

University of Kentucky

UKnowledge

---

Animal and Food Sciences Faculty Publications

Animal and Food Sciences

---

4-12-2017

## Glucans and the Poultry Immune System

Jacque Jacob

*University of Kentucky*, jacque.jacob@uky.edu

Anthony J. Pescatore

*University of Kentucky*, anthony.pescatore@uky.edu

Follow this and additional works at: [https://uknowledge.uky.edu/animalsci\\_facpub](https://uknowledge.uky.edu/animalsci_facpub)



Part of the [Animal Sciences Commons](#), [Food Science Commons](#), and the [Immunology and Infectious Disease Commons](#)

[Right click to open a feedback form in a new tab to let us know how this document benefits you.](#)

---

### Repository Citation

Jacob, Jacquie and Pescatore, Anthony J., "Glucans and the Poultry Immune System" (2017). *Animal and Food Sciences Faculty Publications*. 16.

[https://uknowledge.uky.edu/animalsci\\_facpub/16](https://uknowledge.uky.edu/animalsci_facpub/16)

This Article is brought to you for free and open access by the Animal and Food Sciences at UKnowledge. It has been accepted for inclusion in Animal and Food Sciences Faculty Publications by an authorized administrator of UKnowledge. For more information, please contact [UKnowledge@lsv.uky.edu](mailto:UKnowledge@lsv.uky.edu).

---

## Glucans and the Poultry Immune System

Digital Object Identifier (DOI)

<https://doi.org/10.3844/ajisp.2017.45.49>

### Notes/Citation Information

Published in *American Journal of Immunology*, v. 13, issue 1, p. 45-49.

© 2017 Jacqueline Jacob and Anthony Pescatore.

This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

# Glucans and the Poultry Immune System

Jacqueline Jacob and Anthony Pescatore

Department of Animal and Food Sciences, University of Kentucky, KY, USA

## Article history

Received: 15-11-2016

Revised: 10-01-2017

Accepted: 29-03-2017

## Corresponding Author:

Jacqueline Jacob

Department of Animal and Food Sciences, University of Kentucky, KY, USA

Tell: 859-257-7613

Email: Jacquie.jacob@uky.edu

**Abstract:** With the reduced availability of antibiotics, poultry producers are looking for feed additives to stimulate the immune system of their birds. Some beta-glucans have been shown to improve gut health in poultry subjected to a bacterial challenge, to increase the flow of new immunocytes into the various lymphoid organs, to increase macrophage function, to increase antibody titers after a vaccination and to function as an anti-inflammatory immunomodulator. As a result, beta-glucans may provide a tool for producers trying to reduce or eliminate the use of antibiotics in poultry diets.

**Keywords:** Glucans, Poultry, Immunity

## Introduction

Bacterial infections in poultry are an important concern for both animal health and productivity. In addition, some bacteria are food borne pathogens like *Salmonella* and *Campylobacter*. The subtherapeutic use of antibiotics has reduced the impact of bacterial infections, but has led to concerns regarding development of antibiotic resistance. The result has been the withdrawal of several antibiotics from the toolbox available to poultry producers. As poultry producers work to reduce the use of antimicrobials, alternative management strategies are being developed. There has been increased interest in 'natural' feed additives that can stimulate the immune system of poultry. There has been some research to show that  $\beta$ -glucans may play a role in replacing antibiotics and stimulating the immune system. Rathgeber *et al.* (2008) reported that yeast  $\beta$ -glucan was as effective in promoting growth of broiler chickens as virginiamycin, an antibiotic routinely added to broiler diets at subtherapeutic levels. Similarly, Zhang *et al.* (2008) reported that supplementing broiler diets with  $\beta$ -glucans can improve growth performance in terms of both average daily gain and feed conversion ratio.

The immunomodulatory effects of  $\beta$ -glucans have been well documented in mammals, but, while the mammalian and avian immune systems are similar, they are different enough that it is not possible to extrapolate mammal research to poultry.

## Glucans

Glucans are complex glucose polymers that are structural components of the cell walls of many organisms including bacteria, yeast, fungi and algae. They are also found in the cereals oats and barley. The six-sided glucose rings are connected together in linear or branched forms with glycosidic linkages. The nature of these linkages will affect the functionality of the molecules. The glucans have a backbone of glucose molecules linked at the 1 and 3 carbon atoms. Bacterial glucans are simply this backbone of glucose molecules. Fungal and yeast glucans, however, have side-chains linked at the 1 and 6 carbon atoms. This gives them a branched structure. Comparing the two, fungal glucans typically have shorter side chains than those found in yeast. By comparison, barley glucans have the glucose molecules linked at the 1 and 4 carbon atoms with some linkages between the 1 and 3 carbon atoms. The 1 $\rightarrow$ 3 linkages give the molecule a semi-flexible nature and the 1 $\rightarrow$ 4 add kinks to that chain.

Table 1. Types of  $\beta$ -glucan linkages generally found in different sources

Source	Linkage
Bacteria	(1 $\rightarrow$ 3)
Fungal	(1 $\rightarrow$ 3)(1 $\rightarrow$ 6) with short branches
Yeast	(1 $\rightarrow$ 3)(1 $\rightarrow$ 6) with longer branches
Cereal (barley and oats)	(1 $\rightarrow$ 3)(1 $\rightarrow$ 4)

## Effect of $\beta$ -Glucan Supplementation on Poultry Immune System

The intestinal lining provides the innate defense barrier against most intestinal pathogens. A *Salmonella* Typhimurium challenge has been shown to have major negative effects on the intestinal mucosa (Shao *et al.*, 2013). There is a significant reduction in villus height and the villus height to crypt depth ratio in the jejunum. Dietary yeast  $\beta$ -glucan supplementation for chicks challenged with *S. Typhimurium*, however, resulted in higher villus height and villus height to crypt depth ratio than for the unsupplemented chicks (Shao *et al.*, 2013). Goblet cells in the intestinal mucosa secrete mucus which provides the first line of defense against intestinal injury. In *S. Typhimurium* challenged chicks, the number of goblet cells decreased. Supplementation with  $\beta$ -glucan, however, significantly increased the number of goblet cells in the jejunum of challenged chicks (Shao *et al.*, 2013). This suggests that the  $\beta$ -glucan is playing a role in improving gut health during a bacterial challenge.

All cells involved in the immune reactions originate from common precursors.  $\beta$ -glucan stimulates the production of precursor cells in bone marrow, resulting in an increase flow of new immunocytes into the various lymphoid organs throughout the body (Petračić-Tomenac *et al.*, 2010). This increases protection from potential invaders.

Macrophages are large white blood cells that phagocytize invading pathogens (Qureshi, 2003). In a typical immune response, macrophages recognize specific markers on the surface of the pathogens, referred to as Microbial Associated Marker Proteins (MAMPs). The MAMPs bind to the receptors on the phagocytic cells known as Toll-like Receptors (TLRs) (Medzhitov *et al.*, 1997). Birds have 10 known TLRs, although only five are orthologous to both birds and mammals (Temperley *et al.*, 2008). Key TLRs in chickens are TLR-2 that recognizes peptidoglycan, TLR-4 that binds LPS common in gram-negative bacteria, TLR-5 that recognizes flagellin common in bacteria flagella and TLR-21 which recognizes unmethylated CpG DNA commonly found in bacteria (Kestra *et al.*, 2010). The macrophage binds the MAMPs of the invading pathogens to the TLRs on their outer surface and brings the organism into their cytoplasm in the form of a phagosome. The phagosome then fuses with a lysosome and the enzymes in the lysosome degrade the engulfed organism. Macrophage function has been shown to be responsive to dietary  $\beta$ -glucans. Dietary supplementation with  $\beta$ -glucans from the yeast *Saccharomyces cerevisiae* has been shown to increase phagocytic activity in broiler chicks (Guo *et al.*, 2003). Lowry *et al.* (2005) reported that feeding day-old chicks

with purified  $\beta$ -glucan significantly decreased the levels of *Salmonella* Enteritidis (SE), a potential human pathogen, in the liver and spleen. The  $\beta$ -glucans appear to have functioned by increasing phagocytosis, bactericidal killing and oxidative burst from the heterophils. This was confirmed by Chen *et al.* (2008) who reported that supplementation with yeast  $\beta$ -glucan enhanced the chick's defense against SE by directly upregulating both the phagocytosis and bactericidal activity of abdominal macrophages. Dietary supplementation of  $\beta$ -1,3-1,6-glucan of the fungus *Schizophyllum commune* was also shown to increase the phagocytic and bactericidal capabilities of intestinal macrophages (Chen *et al.*, 2008; Muthusamy *et al.*, 2013).

Macrophages also produce the enzyme iNOS. This enzyme results in the production of nitric oxide which reacts with superoxide anions to generate toxic derivatives which allow the macrophage to kill several types of pathogens. Cox *et al.* (2010) reported upregulation of iNOS in response to  $\beta$ -glucan supplementation, implying increased capacity of macrophages to destroy pathogens.

Animals are able to recognize cells as being self from those that are not self. This is done through specific markers on the surface of the body cells referred to as Major Histocompatibility Complex (MHC). The TLRs are non-self-proteins on invading organisms and are referred to as antigens because they stimulate the production of antibodies. When a macrophage digests a pathogen it does not destroy the antigens. Instead, the antigens are processed and moved to the surface of the macrophage producing a MHC-antigen complex and creating an antigen-presenting macrophage which triggers the adaptive immune system.

As with mammals, the avian adaptive immune system has two major segments: Humoral and cellular immunity. Humoral immunity involves antibody production from B-lymphocytes which are associated with the Bursa of Fabricius, which is unique to birds. The principal cells involved in cellular immunity are T-lymphocytes which are associated with the thymus. After the foreign cell is degraded, the macrophage presents bacterial peptides or antigenic determinants to T- and B-cells (Qureshi, 2003; Genovese *et al.*, 2013).

There are different types of T-cells. The cytotoxic T-cell (Tc) binds to the MHC-antigen complex on the macrophage. The macrophage then secretes the chemical Interleukin-1 (IL-1). In response, the type-1 helper T-cell (Th1) binds to another MHC-antigen on the macrophage and releases Interleukin-2 (IL-2). IL-2 then activates the already bound Tc, causing it to go through several rounds of cell division to create a large population of Tc cells. The population of Tc cells then divides into two subpopulations of effector cells, which will go out and destroy infected body cells and memory cells, which are

held in reserve for future exposures to the pathogen. The effector cells will bind to infected body cells and secrete perforins which punch holes in the surface of the infected cell causing it to spill out its contents. The effector cell then moves on in search of other infected body cells. After the infection is over, the number of effector cells will decrease.

Macrophages also produce IL-1 $\beta$ , interferon (IFN)- $\gamma$ , IL-2, Tumor Necrosis Factor (TNF)- $\alpha$  and IL-4. The production of IL-4 results in the development of type-2 helper T Cells (Th2). Th2 cells then produce IL-5, IL-6, IL-10 and IL-13 which are involved in the humoral immune response. Macrophages can also produce IL-8 (sometimes referred to as CXCLi2), which is a chemokine that is an important mediator of the innate immune system. Its primary function is to induce the migration of its target cells to inflammation sites (Cox *et al.*, 2010). IL-18 is a pro-inflammatory cytokine largely produced by macrophages. After exposure to a pathogen, IL-18 works with IL-12 to induce a cell-mediated immune response. It primarily targets Th cells which will then secrete interferon- $\gamma$ . IFN- $\gamma$  plays an essential role in activating macrophages (Cox *et al.*, 2010).

$\beta$ -glucan supplementation has been shown to alter the cytokine profiles of unchallenged, day-old chicks (Cox *et al.*, 2010). Compared with unsupplemented controls, broiler chicks receiving  $\beta$ -glucan from *Saccharomyces cerevisiae* were shown to down-regulate IL-8. The down-regulation of the IL-8 gene with  $\beta$ -glucan supplementation suggests that the  $\beta$ -glucan functions as an anti-inflammatory immunomodulatory (Cox *et al.*, 2010).

Cox *et al.* (2010) were the first to report on the effects of dietary  $\beta$ -glucan on IL-18. Expression of the pro-inflammatory cytokine IL-18 was found to be up-regulated in the jejunum seven days after feeding  $\beta$ -glucan to day-old chicks. By day 14, however, there was a decrease in IL-18 in the duodenum. IFN- $\gamma$ , which plays a central role in promoting Th1 cell differentiation and activating macrophages, was reported to be down-regulated in the  $\beta$ -glucan supplemented chicks compared to those receiving the unsupplemented control diet. Supplementation with  $\beta$ -glucan for seven-days post-hatch up-regulated IL-4 expression in the duodenum, jejunum and ileum. By day 14, however, IL-4 expression was up-regulated in the duodenum and ileum (Cox *et al.*, 2010). The functions of IL-13 overlap with those of IL-4. Similar to what happened with the IL-4 expression on day seven, IL-13 gene expression was also down-regulated in all intestinal sections. Having both IL-4 and IL-13 being down-regulated would suggest that  $\beta$ -glucan supplementation does not support a Th2 cell-mediated response (Cox *et al.*, 2010).

It has been hypothesized that  $\beta$ -glucans link to the Major Histocompatibility Complex (MHC) and

subsequently activate monocytes and macrophages (Zhang *et al.*, 2008). This results in increased immune response including the activation of cytotoxic macrophages, T-helper and natural killer cells, as well as the promotion of T-cell proliferation and differentiation. IL-1, IL-2, IFN- $\gamma$  and TNF- $\alpha$  may act as the signal regulating this immune network.

B-cells are lymphocytes with bound antibodies on their surface. The antibodies will match specific antigens typically found on pathogens. When the B-cell with the antibody for a specific antigen encounters the pathogen with this antigen, the two become bound together, priming the B-cell. The IL-2 from the T-cell acts as a signal for the B-cell to divide to create a large population of B-cells. The B-cells divide into two subpopulations, effector and memory cells. The effector cells are involved in the production of antibodies while the memory cells remain in preparation for future infections. The antibodies will circulate in the blood and attach to the pathogen. It is difficult for the pathogen to infect other body cells when the antibodies are attached. The antibodies also mark the pathogens for destruction. The destruction can be carried out by macrophages, natural killer cells and heterophils. Natural Killer (NK) cells are a type of cytotoxic lymphocyte. They play a role similar to cytotoxic T-cells. Heterophils are granulocytic phagocytes and functionally equivalent to neutrophils in mammals. The cytoplasmic granules of heterophils contain lysozyme and proteins needed for bactericidal activity. Unlike mammalian neutrophils, however, heterophils do not have peroxylase activity. Inclusion of yeast  $\beta$ -glucan and a commercial probiotic product in the diet of chicks resulted in significantly higher antibody titers against Newcastle disease or infectious bronchitis virus as compared to unsupplemented control chicks (An *et al.*, 2008).

The intestinal tract is an important interface between the external environment and the internal body itself. Gut immunity, therefore, is important as the first line of defense from various pathogenic organisms. In the chicken, the Gut-Associated Lymphoid Tissues (GALT) include cecal tonsils and Peyer's patches, as well as lymphoid aggregates in the urodeum and proctodium of the cloaca. GALT structures are the sites of immune induction including the production of Immunoglobulin A (IgA) from B-cells. IgA is found in serum bile, mucosal surfaces, saliva and tears. They function in mucosal immunity. They neutralize virus and microbial toxins and prevent the adherence and colonization of microbial pathogens. As such, IgA is an important intestinal defense. A *S. Typhimurium* challenge increased the number of IgA-expressing cells as well as the level of IgA produced. These levels were even higher in the chickens receiving  $\beta$ -glucan supplemented diets (Shao *et al.*, 2013).  $\beta$ -Glucan supplementation also reduced the levels of *S.*

Thyphimurium infiltration as indicated by lower cecal and liver levels.

Much of the research with  $\beta$ -glucans and poultry has been with yeast products. Cheng *et al.* (2004) showed that  $\beta$ -1,3-glucan of the fungus *Schizophyllum commune* significantly increased the chemotaxis activity of macrophages, but had no effect on lymphocyte blastogenesis after exposure to different mitogens and no effect on antibody titer after a Newcastle Disease vaccination. The difference in results may be the level of  $\beta$ -glucan used since Cheng *et al.* (2004) used low levels of  $\beta$ -glucan inclusion.

A  $\beta$ -glucan from mushrooms has been shown to promote splenocyte proliferation and interleukin-2 production in broilers (Chen *et al.*, 2008). Purified  $\beta$ -glucan from the edible mushroom *Pleuratusflorida* was shown to provide broiler chicks with some protection from Newcastle disease (Paul *et al.*, 2012). IL-2 production was reported to have increased significantly.

Zhang *et al.* (2012) fed a commercial  $\beta$ -glucan powder to broilers. The  $\beta$ -glucan was derived from the bacterium *Agrobacterium* species. The powder was guaranteed to contain 86.1%  $\beta$ -1,3/1,6 glucan. While Zhang *et al.* (2012) did not observe any changes in blood profiles, feeding  $\beta$ -glucan significantly increased (3.9%) the relative weight of the spleen. In chickens the spleen serves as both a reservoir of leukocytes as well as their activation. Activation of spleen genes reflects systemic immune function (Redmond *et al.*, 2010). This would include genes responsible for interleukin (IL)-4 and IL-18. Different chicken genotypes have been shown to react differently to  $\beta$ -glucan supplementation (Redmond *et al.*, 2010). Gene expression of IL-4 and L-18 in a broiler line was not changed in response to a 3-week  $\beta$ -glucan supplementation, but did respond in chickens of the Fayoumi line. This would indicate that the effect of immunomodulatory supplementation may not have the same beneficial effects in all gene lines.

An additional impact of  $\beta$ -glucans on the immune system involves protection from mycotoxins. Mycotoxins such as aflatoxin B1 (AFB1) cause major health issues in poultry. Chronic AFB1 intake has been shown to result in poor health, a loss in productivity, as well as immune system dysfunction. This appears to be due to the damage to cell DNA (Zimmermann *et al.*, 2014).  $\beta$ -Glucans from the yeast *Saccharomyces cerevisiae* have been shown in vitro to be cytoprotective and genoprotective of chicken lymphocytes (Zimmermann *et al.*, 2015). The researchers speculated that the cytoprotection was a result of  $\beta$ -glucan assisting in repair or blocking of mutations in the life cycle of the cell. This could be due to  $\beta$ -glucan adsorbent capacity, interacting chemically with AFB1.

Le *et al.* (2011) reported that a commercial  $\beta$ -glucan was a potential effector, enhancing the immune response of the avian influenza H5 vaccines tested. The avian

influenza vaccines produced in oil-emulsified inactivated form, were shown to induce immunity in chickens, Muscovy ducks and Vietnamese ducks. They added a  $\beta$ -1,3-1,6 glucan produced by the black yeast *Aureobasidium pollulans* to the vaccine and assessed the ELISA and hemagglutination inhibition assays. Animals with the  $\beta$ -glucan enhanced vaccine had titers significantly higher than those vaccinated with the unsupplemented vaccine.

## Conclusion

There are different types of  $\beta$ -glucans. Glucans from cereals such as barley and oats, have very detrimental effects on poultry health and performance. The inclusion of some of the varieties of barley and oats in poultry diets is made possible by supplementation with feed enzymes which digest the  $\beta$ -glucan. Other  $\beta$ -glucans, such as those from fungi and yeast, have been shown to be beneficial as dietary supplements. They have been shown to increase immune responses in poultry.  $\beta$ -Glucan supplementation was shown to increase the production of new immunocytes. Supplementation has also been shown to increase the phagocytic function of macrophages.  $\beta$ -Glucans increase the phagocytic, bactericidal killing and oxidative burst of heterophils.  $\beta$ -Glucan has also been shown to enhance immune response by altering the cytokine profiles of chickens.

$\beta$ -Glucans, therefore, may provide a tool for producers trying to reduce or eliminate the use of antibiotics in poultry diets. Supplementation with  $\beta$ -glucans has also been shown to be beneficial in protection from mycotoxins. In addition, inclusion of  $\beta$ -glucans in vaccines, may enhance the immune response they elicit.

## Author's Contributions

Both authors contributed to the writing of this paper.

## Ethics

No ethical issues are expected to arise from the publication of this manuscript.

## References

- An, B.K., B.L. Cho, S.J. You, H.D. Paik and H.I. Chang *et al.*, 2008. Growth performance and antibody response of broiler chicks fed yeast derived  $\beta$ -glucan and single-strain probiotics. Asian-Aust. J. Anim. Sci., 21: 1027-1032. DOI: 10.5713/ajas.2008.70571
- Chen, K.L., B.C. Weng, M.T. Chang, Y.H. Liao and T.T. Chen *et al.*, 2008. Direct enhancement of the phagocytic and bactericidal capability of abdominal macrophage of chicks by  $\beta$ -1,3-1,6-glucan. Poul. Sci., 87: 2242-2249. DOI: 10.3382/ps.2008-00147

- Cheng, Y.H., D.N. Lee, C.M. Wen and C.F. Weng, 2004. Effects of  $\beta$ -glucan supplementation on lymphocyte proliferation, macrophage chemotaxis and specific immune responses in broilers. *Asian Aust. J. Anim. Sci.*, 17: 1145-1149. DOI: 10.5713/ajas.2004.1145
- Cox, C.M., L.H. Stuard, S. Kim, A.P. McElroy and M.R. Bedford *et al.*, 2010. Performance and immune responses to dietary  $\beta$ -glucan in broiler chicks. *Poult. Sci.*, 89: 1924-1933. DOI: 10.3382/ps.2010-00865
- Genovese, K.J., H. He, C.L. Swaggerty and M.H. Kogut, 2013. The avian heterophil. *Dev. Comparative Immunol.* 41: 334-340. DOI: 10.1016/j.dci.2013.03.021
- Guo, Y., R.A. Ali and M.A. Qureshi, 2003. The influence of  $\beta$ -glucan on immune responses in broiler chicks. *Immunopharmacol. Immunotoxicol.*, 3: 461-472. DOI: 10.1081/IPH-120024513
- Keestra, A.M., M.R. de Zoete, L.I. Bouwman and J.P. van Putten, 2010. Chicken TLR21 is an innate CpG DNA receptor distinct from mammalian TLR9. *J. Immunol.*, 185: 460-467. DOI: 10.4049/jimmunol.0901921
- Le, T.H., T.B. Le, T.H.T. Doan, D.V. Quyen and K.X.T. Le *et al.*, 2011. The adjuvant effect of Sophy  $\beta$ -glucan to the antibody response in poultry immunized by the avian influenza A H5N1 and H5N2 vaccines. *M. Microbiol. Biotechnol.*, 21: 405-411. PMID: 21532325
- Lowry, V.K., M.B. Farnell, P.J. Ferro, C.L. Swaggerty and A. Bahl *et al.*, 2005. Purified  $\beta$ -glucan as an abiotic feed additive up-regulates the innate immune response in immature chickens against *Salmonella enterica* serovar *Enteritidis*. *Int. J. Food Microbiol.*, 98: 309-318. DOI: 10.1016/j.ijfoodmicro.2004.06.008
- Medzhitov, R., P. Preston-Hurlburt and C.A. Janeway Jr., 1997. A human homologue of the *Drosophila* Toll protein signals activation of adaptive immunity. *Nature*, 388: 394-397. DOI: 10.1038/41131
- Muthusamy, G., S.N. Joardar, I. Samanta, D.P. Isore and B. Roy *et al.*, 2013.  $\beta$ -Glucan from edible mushroom (*Pleurotus florida*) enhances mucosal immunity in poultry. *Adv. Anim. Vet. Sci.*, 1: 116-119.
- Paul, I., D.P. Isore, S.N. Joardar, I. Samanta and U. Biswas *et al.*, 2012. Orally administered  $\beta$ -glucan of edible mushroom (*Pleurotus florida*) origin upregulates innate immune response in broilers. *Indian J. Anim. Sci.*, 82:745-748.
- Petravić-Tomenac, V., V. Zechner-Krpan, S. Grba, S. Screčec and I. Panjkota-Krbavčić *et al.*, 2010. Biological effects of yeast  $\beta$ -glucan. *Agric. Conspectus Scientificus*, 75: 149-158.
- Qureshi, M.A., 2003. Avian macrophage and immune response: An overview. *Poult. Sci.*, 82: 691-698. DOI: 10.1093/ps/82.5.691
- Rathgeber, B.M., K.L. Budgell, J.L. MacIsaac, M.A. Mirza and K.L. Doncaster, 2008. Growth performance and spleen and bursa weight of broilers fed yeast beta-glucan. *Can. J. Anim. Sci.*, 88: 469-473. DOI: 10.4141/CJAS07101
- Redmond, S.B., R.M. Tell, D. Coble, C. Mueller and D. Palić *et al.*, 2010. Differential splenic cytokine responses to dietary immune modulation by diverse chicken lines. *Poult. Sci.*, 89: 1635-1641. DOI: 10.3382/ps.2010-00846
- Shao, Y., Y. Guo and Z. Wang, 2013.  $\beta$ -1,3/1,6-Glucan alleviated intestinal mucosal barrier impairment of broiler chickens challenged with *Salmonella enterica* serovar Typhimurium. *Poult. Sci.*, 92: 1764-1773. DOI: 10.3382/ps.2013-03029
- Temperley, N.D., S. Berlin, I.R. Paton, D.K. Griffin and D.W. Burt, 2008. Evolution of the chicken toll-like receptor gene family: A story of gene gain and gene loss. *BMC Genom.*, 9: 62-74. DOI: 10.1186/1471-2164-9-62
- Zhang, B.Z., Y. Guo and Z. Wang, 2008. The modulating effect of  $\beta$ -1,3/1,6-glucan supplementation in the diet on performance and immunological responses of broiler chickens. *Asian-Aust. J. Anim. Sci.*, 21: 237-244. DOI: 10.5713/ajas.2008.70207
- Zhang, Z.F., T.X. Zhou, X. Ao and I.H. Kim, 2012. Effects of  $\beta$ -glucan and *Bacillus subtilis* on growth performance, blood profiles, relative organ weight and meat quality in broilers fed maize-soybean meal based diets. *Livestock Sci.*, 150: 419-424. DOI: 10.1016/j.livsci.2012.10.003
- Zimmermann, C.E.P., A.K. Machado, F.C. Cadoná, J.A.S. Jaques and K.B. Schlemmer *et al.*, 2014. *In-vitro* cytotoxicity of aflatoxin B1 to broiler lymphocytes of broiler chickens. *Braz. J. Poultry Sci.*, 16: 307-312. DOI: 10.1590/1516-635x1603307-312
- Zimmermann, C.E.P., I.B.M. Cruz, F.C. Cadoná, A.K. Machado and C. Assmann *et al.*, 2015. Cytoprotective and genoprotective effects of  $\beta$ -glucans against aflatoxin B<sub>1</sub>-induced DNA damage in broiler chicken lymphocytes. *Toxicol. Vitro*, 29: 538-543. DOI: 10.1016/j.tiv.2015.01.005