF α -Tg rats compared to the non-Tg animals only under control conditions without ischemia ($p \leq 0.01$; ANOVA/Fisher's test). ** $p \leq 0.01$; *** $p \leq 0.001$

Stereological analysis of vascular structures in the cortex showed that there was no change in micro-vascular anatomy caused by expression of the transgenic construct in the brain of the TNF α -Tg rat. The mean percent cortical area occupied by vascular structures in the cerebral cortex of the TNF α -Tg rats not subjected to MCAO was 6.20 ± 0.20 , compared to 6.57 ± 0.31 in non-Tg littermates ($p = \text{NS}$). Figure 14 shows representative photomicrographs of micro-vascular structures within the cerebral cortex in both animal types.

Discussion

This work confirms that the TNF α -Tg rat survives to adulthood, reproduces effectively, demonstrates upregulated expression of TNF α mRNA and protein in brain, and is susceptible to infarction by MCAO. Our results show that the high level of TNF α mRNA expressed constitutively in the brain of the transgenic animal is derived almost completely from the murine construct and not from the native rat gene. TNF α protein is increased selectively in the brain and by a statistically insignificant margin in the heart, kidney, and skeletal muscle, but remains low in serum and other external organs. TNF α -immunoreactive cells of neuronal morphology are present only in the neocortex of the transgenic animal. The distribution of the various cell

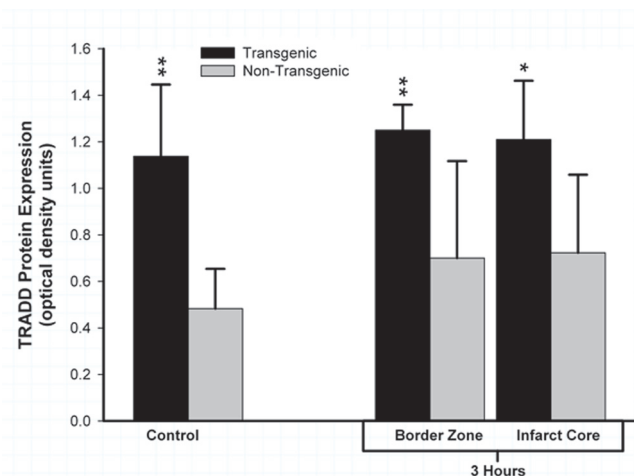


Figure 8
Biological activity of TNF α in transgenic rat brain shown by TRADD protein expression. This bar graph shows expression of TNF receptor-associated protein death domain (TRADD) protein in brain homogenates obtained from TNF α -transgenic (TNF α -Tg) and non-Tg rats. Samples were obtained after 3 hours of reperfusion following one hour of focal cerebral ischemia. TRADD protein expression is shown as mean \pm SD optical density units measured from digitized electrophoretic bands. In ischemic TNF α -Tg brain, TRADD protein was expressed more intensely within the border zone approximating the ischemic penumbra ($n = 5$ per group; $p \leq 0.01$; ANOVA/Fisher's test) and in the developing infarct core ($p \leq 0.05$; ANOVA/Fisher's test) when compared to non-Tg littermates. TRADD was also elevated significantly in the normally-perfused hemisphere (control) of the TNF α -Tg rat compared to non-Tg animals ($p \leq 0.01$; ANOVA/Fisher's test), reflecting increased constitutive expression of protein in response to interaction between active, soluble TNF α and its p55/TNFR1 receptor. There were no significant differences between TRADD protein expression in the normally-perfused hemisphere and the ischemic border zone or infarct core in TNF α -Tg rats ($p = NS$). * $p \leq 0.05$; ** $p \leq 0.01$

populations in the brain of the TNF α -Tg rat appears no different than in the non-Tg animal, suggesting that the transgene has no measurable effect on the structural organization of the neocortex.

Our immunohistochemical studies indicate that the predominant cell type secreting TNF α in the brain of the transgenic rat is of neuronal lineage. This observation of constitutive neuronal expression differs importantly from previous studies showing that TNF α is frequently undetectable in quiescent neural cell populations, becoming upregulated in neurons and astroglial cells only after ischemic or post-traumatic stress [40,41]. In a comprehensive study of the evolution of TNF α expression in the brains of human patients with acute ischemic stroke,

Sairanen and colleagues [6] found that TNF α expression appeared in neurons located within the infarct core and in the adjacent border zone during 24 hours after the onset of infarction. Reactive astrocytes then expressed TNF α immunoreactivity in both the injured hemisphere and in the contralateral hemisphere, beginning within 24 hours of stroke and persisting for up to 18 days afterward. These findings indicate that multiple neural cell types have the capacity to express TNF α , thereby emphasizing that this proinflammatory cytokine participates in an evolving brain injury response culminating in delayed involvement of "unaffected" tissue.

The suture-occlusion model of focal cerebral ischemia is feasible in the TNF α -Tg rat and produces a larger mean volume of infarction than in non-Tg littermates. We speculate that the greater infarct volume observed in the transgenic animal resulted from chronic overexpression of TNF α protein within the brain, causing receptor-mediated caspase activation that promoted neuronal apoptosis [42]. Binding of active, soluble TNF α to p55/TNFR1 will recruit TRADD, activate effector caspase-8, and cleave the executioner caspase-3 for activation [43]. Our observation that TRADD protein is expressed intensely within and outside of the ischemic focus in TNF α -Tg rat brain indicates that the cytokine produced in this animal is biologically active. We found significantly elevated levels of TNF α within ischemic tissue in the brain of the TNF α -Tg rat after 24 hours of post-ischemic reperfusion, but without corresponding increases in the component of mRNA transcribed from the murine TNF α gene. We speculate that the TNF α -Tg rat may have elevated TACE activity to facilitate synthesis of TNF α from its inactive precursor, thereby requiring no additional transcription of TNF α mRNA. Overall, our results suggest that it is soluble TNF α , not its membrane-bound precursor, which is active in the brain of the transgenic animal. Furthermore, our present findings allow us to conclude only that TNF α binds to p55/TNFR1 in the TNF α -Tg rat.

Our supposition that the pivotal mediator of increased lesion volume is the excess brain content of TNF α protein in the transgenic rat is corroborated by the work of Barone and colleagues [13]. To directly examine the role of early TNF α production in the evolution of secondary ischemic injury, these investigators administered exogenous TNF α by intracerebroventricular (ICV) injection 24 hours before MCAO to achieve permanent or transient focal ischemia in rats. They found that motor function, hemispheric edema, and infarct volume were affected adversely in a dose-dependent manner by TNF α . Furthermore, the specificity of the effect of higher doses of TNF α to cause progressively larger infarcts was confirmed by reduction of infarct volume after ICV injections of an anti-TNF α neutralization antibody or of soluble TNFR1 before and after

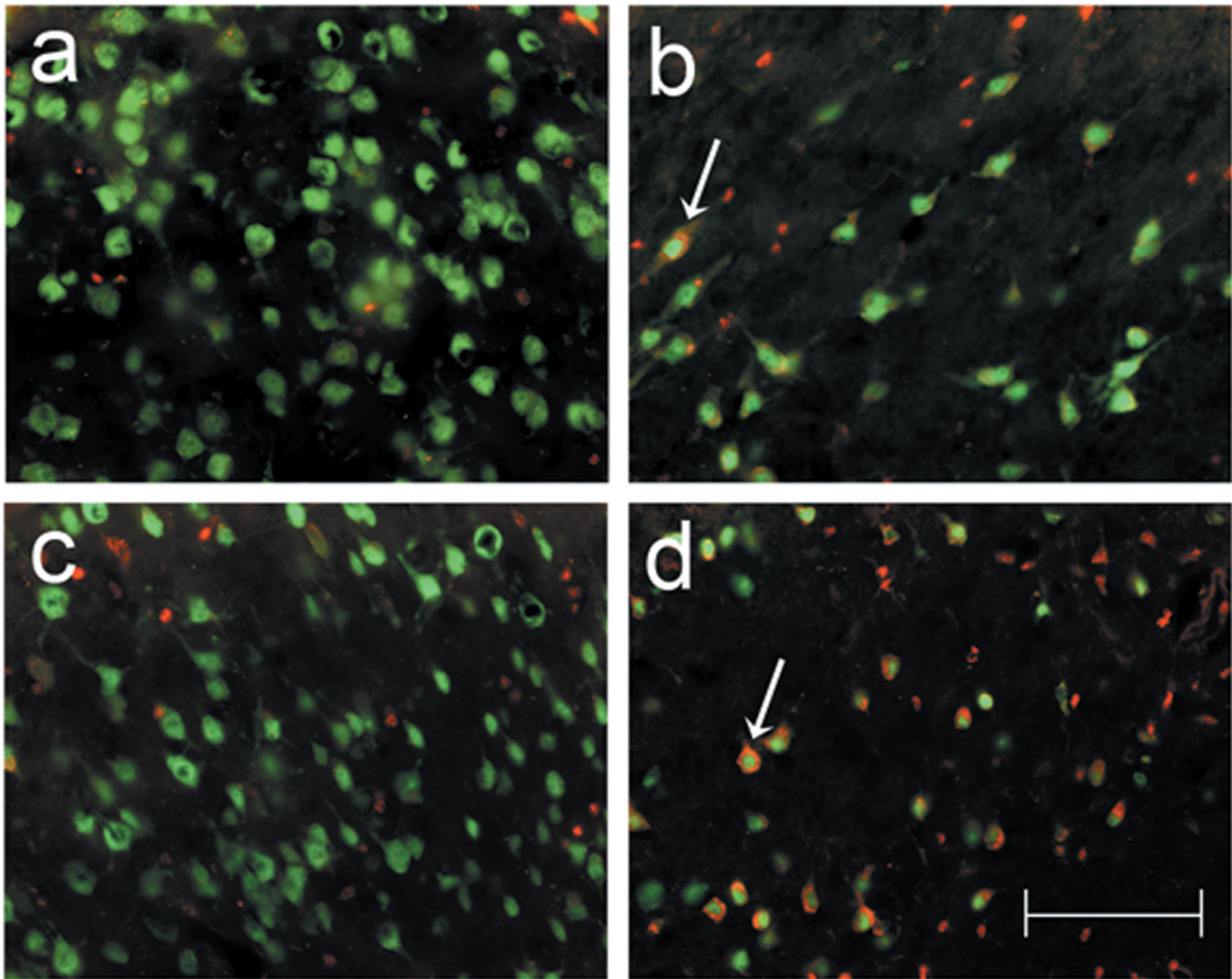


Figure 9
Co-localization studies identifying neuronal apoptosis after focal cerebral ischemia. A collage of 40× photomicrographs shows localization of activated caspase-3 (red) in NeuN-positive striatal neurons (green) after focal cerebral ischemia in TNF α -transgenic (TNF α -Tg) and non-Tg rats. Panels a and c are representative images of striatum in the left cerebral hemisphere of a non-Tg (a) and a TNF α -Tg (c) rat subjected to one hour of suture-occlusion of the right middle cerebral artery, followed by 24 hours of reperfusion. Note the absence of activated caspase-3 in NeuN-positive striatal neurons of the cerebral hemisphere contralateral to the ischemic injury in both animals. Panels b and d are taken from the ischemic striatum in the same non-Tg (b) and TNF α -Tg (d) rats. The presence of activated caspase-3 is clearly evident in NeuN-positive striatal neurons (arrows), many of which exhibit morphological changes consistent with ongoing cell death. Scale bar = 100 μ m.

MCAO. This relationship between lesion volume and ambient level of TNF α has been confirmed repeatedly in animal models of focal cerebral ischemia [14,15,44,45] and has even been observed in human stroke victims. Montaner and co-workers [46] found positive correlations between elevated blood levels of TNF α or interleukin-6 and the volume of hypoperfused brain observed after bolus-tracking perfusion MRI studies in stroke patients.

We examined brain perfusion by LDF to determine if the larger mean infarct volume in the TNF α -Tg rat was caused by an unanticipated effect of the constitutively expressed TNF α mRNA or translated protein on the cerebral circulation. We found that cortical perfusion within the infarct core in the brain of the TNF α -Tg rat fell to 20% of pre-ischemic baseline within 5 minutes after MCAO. Beyond the first 10 minutes of MCAO, there was no significant difference in cortical flow between transgenic and non-Tg animals for the remaining 50 minutes of focal ischemia

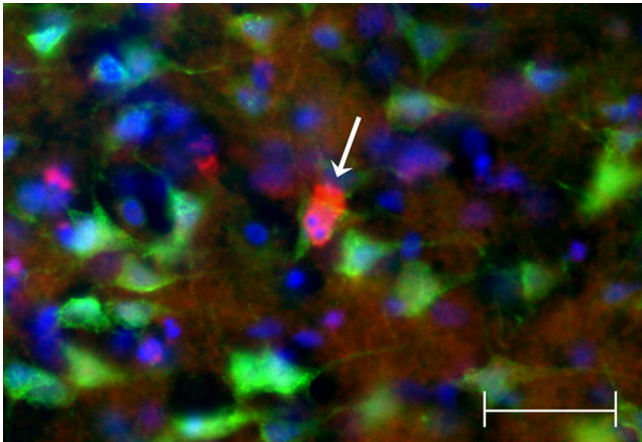


Figure 10
Co-localization study revealing apoptotic bodies within degenerating neuron in ischemic TNF α -transgenic rat brain. A single 40 \times photomicrograph shows localization of activated caspase-3 (red) in NeuN-positive cortical neurons (green) after focal cerebral ischemia in the TNF α -transgenic (TNF α -Tg) rat. The right cerebral cortex was sampled after one hour of suture occlusion of the ipsilateral middle cerebral artery, followed by 24 hours of reperfusion. In the representative NeuN-positive neuron identified by the arrow, there is caspase-3 activation with apoptotic bodies identified by Hoechst staining (blue). Scale bar = 50 μ m.

and 30 minutes of post-ischemic reperfusion. This result suggests that constitutive upregulation of TNF α protein could accentuate brain response to focal circulatory arrest and may have contributed to the larger mean percent infarct volume observed in transgenic animals. However, the transience of the effect on LDF in the ischemic TNF α -Tg rat is self-evident and was not corroborated by underlying changes in micro-vascular anatomy, as was shown in our stereological analysis of cortical vascular density.

Our speculation of a link between elevated TNF α in brain and cortical perfusion is corroborated by Lavine and colleagues [45] who found that pre-ischemic administration of an anti-TNF α neutralization antibody one hour before MCAO significantly improved perfusion in the infarct core and in the peri-infarct region after reversal of MCAO in non-Tg animals. Alternately, Savitz and others [47] showed that pre-ischemic administration of carvedilol, a mixed α and β adrenoreceptor antagonist, reduced TNF α mRNA copy number, infarct volume, and ischemic apoptosis after MCAO but had no effect on LDF. Recent studies suggest that TNF α may affect brain endothelial cell function or microvascular integrity, even if its influence on regulation of cerebral hemodynamic function after ischemia remains ambivalent. Kimura and coworkers [48] reported that TNF α induced cleavage of caspase-3 and promoted apoptosis in cultured brain microvascular endothelial

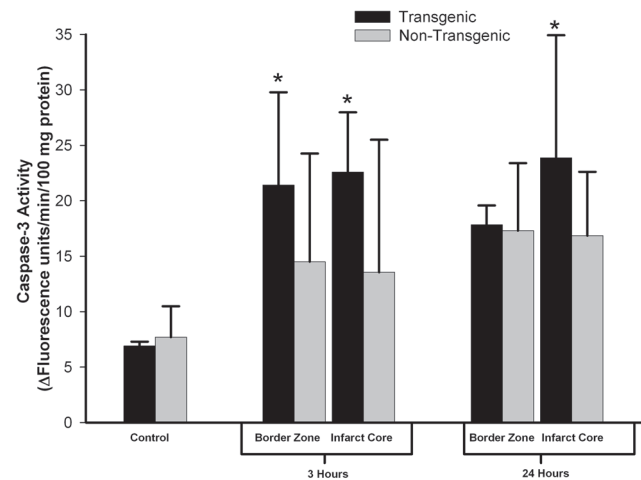


Figure 11
Caspase-3 activity in TNF α -transgenic and non-transgenic rat brain after focal ischemia. The bar graph shows caspase-3 activity in the brains of TNF α -transgenic (TNF α -Tg) and non-Tg rats (n = 3 – 4 per group and sampling time) with or without ischemic brain injury. TNF α -transgenic (TNF α -Tg) and non-Tg rats underwent one hour of focal brain ischemia achieved by unilateral middle cerebral artery occlusion. Tissue samples were obtained from the developing infarct core and the border zone approximating the ischemic penumbra in both groups of animals after 3 or 24 hours of post-ischemic reperfusion. Caspase-3 activity is presented as Δ fluorescence units/minute/100 mg protein (mean \pm SD). Transgenic animals had significantly greater caspase-3 activity in the border zone region after 3 hours of reperfusion and in the infarct core at both 3 and 24 hours (p \leq 0.05; ANOVA/Fisher's test). *p \leq 0.05 compared to respective control.

cells. Hosomi and others [44] found that TNF α may increase microvascular permeability after ischemia by upregulating matrix metalloproteinases (MMPs). They observed that intravenous infusion of an anti-TNF α neutralization antibody immediately after reversal of MCAO inhibited upregulation of MMP-9 and membrane type 1-MMP, thereby lowering infarct volume and attenuating ischemic brain edema.

We have confidence that the unique cellular anatomy and the response to ischemic brain injury observed in our transgenic animal cannot be attributed to random genetic heterogeneity. Transgenic animals are often produced on a hybrid background and then backcrossed to optimize genetic homogeneity. The TNF α -Tg rats were produced on the outbred S-D rat strain. All subsequent generations of transgenic animals have been prepared by backcrossing onto this same strain. We have used non-Tg littermates as controls in all the assays we performed to quantify TNF α mRNA and protein levels and verify cellular anatomy.

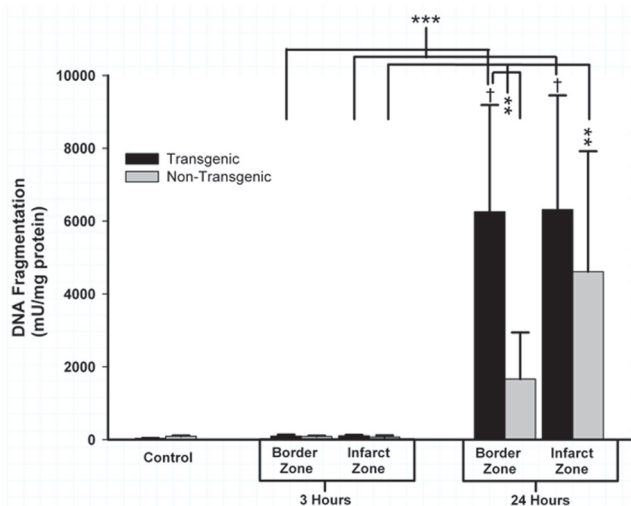


Figure 12
Quantification of apoptotic cell death by DNA fragmentation in TNF α -transgenic and non-transgenic rat brain after focal ischemia. The bar graph shows DNA fragmentation representing apoptotic cell death in brain sampled from TNF α -transgenic (TNF α -Tg) and non-Tg rats (n = 6 – 7 per group and sampling time) after one hour of unilateral middle cerebral artery occlusion (MCAO). Tissue samples were obtained from the developing infarct core and the border zone approximating the ischemic penumbra in both groups of animals after 3 or 24 hours of post-ischemic reperfusion. Neural cell death represented by DNA fragmentation is presented as mU/mg protein (mean \pm SE). Cell death represented by DNA fragmentation was present at only negligible levels in non-ischemic TNF α -Tg and non-Tg controls (n = 12 per group) and after MCAO followed by 3 hours of post-ischemic reperfusion (p = NS for all pairwise comparisons). However, DNA fragmentation increased markedly after 24 hours of post-ischemic reperfusion in both TNF α -Tg and non-Tg rats. In the border zone approximating the ischemic penumbra in TNF α -Tg brain sampled at 24 hours, mean DNA fragmentation was increased by almost 200-fold compared to the respective non-ischemic control (p \leq 0.0001; ANOVA/Fisher's test) and over 60-fold compared to border zone tissue sampled from TNF α -Tg brain at 3 hours (p \leq 0.001; ANOVA/Fisher's test). DNA fragmentation in this region sampled from TNF α -Tg brain was almost 4-fold higher than that observed in non-Tg brain after 24 hours of post-ischemic reperfusion (p \leq 0.01; ANOVA/Fisher's test). Within the infarct zone of TNF α -Tg brain sampled at 24 hours, mean DNA fragmentation was again increased by almost 200-fold compared to the respective non-ischemic control (p \leq 0.0001; ANOVA/Fisher's test) and 60-fold compared to infarct zone tissue sampled from TNF α -Tg brain at 3 hours (p \leq 0.001; ANOVA/Fisher's test). There was no significant difference observed between the elevated mean levels of DNA fragmentation within the infarct zone in TNF α -Tg and non-Tg rat brain after 24 hours of post-ischemic reperfusion (p = NS). **p \leq 0.01; ***p \leq 0.001; †p \leq 0.0001

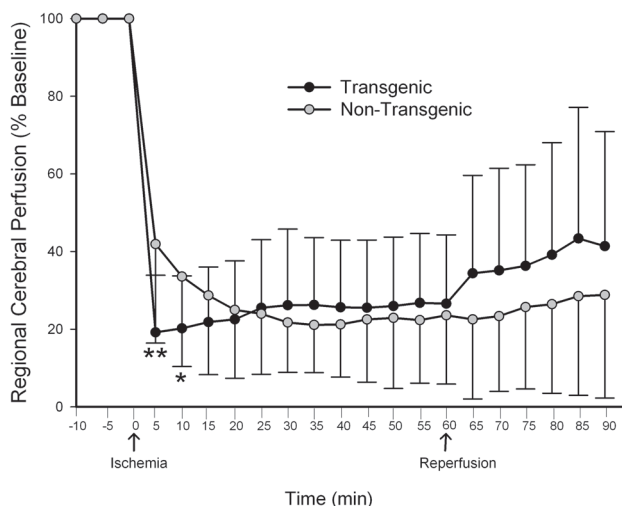


Figure 13
Measurement of cortical perfusion during focal cerebral ischemia in TNF α -transgenic and non-transgenic rats. The multi-line plot shows laser-Doppler flowmetry (LDF) measurements of cortical perfusion expressed as a percentage of pre-ischemic baseline in TNF α -transgenic (TNF α -Tg; n = 19) rats and non-Tg littermates (n = 21). Measurements were made every 5 minutes during 10 minutes of pre-ischemic baseline, 60 minutes of ischemia, and 30 minutes of post-ischemic reperfusion. Values were calculated as a percentage of pre-ischemic baseline and are presented as means \pm SD. The TNF α -Tg rats had significantly lower mean LDF at 5 and 10 minutes immediately after the onset of ischemia (P = 0.0016 and 0.034, respectively; repeated measures ANOVA). *p \leq 0.05; **p \leq 0.01

TNF α -Tg animals express a uniform phenotype that differs consistently from the non-Tg littermates. Therefore, we believe that the unique features of the TNF α -Tg rat must be attributed to segregation of the transgene allele.

With its chronic upregulation of TNF α in brain, the TNF α -Tg rat is a clinically relevant model of secondary inflammatory changes after cerebral infarction in humans. Although non-Tg animals show increased levels of endogenous TNF α in response to ischemia, the transgenic rat has greater research potential because of the ease with which signal transduction and pro-apoptotic pathways may be studied under conditions of prolonged activation *in vivo*. Even though TNF α -Tg mice share this asset, their role in the assessment of experimental therapeutic agents will be limited by small size, restricted vascular access, and constrained use in functional assessment of motor and cognitive behavior. The TNF α -Tg rat is a valuable new tool for the study of cytokine-mediated signal transduction paths in ischemic brain injury.

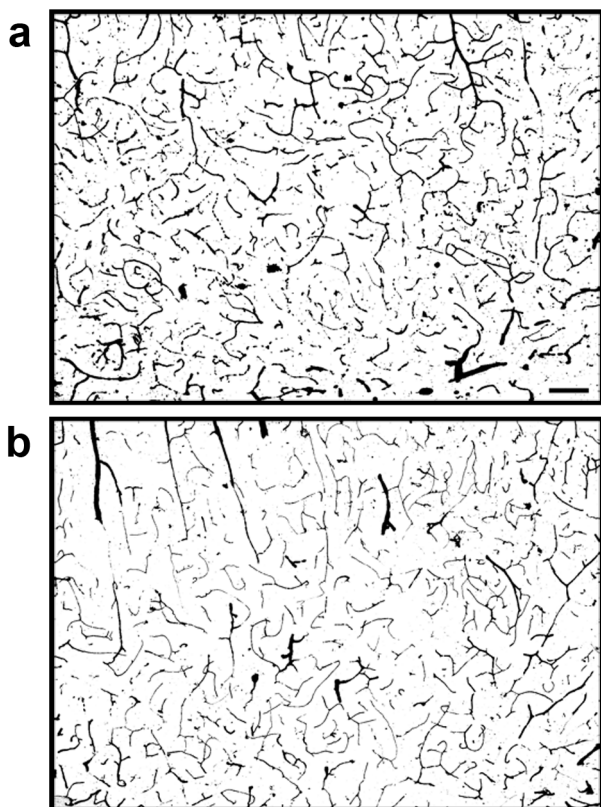


Figure 14
Micro-vascular structures in the brains of TNF α -transgenic and non-transgenic rats. These representative photomicrographs show ink-stained micro-vascular structures within the cortex of the TNF α -transgenic rat (Panel a) and a non-transgenic littermate (Panel b). There was no significant difference between samples in the number of micro-vascular structures quantified by stereology. Scale bar = 100 μ m

Competing interests

David S. Grass, Ph.D., is a salaried employee of Xenogen Biosciences, Cranbury, New Jersey, USA. Xenogen Biosciences provided no financial support for the submitted work, will not realize financial gain from publication of the work, and did not pay the article-processing charge. All other authors declare that they have no financial or non-financial competing interests.

Authors' contributions

LCP conceived of the study, supervised its design and coordination, and wrote the manuscript. MSK obtained the transgenic animal described in the study and contributed to the manuscript. SS performed quantitative stereology and contributed to the manuscript. JES performed histological studies and contributed to the manuscript. RJK performed statistical analyses and contributed to the

manuscript. YL conducted preliminary studies with the transgenic animal. DSG provided the transgenic animal used in the study and contributed to the manuscript. All authors read and approved the final manuscript.

Acknowledgements

Financial support was provided by NIH/NINDS grants R01 NS047395 (LCP) and R01 NS039588 (MSK) and by a Visiting Research Scientist Award provided by Shandong University, China (YL) for training at the University of Kentucky in Lexington, Kentucky. Dr. George Kollias prepared the microinjection construct containing the murine TNF α gene and its promoter that was used to make the transgenic animal. Mr. Richard Coffee constructed the animal at Xenogen Biosciences in Cranbury, New Jersey. We thank Mary Louise Holtz, Ph.D., Xiaio-Hong Song, M.D., Susan D. Craddock, and Ravikumar Rao for technical contributions to this work and Sherry Chandler Williams, ELS, for editing the manuscript and preparing the illustrations.

References

1. Emsley HC, Tyrrell PJ: **Inflammation and infection in clinical stroke.** *J Cereb Blood Flow Metab* 2002, **22**:1399-1419.
2. Mayne M, Ni W, Yan HJ, Xue M, Johnston JB, Del Bigio MR, Peeling J, Power C: **Antisense oligodeoxynucleotide inhibition of tumor necrosis factor- α expression is neuroprotective after intracerebral hemorrhage.** *Stroke* 2001, **32**:240-248.
3. Tomimoto H, Ihara M, Wakita H, Ohtani R, Lin JX, Akiguchi I, Kinoshita M, Shibasaki H: **Chronic cerebral hypoperfusion induces white matter lesions and loss of oligodendroglia with DNA fragmentation in the rat.** *Acta Neuropathol (Berlin)* 2003, **106**:527-534.
4. Lotocki G, Alonso OF, Dietrich WD, Keane RW: **Tumor necrosis factor receptor 1 and its signaling intermediates are recruited to lipid rafts in the traumatized brain.** *J Neurosci* 2004, **24**:11010-11016.
5. Tomimoto H, Akiguchi I, Wakita H, Kinoshita A, Ikemoto A, Nakamura S, Kimura J: **Glial expression of cytokines in the brains of cerebrovascular disease patients.** *Acta Neuropathol (Berlin)* 1996, **92**:281-287.
6. Sairanen T, Carpén O, Karjalainen-Lindsberg ML, Paetau A, Turpeinen U, Kaste M, Lindsberg PJ: **Evolution of cerebral tumor necrosis factor- α production during human ischemic stroke.** *Stroke* 2001, **32**:1750-1758.
7. Carlstedt F, Lind L, Lindahl B: **Proinflammatory cytokines, measured in a mixed population on arrival in the emergency department, are related to mortality and severity of disease.** *J Intern Med* 1997, **242**:361-365.
8. Vila N, Castillo J, Dávalos A, Chamorro A: **Proinflammatory cytokines and early neurological worsening in ischemic stroke.** *Stroke* 2000, **31**:2325-2329.
9. Zaremba J, Skrobanski P, Losy J: **Tumor necrosis factor- α is increased in the cerebrospinal fluid and serum of ischaemic stroke patients and correlates with the volume of evolving brain infarct.** *Biomed Pharmacother* 2001, **55**:258-263.
10. Sairanen TR, Lindsberg PJ, Brenner M, Carpén O, Sirén AL: **Differential cellular expression of tumor necrosis factor- α and Type I tumor necrosis factor receptor after transient global forebrain ischemia.** *J Neurol Sci* 2001, **186**:87-99.
11. Saito K, Suyama K, Nishida K, Sei Y, Basile AS: **Early increases in TNF- α , IL-6, and IL-1 β levels following transient cerebral ischemia in gerbil brain.** *Neurosci Lett* 1996, **206**:149-152.
12. Liu Y, Jacobowitz DM, Barone F, McCarron R, Spatz M, Feuerstein G, Hallenbeck JM, Siren AL: **Quantitation of perivascular monocytes and macrophages around cerebral blood vessels of hypertensive and aged rats.** *J Cereb Blood Flow Metab* 1994, **14**:348-352.
13. Barone FC, Arvin B, White RF, Miller A, Webb CL, Willette RN, Lysko PG, Feuerstein GZ: **Tumor necrosis factor- α : a mediator of focal ischemic brain injury.** *Stroke* 1997, **28**:1233-1244.
14. Dawson DA, Martin D, Hallenbeck JM: **Inhibition of tumor necrosis factor- α reduces focal cerebral ischemic injury in the spontaneously hypertensive rat.** *Neurosci Lett* 1996, **218**:41-44.

15. Nawashiro H, Martin D, Hallenbeck JM: **Inhibition of tumor necrosis factor and amelioration of brain infarction in mice.** *J Cereb Blood Flow Metab* 1997, **17**:229-232.
16. Nawashiro H, Tasaki K, Ruetzler CA, Hallenbeck JM: **TNF-alpha pretreatment induces protective effects against focal cerebral ischemia in mice.** *J Cereb Blood Flow Metab* 1997, **17**:483-490.
17. Sharif SF, Hariri RJ, Chang VA, Barie PS, Wang RS, Ghajar BG: **Human astrocyte production of tumor necrosis factor- α , interleukin-1 β and interleukin-6 following exposure to lipopolysaccharide endotoxin.** *Neurol Res* 1993, **15**:109-112.
18. Sawada M, Kondo N, Suzumura A, Marunouchi T: **Production of tumor necrosis factor- α by microglia and astrocytes in culture.** *Brain Res* 1989, **491**:394-397.
19. Wajant H, Pfizenmaier K, Scheurich P: **Tumor necrosis factor signaling.** *Cell Death Differ* 2003, **10**:45-65.
20. Bruce AJ, Boling W, Kindy MS, Peschon J, Kraemer PJ, Carpenter MK, Holtzberg FW, Mattson MP: **Altered neuronal and microglial responses to excitotoxic and ischemic brain injury in mice lacking TNF receptors.** *Nat Med* 1996, **2**:788-794.
21. Gary DS, Bruce-Keller AJ, Kindy MS, Mattson MP: **Ischemic and excitotoxic brain injury is enhanced in mice lacking the p55 tumor necrosis factor receptor.** *J Cereb Blood Flow Metab* 1998, **18**:1283-1287.
22. Probert L, Akassoglou K, Pasparakis M, Kontogeorgos G, Kollias G: **Spontaneous inflammatory demyelinating disease in transgenic mice showing central nervous system-specific expression of tumor necrosis factor.** *Proc Natl Acad Sci USA* 1995, **92**:11294-11298.
23. Kollias G, Hurst J, deBoer E, Grosveld F: **The human beta-globin gene contains a downstream developmental specific enhancer.** *Nucleic Acid Res* 1987, **15**:5739-5747.
24. Brinster RL, Chen HY, Trumbauer ME, Yagle MK, Palmiter RD: **Factors affecting the efficiency of introducing foreign DNA into mice by microinjecting eggs.** *Proc Natl Acad Sci USA* 1985, **82**:4438-4442.
25. Holtz ML, Craddock S, Pettigrew LC: **Rapid expression of neuronal and inducible nitric oxide synthases during post-ischemic reperfusion in rat brain.** *Brain Res* 2001, **898**:49-60.
26. Minger SL, Geddes JW, Holtz ML, Craddock SD, Whiteheart SW, Siman RG, Pettigrew LC: **Glutamate receptor antagonists inhibit calpain-mediated cytoskeletal proteolysis in focal cerebral ischemia.** *Brain Res* 1998, **810**:181-199.
27. Pettigrew LC, Holtz ML, Craddock SD, Minger SL, Hall N, Geddes JW: **Microtubular proteolysis in focal cerebral ischemia.** *J Cereb Blood Flow Metab* 1996, **16**:1189-1202.
28. Longa EZ, Weinstein PR, Carlson S, Cummins R: **Reversible middle cerebral artery occlusion without craniotomy in rats.** *Stroke* 1989, **20**:84-91.
29. Dubal DB, Kashon ML, Pettigrew LC, Ren JM, Finklestein SP, Rau SW, Wise PM: **Estradiol protects against ischemic injury.** *J Cereb Blood Flow Metab* 1998, **18**:1253-1258.
30. Belayev L, Alonso OF, Busto R, Zhao W, Ginsberg MD: **Middle cerebral artery occlusion in the rat by intraluminal suture. Neurological and pathological evaluation of an improved model.** *Stroke* 1996, **27**:1616-1623.
31. Bederson JB, Pitts LH, Germano SM, Nishimura MC, Davis RL, Bartkowski HM: **Evaluation of 2,3,5-triphenyltetrazolium chloride as a stain for detection and quantification of experimental cerebral infarction in rats.** *Stroke* 1986, **17**:1304-1308.
32. Swanson RA, Morton MT, Tsao-Wu G, Savalos RA, Davidson C, Sharp FR: **A semiautomated method for measuring brain infarct volume.** *J Cereb Blood Flow Metab* 1990, **10**:290-293.
33. McEwen ML, Springer JE: **A mapping study of caspase-3 activation following acute spinal cord contusion in rats.** *J Histochem Cytochem* 2005, **53**:809-819.
34. Shimizu H, Ohgoh M, Ikeda M, Nishizawa Y, Ogura H: **Caspase-3-like protease activity-independent apoptosis at the onset of neuronal cell death in the gerbil hippocampus after global ischemia.** *Biol Pharm Bull* 2007, **30**:1950-1953.
35. Cholet N, Seylaz J, Lacombe P, Bonvento G: **Local uncoupling of the cerebrovascular and metabolic responses to somatosensory stimulation after neuronal nitric oxide synthase inhibition.** *J Cereb Blood Flow Metab* 1997, **17**:1191-1201.
36. Scheff SW, Bernardo LS, Cotman CW: **Decrease in adrenergic axon sprouting in the senescent rat.** *Science* 1978, **202**:775-778.
37. Paxinos G, Watson C: *The rat brain in stereotaxic coordinates* 4th edition. San Diego: Academic Press; 1998.
38. Mouton PR: *Principles and practices of unbiased stereology: an introduction for bioscientists* Baltimore: Johns Hopkins University Press; 2002.
39. Allan SM, Rothwell NJ: **Cytokines and acute neurodegeneration.** *Nat Rev Neurosci* 2001, **2**:734-744.
40. Feuerstein GZ, Wang X, Barone FC: **The role of cytokines in the neuropathology of stroke and neurotrauma.** *Neuroimmunomodulation* 1998, **5**:143-159.
41. Harrington JF, Messier AA, Levine A, Szmydynger-Chodobska J, Chodobski A: **Shedding of tumor necrosis factor type I receptor after experimental spinal cord injury.** *J Neurotrauma* 2005, **22**:919-928.
42. Martin-Villalba A, Herr I, Jeremias I, Hahne M, Brandt R, Vogel J, Schenkel J, Herdegen T, Debatin KM: **CD95 ligand (Fas-L/APO-1L) and tumor necrosis factor-related apoptosis-inducing ligand mediate ischemia-induced apoptosis in neurons.** *J Neurosci* 1999, **19**:3809-3817.
43. Muzio M, Salvesen GS, Dixit VM: **FLICE induced apoptosis in a cell-free system. Cleavage of caspase zymogens.** *J Biol Chem* 1997, **272**:2952-2956.
44. Hosomi N, Ban CR, Naya T, Takahashi T, Guo P, Song XY, Kohno M: **Tumor necrosis factor-alpha neutralization reduced cerebral edema through inhibition of matrix metalloproteinase production after transient focal cerebral ischemia.** *J Cereb Blood Flow Metab* 2005, **25**:959-967.
45. Lavine SD, Hofman FM, Zlokovic BV: **Circulating antibody against tumor necrosis factor-alpha protects rat brain from reperfusion injury.** *J Cereb Blood Flow Metab* 1998, **18**:52-58.
46. Montaner J, Rovira A, Molina CA, Arenillas JF, Ribo M, Chacon P, Monasterio J, Alvarez-Sabin J: **Plasmatic level of neuroinflammatory markers predict the extent of diffusion-weighted image lesions in hyperacute stroke.** *J Cereb Blood Flow Metab* 2003, **23**:1403-1407.
47. Savitz SI, Erhardt JA, Anthony JV, Gupta G, Li X, Barone FC, Rosenbaum DM: **The novel beta-blocker, carvedilol, provides neuroprotection in transient focal stroke.** *J Cereb Blood Flow Metab* 2000, **20**:1197-1204.
48. Kimura H, Gules I, Meguro T, Zhang JH: **Cytotoxicity of cytokines in cerebral microvascular endothelial cell.** *Brain Res* 2003, **990**:148-156.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

