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Review

Role of LAB in silage fermentation: Effect on nutritional quality and organic acid production—An overview

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Abstract: Lactic acid bacteria (LAB) inocula play a key role in the preservation and fermentation of forage crops within inoculated silages. LAB is a significant group of the bacterial community as they successfully reduce pH, inhibit the survival of undesirable microorganisms and control nutrient loss in fermented silage. Ensiled plants and metabolites such as simple plant carbohydrates have been utilized by LAB (homo-fermentative and hetero-fermentative LAB) to initiate the production of organic acids including lactic and acetic acids. LAB as a biological silage additive provides stable feed value and secondary metabolic products during rapid anaerobic primary silage fermentation. They are able to ferment a large number of forage crops and also to reduce pH levels in fermented forages, which helps to suppress the growth of spoilage microorganisms. Furthermore, silage inoculants can enhance silage quality, nutritional recovery and shelf life of the inoculated product. When ingested silage, Lactobacilli in the rumen may degrade secondary plant metabolites as part of the rumen microbiota, along with endogenous enzymes. Also, the forages harvesting time are key factors in the development of essential metabolites particularly carbohydrates and proteins which is essential nutrition for LAB survival and production of organic acids. The higher population of LAB could reduce the pH faster and control of deleterious microbial growth in silage. This review presents LAB function in silage production and the potential impacts of its

fermentative activity. In addition, the advantage of LAB additives in silage production is discussed, with a focus on recent literature.

Keywords: LAB additives; silage; fermentation; organic acids; metabolite degradation

1. Introduction

Recent research on lactic acid bacteria (LAB) has enhanced our understanding of their many advantageous properties; they can be used as biopreservatives, feed grade enzymes, veterinary medicines, health care products and, especially, food and beverage additives [1–4]. *Lactobacilli* are a fermentative, facultative anaerobe that is one of the first-evolving groups of bacteria to have beneficial effects for both humans and animals. LAB have been used as bio-additives in ensiling forage to improve fermentation and preserve nutritional content. Silage results from fermentation of a green chopped forage crop and storage of the product with removal of O₂ in a silo (ensiling). Ensiled forages contribute 40–60% of all nutrients for livestock animals. The process involves a wide range of factors, including plant cultivation, harvesting and storage practices [5,6]. *Lactobacilli* play a critical role in preservation of silage nutrient quality through production of organic acids and inhibition of deleterious spoilage from the remaining microbial community. Also, LAB effectively controls secondary fermentation within inoculated forages [7]. In silage, secondary fermentation is an undesirable acidification that is carried out mainly by *Enterobacteria*, *Clostridia* (butyric acid producer), and yeasts (ethanol producer).

LAB utilize water-soluble carbohydrates (WSC) and convert them into mixtures of organic acids. The use of LAB in starter cultures can enhance feed quality while minimizing the loss of nutrients (such as carbohydrates, crude proteins, volatile free fatty acids and minerals). The use of selected LAB additives enhances the fermentation process, leading to control of dry matter (DM) loss and pathogenic activity [8]. In silage production, homofermentative LAB are used most broadly due to the high amounts of lactic acid production that occur during fermentation (by e.g: *Lactobacillus*, *Pediococcus*, *Streptococcus spp.*, and *Enterococcus*). Currently, many researchers are using heterofermentative LAB (e.g: *Leuconostoc spp.* and some *Lactobacillus* strains) as silage additives with a view towards production of acetic acid and butanol for fuel production [9–11].

Silage microflora can be categorized into two main groups, desirable and undesirable organisms. LAB are desirable microbes, while undesirable microorganisms (*Enterococcus*, yeast and molds) can cause anaerobic or aerobic spoilage during silage fermentation. These undesirable epiphytic microorganisms can decrease the nutritional quality of the silage and also affect animal health and/or milk production [12,13]. Silage additives can be classified into six main groups: homofermentative LAB (hoLAB), heterofermentative LAB (heLAB), customized inoculants (such as hoLAB + heLAB), chemicals, and enzymes. The hoLAB quickly decrease pH and increase lactic acid production in corn, sorghum, and sugarcane silages [14]. Also, the LAB suppresses the growth of undesirable microorganisms and thus reduce proteolysis and DM loss in early fermentation [15]. Blajman et al. [16] reported that LAB inoculation reduces undesirable yeast and mold growth in treated silage, and improves aerobic stability and LAB count in corn silage. However, the effect is highly dependent on the types of inoculants (hoLAB or heLAB) used during silage fermentation. Also, Oliveira et al. [17]

explored meta- analysis using silage, LAB species, and silo scale (laboratory or farm-scale) as important factors that determine silage quality.

In recent decades, silage research has driven novel innovations by private companies, which have experimented with silage inoculums, forage harvesters and plastic films for making in silos [5,18]. In addition, recent silage research has addressed the different parameters of the process, including biochemical, microbiological, agronomical and nutritional aspects. The purpose of silage production is to preserve the nutritional value and original energy content of the forage. Energy loss may occur as a result of many factors, such as field loss, harvesting-related loss, secondary metabolism during fermentation of ensilage material, aerobic spoilage and oxidation when the silage is unwrapped to serve to ruminants [19]. Aerobic spoilage is mainly associated with penetration of O₂ into the silage during the storage or feeding period. The nutritional composition and feed value of the silage is dependent on the effectiveness of the silage inoculums in the fermentation process. Proper production and management of silage is very important since contaminated or poorly prepared silage can host undesirable organisms, such as *Listeria monocytogenes*, *Bacillus spp.*, *Clostridium spp.*, *Salmonella*, *Lactococcus* and molds [5]. Growth of pathogenic organisms in silage may reduce the safety and quality of the feed and decrease dairy cattle performance, in particular milk and meat production [20,21]. Some concepts necessary to consider when inoculating a silage ball/pack with LAB are exclusion of air, availability of adequate water-soluble carbohydrates, suitable moisture content and the initiation of an early and prompt fermentation [22]. Under appropriate conditions, silage fermentation should preserve over 90% of the harvested energy, sugar, crude protein and metabolites over long storage periods [5]. In this review, we summarize recent information about the functions of LAB strains in silage fermentation as well as their primary fermented products and methods of improving the quality of silage and its shelf life.

2. Biological properties of LAB

LAB is gram-positive, microaerophilic, non-spore forming, cocci- or rod-shaped bacteria. They consist of different genera, such as *Lactobacillus*, *Leuconostoc*, *Pediococcus*, *Streptococcus*, *Enterococcus* and *Lactococcus* [23]. Classification of LAB into different genera is largely dependent on morphology, action of fermentable carbohydrates, growth at different temperatures, ability to survive under acid or alkaline conditions and production of various organic acids. Though the majority of silage LAB can grow under mesophilic conditions, including different temperatures (20–50 °C), the optimum temperature for growth is between 25 and 40 °C. LAB strains are naturally present in various resources, including fermented foods (yogurt, kimchi, meat), forage crops (Italian rye grass, alfalfa, rye, sorghum, corn and triticale), animal manure, dairy products, rumen juice and infant feces [24–26]. Some LAB are facultative aerobes, but some have a preference for anaerobic conditions. LAB are able to reduce silage pH to between 3–5 depending on the selected strains and the type of forage crop [27]. The main function of LAB is to ferment WSC to organic substances as their major end product. LAB generally belong to the genera *Lactobacillus* and *Bifidobacterium*, which are most widely recognized and used as probiotics in feed development, including dairy foods and beverages. Also, the consumption of probiotics is associated with a wide range of health benefits in humans and livestock animals [28].

3. LAB as silage additives in livestock feed

In order to improve the quality of silage fermentation, different silage additives have been developed. Recently, biological additives (inoculants) are becoming more widely used as silage preservatives; these can be added to forage in order to increase organic acid production and rapidly decrease silage pH from 6.5 to 3.5. The most important inoculants consist of preferred strains of hoLAB, for example, *Lactobacillus* spp. (*L. plantarum*, *L. acidophilus*, *L. lactis*, *L. bulgaricus*), *Enterococcus faecium* and *Pediococcus* spp. (*P. pentosaceus*, *P. acidilactici*, *P. cellicola*); these can produce precise quantities of lactic acid in a short fermentation period and so stabilize the silage with minimal nutritional and DM losses [29,30]. Various factors reduce DM and quality loss during the ensiling steps, including the field and pre-ensiling conditions, temperature, fermentation patterns, moisture content during ensiling, methods and materials used to pack the silage and aerobic deterioration during the feed out phases [31].

In recent years, customized bacterial inoculants have been introduced to enhance organic acid production in silage [32,33]. The primary purpose of these customized inoculants, which include hoLAB strains, is to encourage early fermentation in order to increase the efficiency of fermentation and quickly reduce silage pH. Driehuis et al. [34] LAB inoculated perennial ryegrass with three treatments, *L. buchneri* alone; *L. buchneri* with *L. plantarum*, and *P. pentosaceus* and non-inoculated silage served as control. The combination inoculants exhibited similar fermentation to the homofermentative *L. plantarum* and *P. pentosaceus* treatment over the first 14 days. However, the combination of inoculants and *L. buchneri* alone resulted in reduced yeast colony counts and improved aerobic stability of treated silage compared with the untreated control during a 90-day fermentation period. Aerobic stability is defined as the length of storage time the silage remains as well-preserved dry matter content with the lowest amount of yeast and mold spores, even after it is exposed to air. Reports of the use of combination inoculants in various types of silage have been published, with successful outcomes [35].

The production of organic acids by LAB depends on the amount of WSC present in the fresh crop and the precise nature of the bacterial strains used [30,36]. The carbohydrates in the fresh crop are catabolized to produce organic acids and other substances. Table 1 illustrates the effect of LAB inoculation on organic acid production in silage fermentation. Different types of fermentation may take place in the silo environment, mainly depending on the concentration of WSC, volatile free fatty acids, crude proteins, minerals, and moisture in the forage crop and the microbial community present during the ensiling period [17,31].

Table 1. Organic acid production by LAB during silage fermentation.

Species	Type of silages	Fermentation products	Functions	Cited
<i>Lactobacillus plantarum</i> , <i>Lactococcus lactis</i> , and <i>Lactobacillus buchneri</i>	<i>Lolium perenne</i>	Lactic acid and other organic acids	Productivity of Holstein Friesian dairy cattle	[37]
<i>Lactobacillus plantarum</i> and <i>Pediococcus pentosaceus</i>	Soybean	Enhance the lactic acid, and crude protein content	Inhibit the growth of undesirable microbes	[38]
<i>Lactobacillus plantarum</i>	Rice straw	Improve lactic acid/ acetic acid ratios	Hemicellulose degradation and inhibition of clostridia fermentation	[39]
<i>Lactobacillus plantarum</i> , <i>Lactobacillus buchneri</i> , and <i>Lactobacillus rhamnosus</i>	Maize and Amaranthus	Production of lactic acid and a specific methane yield	Organic juice into biogas production	[40]
<i>Lactobacillus plantarum</i>	Alfalfa	Enhance LA content and LA/AA	Non-irradiated or irradiated alfalfa forage	[41]
<i>Lactobacillus buchneri</i>	Sugarcane	Improve the WSC, lactic acid, acetic acid and ethanol concentration	Improvement on aerobic stability	[39]
<i>L. plantarum</i>	Rice straw silage	Enhance its fermentation quality, nutritive characteristics and <i>in vitro</i> digestibility	<i>In vitro</i> gas production of rice straw silage	[32]
<i>Bacillus megaterium</i> , <i>Pediococcus acidilactici</i> and <i>Lactobacillus plantarum</i>	<i>Medicago sativa</i>	Improve the of γ - aminobutyric acid, and 4-hydroxy benzoic acid and phenyl lactic acid production.	Regulating the organic acid and toxin contents	[42]
<i>Lactobacillus amylovorus</i> , <i>Lactobacillus mucosae</i>	Cattle feed	Improve the lactic acid production	Inhibition of <i>Salmonella</i> spp.	[43]
<i>L. parafarraginis</i> , <i>Lactobacillus brevis</i>	Corn stover	Improve the lactic acid and acetic acid contents	Improving the corn stover silage quality	[44]
<i>L. buchneri</i> , <i>Leuconostoc mesenteroides</i>	Corn silage	Production of organic acids	Control the <i>E.coli</i> growth	[45]

3.1. Carbohydrate metabolism in silage production

Silage microflora ferment carbohydrates to obtain energy for growth and survival and produce organic acids and other metabolites as end products. In general, silage fermentation utilizes hexoses as the primary substrate. Hexoses are degraded by a different class of silage bacteria and produce organic acids as metabolic end products [46–48]. The concentrations of these carbohydrates may vary depending on the type of forage, the time of harvest and environmental conditions [49]. The main pathway used by LAB is glycolysis (Embden-Meyerhoff), in which glucose-containing substrates are catabolized to pyruvate, and acetyl coA is formed as an intermediate product of the Krebs cycle [50,51]. During the ensiling process, both homofermentative and heterofermentative LAB partially hydrolyze hemicellulose to yield pentoses, such as xylose and arabinose, which are fermented into organic acids by the phosphoketolase pathway [52].

There are three main metabolic pathways found in LAB, the homofermentation, heterofermentation and bifidum pathways. Homofermentation converts glucose to lactic acid at a rate that exceeds 80% of the theoretical efficiency. The most abundant organic acid produced by homofermentation is lactic acid. Many LAB strains, such as *Lactobacillus*, *Lactococcus* and some *Streptococci*, carry out homolactic acid fermentation. Very small quantities of formic acid, acetic acid, propionic acid and tartaric acid are also produced as fermentation products (Figure 1). The concentration of other organic acids formed is lower, and negligible for our purposes [53]. LAB uses the EMP (glycolysis) pathway to oxidize one glucose molecule into 2 pyruvate molecules, along with 2 ATP and 2 NADH. Both pyruvates are reduced to lactate by the oxidation of NADH to NAD⁺ [54,55]. Some LAB strains (e.g: *Lactobacillus* and *Bifidobacterium*) utilize the heterolactic acid pathway, in which a phosphoketolase enzyme breaks down a pentose sugar molecule into a 3-carbon phosphate and a 2-carbon phosphate molecule [56–58]. As a heterolactic acid fermentation pathway, is mainly initiated by the heLAB, which include *Lactobacillus* and *Leuconostoc* sp. The hetero lactic fermentation major end product is ethanol and CO₂ in addition to lactic acid. The precursor glucose molecules are first metabolized to pyruvate, acetic acid and CO₂ by Pentose phosphate pathway. In addition, heLAB and yeasts use the EMP pathway to synthesize ethanol and CO₂, and also to produce diverse gases, fatty acids and alcohols. In LAB, the primary form of reduced coenzymes nicotinamide adenine dinucleotide (NADH) and nicotinamide adenine dinucleotide phosphate (NADPH) greatly affect decomposition of carbon sources and trends of metabolic synthesis, respectively. NADPH is required to provide the reducing power and to promote synthesize cellular components such as amino acids, nucleotides and lipids, among others, which have glucose intermediates, thereby providing the raw materials for further production of lactic acid [54].

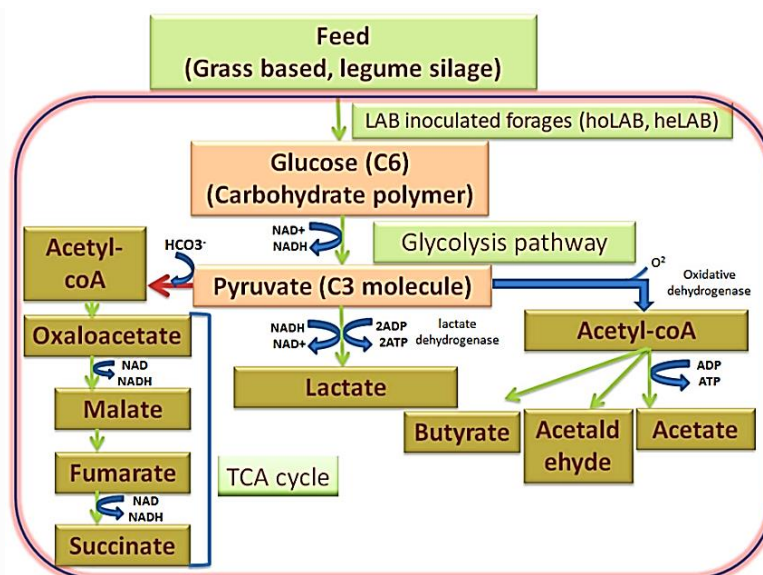


Figure 1. Schematic representation of organic acid metabolic pathways during silage fermentation [54] (C6: 6-carbon molecule; ATP: adenosine triphosphate; ADP: adenosine diphosphate; NAD⁺: Nicotinamide adenine dinucleotide; NADH: Nicotinamide adenine dinucleotide (reduced form); HCO₃: bicarbonate; CO₂: Carbon dioxide).

3.2. Organic acid production using LAB in silage

Normally, ensiled forage crops contain different organic acids, with the quantity and variety of these acids varying depending on LAB species [22,59]. Jones and Barnes [60] compared several forage varieties and found mostly organic acids, such as malic, citric, and succinic acids, with minor concentrations of fumaric and shikimic acids. Figure 1 schematically illustrates the metabolism of organic acids by LAB during silage fermentation. On the other hand, undesirable microbial fermentation in the silo can produce various end products that alter the nutritive value of the forage. LAB produce high-quality silage and control the formation of undesirable compounds from secondary fermentation which can negatively affect animal performance, the environment, and net farm income [61].

4. Biological activity of *Lactobacillus* in silage production

The scientific community has recently been focused on finding novel drugs to inhibit the colonization of multidrug-resistant pathogenic organisms. Some LAB strains can be used as probiotics for ruminants, not only because of the production of lactic acid, but also because they synthesize bacteriocins, bacteriocin-like substances (BLS) and exo-polysaccharides, all of which improve the health of livestock [62–64]. Generally, LAB produce different types of antimicrobial compounds that they are safe substances for animal and human. These metabolites which are synthesized from bacterial ribosomes and inactivated by digestive proteases [65,66]. The most common pathogenic microorganisms found in silage are *E. coli*, *Salmonella* spp., *L. monocytogenes*, *Clostridium* spp. and *Bacillus* spp. [20]. LAB inoculation of silage rapidly reduces the pH to below 4

and has also been shown to prevent *E. coli* multiplication in fermented silage [67]. Similarly, Pedroso et al. [68] reported the effectiveness of three combined LAB commercial inoculants at controlling *E. coli* O157: H7 in corn silages. LAB silage inoculation suppressed pathogen activity within 3 d of ensiling, when the pH dropped below 4.0. The growth inhibition of pathogens in silage was considered to be due to lower pH, and presence of antimicrobial compounds such as organic acids, bacteriocin and BLS in the preserved silages. These metabolites may eliminate the activity of unwanted food-borne pathogens [69]. Similarly, the production of microbial secondary metabolites at lower pH level may also directly reduce the growth of pathogens. Furthermore, pure cultures of *L. buchneri* and *L. plantarum* have shown pH-independent antibacterial activity against *E. coli* O157: H7 [70]. All of these biological additives should be uniformly distributed in the silage for maximum efficiency. It is apparent that to inhibit aerobic spoilage, one must inhibit the growth and activity of microorganisms that cause silage deterioration (i.e. *Clostridia*, *Enterobacteria*, molds and yeasts). In addition, the bacteriocin did not affect the other bacterial communities involved in silage fermentation [71]. These were *L. lactis* CECT 539 and *P. acidilactici* NRRL B-5627, both of which are bacteriocinogenic strains that have shown antilisterial activity *in vitro*, and effectiveness for controlling *L. monocytogenes* in the silo [72]. Figure 2 shows the livestock applications of LAB inoculants in silage production. Gavrilova et al. [73] reported that new LAB strains isolated from clover silage were able to control biofilm formation and inhibit bacterial growth originating from biofilm. In addition, these strains exhibit high organic acid production that completely suppresses the growth of *E. coli* and *S. aureus*.

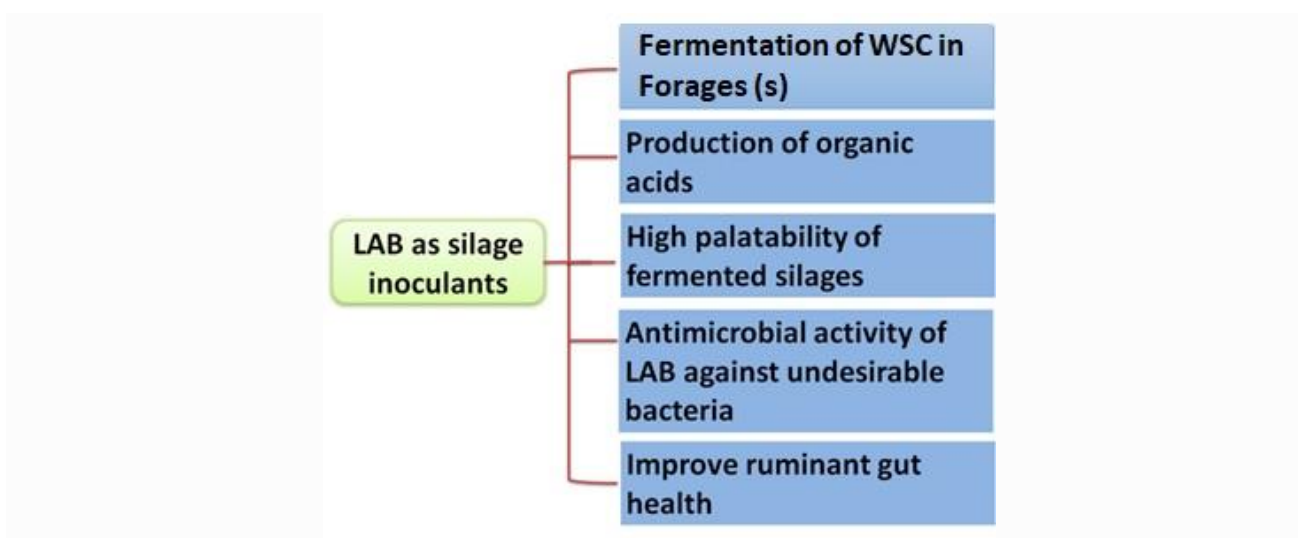


Figure 2. Biological activity of LAB in silage production and development.

Silage additives contain specific LAB strains, such as *L. plantarum*, *P. acidilacti*, *P. pentosaceus*, and *E. faecium*, which enhance homolactic fermentation by inhibiting undesirable bacteria or dominating the normal flora and increasing the rate of acidification in preserved forage. The antibacterial effect of LAB cell-free supernatant (CFS) against pathogens increased with lower pH levels, and no antibacterial activity was identified at pH 6.0. The CFS of the LAB strains showed moderate antibacterial effects against pathogenic bacteria, except *S. aureus* and *L. monocytogenes*, at pH below 5. [74]. Previously, Ogunade et al. [75] demonstrated that *E. coli* O157: H7 growth was

suppressed in silage extract when the pH was below 4. The low pH in silage was achieved due to rapid production of organic acids by the inoculants. When the pH of the silage falls below 4, lactic acid and acetic acid are primarily in their undissociated form. The cell membranes of yeasts and filamentous fungi are more permeable to the acids in this form. The acid is dissociated (carboxyl cation and hydrogen ion) within the cell, due to the high pH (>6.0), releasing H⁺ ions, which reduce proliferation and survival of pathogenic microorganisms in the silage [76,77].

When ryegrass was inoculated with *L. plantarum*, alone and in combination with either *L. lactis* or *P. acidilactici*, the DNA band for the spoilage microorganism disappeared after the 5th day of silage fermentation, indicating a synergistic effect of bacteriocin-producing bacteria. This synergistic effect can be attributed to the low pH achieved (<4.0) within 2 d of fermentation when the combination strains were used in the forage [19]. Low pH values stimulate the release of nisin and pediocin from the cell surfaces of *L. lactis* and *P. acidilactici*, respectively, and both bacteriocin metabolites exhibit strong antibacterial activity against spoilage bacteria [78]. Amado et al. [72] evaluated the effect of pediocin alone or in combination with a mixture of *L. plantarum*, *E. faecium*, and *L. buchneri* strains against *L. monocytogenes*. The LAB strains effectively suppressed growth of *L. monocytogenes* in corn silage via production of pediocin metabolites. Broberg et al. [79] reported that 3-hydroxydecanoic acid (100 µg/mL) inhibited *P. roqueforti*, *Pichia anomala* and *Aspergillus fumigatus*. Also, antifungal activity was seen at pH 4 in the presence of lactic acid (100 mM), 3-hydroxydecanoic acid, 2-hydroxy-4-methylpentanoic acid, benzoic acid and catechol in LAB-inoculated grass silage.

5. Effect of environmental factors on LAB forage ensiling

Although some forages are deficient in essential dietary nutrients for ruminants, a wide range of forages crops has been cultivated for livestock animal feeds. The nutritional value of these crops, including crude protein, VFA (volatile free fatty acids), ADF (Acid detergent fiber), NDF (neutral detergent fiber), and DM, can be enhanced by addition of LAB additives at the time of ensiling. Furthermore, biological additives have beneficial effects on silage fermentation due to improvements in the aerobic stability of silage and via control of the growth of pathogenic bacteria [20]. Some crops, like corn and sorghum, have better nutritional value to begin with and good ensiling characteristics. However, high-quality silage production often requires extensive processing, involving harvesting, crop wilting, selection of additives and ensiling steps, which can make production more expensive [80,81]. In general, hoLAB strains are preferable to the heLAB for preserving. However, some heLAB strains are also quite efficient because of their weaker acids production and possibly their higher pKa in heLAB contribute to aerobic stability effect of silage. [82]. Figure 3 shows the various factors which influence silage production. Importantly, Tanizawa et al. [52] clarified silage fermentation mechanisms from a genomic point of view, especially in cold conditions. Several genes were inactive in the cold condition, but some of the *L. vaccinostercus* strains, clearly demonstrating the ability to adapt to specific ecological niches. Also, Doi et al. [83] reported that LABs grown at room temperature have a significant capacity to improve the quality of silage. On the other hand, it is not clear whether strains showing strong lactic acid production can dominate butyric acid bacteria or coliform bacteria at higher temperatures (>37 °C). Different processes have been established to improve the fermentation quality and reduce dry matter losses of whole-crop silages of paddy rice, legumes and grasses [84,85]. Overall, biological silage additive-induced hoLAB

fermentation is more suitable to crop silage than other types of fermentation because it leads to greater dry matter recovery and improved fermentation end products [86]. LAB inoculated forages did not alter biochemical and nutritional components and it could preserve its quality of silage in terms of crude protein (CP) content, ADF (acid detergent fiber), NDF (neutral detergent fiber), and TDF (total detergent fiber). In addition, the nutrient values are nothing but the quality of the silage such as organic acids, crude protein content, ADF (acid detergent fiber), NDF (neutral detergent fiber), and TDF (total detergent fiber). Although, the organic acids levels and pH can reflect quality of fermentation and silage mass stability. The low pH shows the silage is stable and cannot further develop undesirable microbes too in the in-silo.

6. Advantage of LAB use in livestock industries

The diversity and function of LAB microbes present in a wide range of fermented foods that improve the nutritional value of fermented foods is now well-appreciated [87]. The fermented functional foods have presence of live microorganisms including different bioactive molecules, vitamins other constituents during the fermentation that enhanced safety, functionality, sensory, and nutritional properties of silage product [88]. Table 2 shows the merits of LAB inoculants in silage production. Also, the lactic acid bacteria ensiling benefits and uses of chemical additives were discussed.

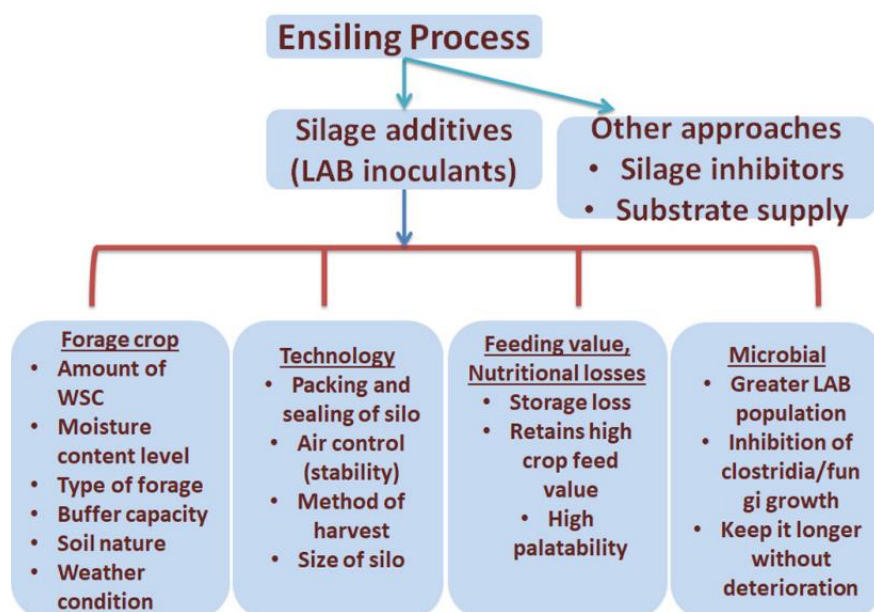


Figure 3. Different factors influencing silage metabolism.

Table 2. Comparison of LAB and Chemical additives in silage fermentation.

Particulars	LAB additives	Chemical additives
Fermentation efficiency	Addition of homofermentative LAB speed up fermentative process and inhibit the growth of pathogenic microbes via production of organic acid	Formic acid, mineral acid, sulphuric acid could also be effective silage additives to control pathogenic microbes and enhance the fermentation process. However they are hazardous and can damage ensiling materials like equipment and polyethylene wrappers etc. Also, undesirable odors.
Economic value	Effective on a wide range of forage crops and economic	Expensive and inefficient on some crop forages
Aerobic stability	Enhances aerobic stability and controls DM loss, on the other hand enhances the nutritive value	Though aerobic stability is high it can sometimes damage the silage cover
Dry matter content	Elimination of fermentation losses is not possible, but the use of silage additives may help minimize them	Fermentation dry matter loss high
Different environment condition	LAB is more tolerant to low moisture conditions (low water activity) than another undesirable anaerobic microorganisms	Low moisture fermentation is less active than other additives

Ruminal microbiota plays a vital role in the conversion of lignocellulose-rich plants into nutrients for ruminants. It has been proposed that different genera, such as bacteria, fungi, and protozoa, hydrolyze complex flavonoid glycosides to the corresponding less polar aglycones prior to gastrointestinal absorption [89]. There is a direct relationship between rumen pH and bacterial community, with significant reductions in bacteria at pH values around 6.8. Table 3 shows the degradation of different plant secondary metabolites by LAB in rumen fluid. In LAB-inoculated potato with wheat straw silage, the chemical composition, silage fermentation characteristics, and *in vitro* gas production were improved after anaerobic storage for 90 d, compared to untreated silage [90]. Contreras-Govea et al. [7] found that, in ruminal *in vitro* fermentation, gas and VFA production were not significantly different between non-inoculated and inoculated silage, although the *L. plantarum* MTD/1-inoculated model produced greater microbial populations than were seen in non-inoculated silage.

Table 3. Degradation of secondary plant metabolites by LAB in rumen.

Name of the LAB	Silage used	Method evaluated	Rumen metabolite degradation	Cited
<i>Lactobacillus rhamnosus</i> GG (ATCC 53013)	Highly contaminated feed	Male Holstein calves	Reduced the release of toxins into plasma	[91]
LAB or a combination.	Potato-wheat straw mixture	<i>In-vivo</i> study	Enhanced VFA production	[90]
<i>Lactobacillus buchneri</i>				
<i>Lactobacillus plantarum</i>				
<i>Propionibacterium acidipropionici</i>				
<i>E. faecalis</i> and ruminal microbiota	<i>Acacia nilotica</i>	<i>In-vitro</i> rumen liquor used for analysis	Improved degradation	tannin [92]
Rumen microbiota	<i>Eucalyptus globulus</i>	Sheep rumen liquor	Enhances breakdown	rutin [93]
Ruminal bacteria	<i>Cinnamomum zeylanicum</i> , <i>Eugenia caryophyllata</i>	<i>In-vitro</i> rumen inoculum	Improved oil degradation	essential [94]
Complex lactic acid bacteria	Corn crops	Crossbred bulls	Improved daily matter intake	dry [32]
Lactic acid bacteria	Corn stover silage	<i>In vitro</i> fermentation	<i>In vitro</i> digestibility improved	[95]

Those authors suggested that enhanced protein preservation during ensiling with a silage inoculant could be one of the reasons behind disturbances in ruminal fermentation. Zhang et al. [32] evaluated the effects of *Lactobacillus rhamnosus* GG on growth performance and hepatotoxicity in calves fed a single dose of aflatoxin B1 (AFB1). *L. rhamnosus* GG administration reduced the concentrations of free AFB1 and AFM1 in rumen fluid and reduced the release of toxins into plasma and urine. Also, Goel et al. [92] reported that even typical hydrolysable tannins are toxic to both ruminants and monogastrics, particularly when present in the forage of such animals in excessive amounts. However, ruminants have developed an adaptation that allows degradation of tannins to simple isomers via the microbial ecosystem and subsequent excretion in feces.

6. Conclusion

LAB inoculants improve silage quality and reduce DM losses under long-term storage. LAB is one of best organic additives that allow for precise control over undesirable bacterial growth in silage. LAB inoculants utilize water-soluble carbohydrates and complex secondary metabolites in forage crops into organic acids and mineral acids, which effectively control pathogenic growth and enhance the nutritional quality of silage for livestock animals. However, organic acid production and its associated biochemical mechanisms differ depending on the moisture content of forage crops and the storage period. Many studies have been done to investigate the effect of different temperature conditions, harvesting times, moisture levels on forage ensiling quality. In addition, further investigation into the optimal fermentation duration for various kinds of forage crops must be

considered, and the effect of LAB additives on fermentation must be investigated, with the overall goal being to save time and cost and to minimize nutrient losses.

Glossary

Forage

Forage is defined as feed for domestic and/or livestock animals.

Silage

Silage is feedstuff prepared from grass or other green crops (such as corn, wheat, sorghum, legumes, grass and triticale) that has been cut and preserved through fermentation under anaerobic conditions in a silo. Purpose, to provide winter feed for livestock animals.

Ensiling

Ensiling is the name given to the process of preparing and storing forage in order to induce conversion to silage in a packed enclosure (silo).

Silage additives (inoculums)

The purpose of using silage additives is to enhance feed quality by encouraging lactic acid fermentation and inhibiting pathogenic microbial growth and by improving its nutritional value. The silage additives may include bacterial cultures, inhibitors of aerobic damage, acids, and nutrients.

Homofermentative and heterofermentative

LAB can be classified into two main groups. Homofermentative LAB catabolize glucose into lactic acid, while heterofermentative LAB convert glucose into ethanol and CO₂ as well as organic acids.

Dry matter

Dry matter refers to foodstuff that remains after removal of water, and the moisture content reflects the amount of water present in the feed ingredient.

Livestock

Livestock commonly refers to animals, such as cattle, pigs, sheep, horses and goats that are kept on a farm for commodity and profit.

Fermented feedstuff

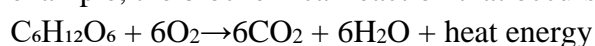
Fermented feedstuff is formed by probiotic microorganisms and mainly encompasses agricultural end products. Fermented feedstuffs contain essential nutrition and gut microbiota because they contain inoculating microorganisms.

Buffering capacity

Silage resistance to pH lowering is called buffering capacity.

Plant respiration

The immediate stage of ensiling involves plant cell respiration and oxidation of WSC (hexose). This is also called the aerobic phase because it can only take place when O₂ is present in a silo. For example, the biochemical reaction that occurs when plant respiration happens in a silo is:



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Conflicts of interest

We declare no conflicts in this work.

References

1. Doonan BM, Kaiser AG, Stanley DF, et al. (2004) Silage in the farming system. In: Kaiser AG, Piltz JW, Burns HM, et al. (Eds), *Successful silage 2nd ed.* Dairy Australia and South Wales Department of Primary Industries, New South Wales, 1–25.
2. Wee YJ, Kim JN, Ryu HW (2006) Biotechnological Production of Lactic Acid and Its Recent Applications. *Food Technol Biotechnol* 44: 163–172.
3. Asmahan AA (2010) Beneficial role of lactic acid bacteria in food preservation and human health: A review. *Res J Microbiol* 5: 1213–1221.
4. Kowalska JD, Nowak A, Śliżewska K, et al. (2020) Anti-Salmonella Potential of New *Lactobacillus* Strains with the Application in the Poultry Industry. *Pol J Microbiol* 69: 5–18.
5. Buxton DR, Muck RE, Harrison JH (2003) Microbiology of ensiling, American Society of Agronomy. *Silage Science and Technology* Madison, Wisconsin USA, 31–93.
6. Guo L, Yao D, Li D, et al. (2019) Effects of lactic acid bacteria isolated from rumen fluid and feces of dairy cows on fermentation quality, microbial community, and in vitro digestibility of alfalfa silage. *Front Microbiol* 10: 2998.
7. Contreras-Govea FE, Muck RE, Broderick GA, et al. (2012) *Lactobacillus plantarum* effects on silage fermentation and in vitro microbial yield. *Anim Feed Sci Technol* 179: 61–68.
8. Pot B, Salvetti E, Mattarelli P, et al. (2019) The potential impact of the *Lactobacillus* name change: The results of an expert meeting organised by the Lactic Acid Bacteria Industrial Platform (LABIP). *Trends Food Sci Technol* 94: 105–113.
9. Holzer M, Mayrhuber E, Danner H, et al. (2003) The role of *Lactobacillus buchneri* in forage preservation. *Trends Biotechnol* 21: 282–287.
10. Tarraran L, Mazzoli R (2018) Alternative strategies for lignocellulose fermentation through lactic acid bacteria: the state of the art and perspectives. *FEMS Microbiol Lett* 365.
11. Hafner SD, Howard C, Muck RE, et al. (2013) Emission of volatile organic compounds from silage: Compounds, sources, and implications. *Atmos Environ* 77: 827–839.
12. Elferink SJWHO, Driehuis F, Gottschal JC, et al. (2000) Silage fermentation processes and their manipulation. *FAO Plant Prod Prot Papers* 17–30.
13. Schnürer J, Magnusson J (2005) Antifungal lactic acid bacteria as biopreservatives. *Trends Food Sci Technol* 16: 70–78.
14. Guan H, Shuai Y, Yan Y, et al. (2020) Microbial community and fermentation dynamics of corn silage prepared with heat-resistant lactic acid bacteria in a hot environment. *Microorganisms* 8: 179.
15. Muck RE, Nadeau EMG, McAllister TA, et al. (2018) Silage review: Recent advances and future uses of silage additives. *J Dairy Sci* 101: 3980–4000.

16. Blajman JE, Paez RB, Vinderola CG, et al. (2018) A meta-analysis on the effectiveness of homofermentative and heterofermentative lactic acid bacteria for corn silage. *J Appl Microbiol* 125: 1655–1669.
17. Oliveira AS, Weinberg ZG, Ogunade IM, et al (2017) Meta-analysis of effects of inoculation with homofermentative and facultative heterofermentative lactic acid bacteria on silage fermentation, aerobic stability, and the performance of dairy cows. *J Dairy Sci* 100: 4587–4603.
18. Bernardes TF, Chizzotti FHM (2012) Technological innovations in silage production and utilization. *Rev Bras Saúde Prod Anim* 13: 629–641.
19. Queiroz OCM, Ogunade IM, Weinberg Z, et al. (2018) Silage review: Foodborne pathogens in silage and their mitigation by silage additives. *J Dairy Sci* 101: 4132–4142.
20. McDonald P, Henderson AR, Heron SJE (1991) *The biochemistry of silage*. Chalcombe publications, 1–45.
21. Xu D, Ding W, Ke W, et al. (2018) Modulation of metabolome and bacterial community in whole crop corn silage by inoculating homofermentative *Lactobacillus plantarum* and heterofermentative *Lactobacillus buchneri*. *Front Microbiol* 9: 3299.
22. Gobbetti M, De Angelis M, Corsetti A, et al. (2005) Biochemistry and physiology of sourdough lactic acid bacteria. *Trends Food Sci Technol* 16: 57–69.
23. Morales-Estrada AI, López-Merino A, Gutierrez-Mendez N, et al. (2016) Partial characterization of bacteriocin produced by halotolerant *Pediococcus acidilactici* Strain QC38 isolated from traditional Cotija Cheese. *Pol J Microbiol* 65: 279–285.
24. Olofsson TC, Vásquez A (2008) Detection and identification of a novel lactic acid bacterial flora within the honey stomach of the honeybee *Apis mellifera*. *Curr Microbiol* 57: 356–363.
25. Papadelli M, Zoumpopoulou G, Georgalaki M, et al. (2015) Evaluation of two lactic acid bacteria starter cultures for the fermentation of natural black table olives (*Olea europaea* L cv Kalamon). *Pol J Microbiol* 64: 265–271.
26. Sornplang P, Piyadeatsoontorn S (2016) Probiotic isolates from unconventional sources: a review. *J Anim Sci Technol* 58: 26.
27. Holzapfel WH (1995) Culture media for non-sporulating Gram-positive food spoilage bacteria. In Corry JEL, Curtis GDW, Baird RM. (Eds), *Progress in Industrial Microbiology*. Elsevier, 89–109.
28. Campana R, van Hemert S, Baffone W (2017) Strain-specific probiotic properties of lactic acid bacteria and their interference with human intestinal pathogens invasion. *Gut Pathog* 9: 12.
29. Wu Z, Xu S, Yun Y, et al. (2019) Effect of 3-phenyllactic acid and 3-phenyllactic acid-producing lactic acid bacteria on the characteristics of alfalfa silage. *Agriculture* 10: 10.
30. Ni K, Wang Y, Li D, et al. (2015) Characterization, identification and application of lactic Acid bacteria isolated from forage paddy rice silage. *PLoS One* 10: e0121967.
31. Borreani G, Tabacco E, Schmidt RJ, et al. (2018) Silage review: Factors affecting dry matter and quality losses in silages. *J Dairy Sci* 101: 3952–3979.
32. Zhang Y, Zhao X, Chen W, et al. (2019) Effects of adding various silage additives to whole corn crops at ensiling on performance, rumen fermentation, and serum physiological characteristics of growing-finishing cattle. *Animals* 9: 695.
33. Soundharajan I, Kim DH, Srisesharam S, et al. (2017) Application of customised bacterial inoculants for grass haylage production and its effectiveness on nutrient composition and fermentation quality of haylage. *3 Biotech* 7: 321.

34. Driehuis F, Elferink SJ, Spoelstra SF (1999) Anaerobic lactic acid degradation during ensilage of whole crop maize inoculated with *Lactobacillus buchneri* inhibits yeast growth and improves aerobic stability. *J Appl Microbiol* 87: 583–594.
35. Muck R (2013) Recent advances in silage microbiology. *Agr Food Sci* 22: 3–15.
36. Cai Y, Benno Y, Ogawa M, et al. (1998) Influence of *Lactobacillus* spp. from an inoculant and of *Weissella* and *Leuconostoc* spp. from forage crops on silage fermentation. *Appl Environ Microbiol* 64: 2982–2987.
37. Ellis JL, Hindrichsen IK, Klop G, et al. (2016) Effects of lactic acid bacteria silage inoculation on methane emission and productivity of Holstein Friesian dairy cattle. *J Dairy Sci* 99: 7159–7174.
38. Ni K, Wang F, Zhu B, et al. (2017) Effects of lactic acid bacteria and molasses additives on the microbial community and fermentation quality of soybean silage. *Bioresour Technol* 238: 706–715.
39. Zhao J, Dong Z, Li J, et al. (2018) Ensiling as pretreatment of rice straw: The effect of hemicellulase and *Lactobacillus plantarum* on hemicellulose degradation and cellulose conversion. *Bioresour Technol* 266: 158–165.
40. Haag NL, Nagele HJ, Fritz T, et al. (2015) Effects of ensiling treatments on lactic acid production and supplementary methane formation of maize and amaranth--An advanced green biorefining approach. *Bioresour Technol* 178: 217–225.
41. Yang L, Yuan X, Li J, et al. (2019) Dynamics of microbial community and fermentation quality during ensiling of sterile and nonsterile alfalfa with or without *Lactobacillus plantarum* inoculant. *Bioresour Technol* 275: 280–287.
42. Bao W, Mi Z, Xu H, et al. (2016) Assessing quality of *Medicago sativa* silage by monitoring bacterial composition with single molecule, real-time sequencing technology and various physiological parameters. *Sci Rep* 6: 28358.
43. Adetoye A, Pinloche E, Adeniyi BA, et al. (2018) Characterization and anti-*Salmonella* activities of lactic acid bacteria isolated from cattle faeces. *BMC Microbiol* 18: 96.
44. Xu Z, He H, Zhang S, et al. (2017) Characterization of feruloyl esterases produced by the four *Lactobacillus* Species: *L. amylovorus*, *L. acidophilus*, *L. farciminis* and *L. fermentum*, isolated from ensiled corn stover. *Front Microbiol* 8: 941.
45. Duniere L, Gleizal A, Chaucheyras-Durand F, et al. (2011) Fate of *Escherichia coli* O26 in corn silage experimentally contaminated at ensiling, at silo opening, or after aerobic exposure, and protective effect of various bacterial inoculants. *Appl Environ Microbiol* 77: 8696–8704.
46. Li M, Shan G, Zhou H, et al. (2017) CO₂ production, dissolution and pressure dynamics during silage production: multi-sensor-based insight into parameter interactions. *Sci Rep* 7: 14721.
47. Mora-Villalobos JA, Montero-Zamora J, Barboza N, et al (2020) Multi-Product lactic acid bacteria fermentations: A review. *Fermentation* 6: 23.
48. Su R, Ni K, Wang T, et al. (2019) Effects of ferulic acid esterase-producing *Lactobacillus fermentum* and cellulase additives on the fermentation quality and microbial community of alfalfa silage. *Peer J* 7: e7712.
49. Buxton DR, Muck RE, Harrison JH (2003) Preharvest plant factors affecting ensiling. *Silage Sci Technol* 42: 199–250.
50. Axelsson L (1998) Lactic acid bacteria: Classification and physiology. In: Salminen S, von Wright A, Ouwehand A. (Eds), *Lactic acid bacteria*. Marcel Dekker Inc, New York, 1–67.
51. Rooke JA, Hatfield RD (2003) *Biochemistry of ensiling*. USDA-ARS/UNL Faculty, 1399.

52. Tanizawa Y, Tohno M, Kaminuma E, et al. (2015) Complete genome sequence and analysis of *Lactobacillus hokkaidonensis* LOOC260 (T), a psychrotrophic lactic acid bacterium isolated from silage. *BMC Genomics* 16: 240.
53. Liu J, Wang Q, Zou H, et al. (2013) Glucose metabolic flux distribution of *Lactobacillus amylophilus* during lactic acid production using kitchen waste saccharified solution. *Microb Biotechnol* 6: 685–693,
54. Wolfe AJ (2015) Glycolysis for Microbiome Generation. *Microbiol Spectr* 3: 10.1128.
55. Melchiorson CR, Jensen NBS, Christensen B, et al. (2001) Dynamics of pyruvate metabolism in *Lactococcus Lactis*. *Biotechnol Bioeng* 74: 271–279.
56. Fushinobu S (2010) Unique sugar metabolic pathways of bifidobacteria. *Biosci Biotechnol Biochem* 74: 2374–2384,
57. Macfarlane GT, Macfarlane S (2012) Bacteria, colonic fermentation, and gastrointestinal health. *JAOAC Int* 95: 50–60.
58. Zaunmuller T, Eichert M, Richter H, et al. (2006) Variations in the energy metabolism of biotechnologically relevant heterofermentative lactic acid bacteria during growth on sugars and organic acids. *Appl Microbiol Biotechnol* 72: 421–429.
59. Iskandar CF, Cailliez-Grimal C, Borges F, et al. (2019) Review of lactose and galactose metabolism in Lactic Acid Bacteria dedicated to expert genomic annotation. *Trends Food Sci Technol* 88: 121–132.
60. Jones EC, Barnes RJ (1967) Non-volatile organic acids of grasses. *J Sci. Food Agric* 18: 321–324.
61. Kung Jr L, Shaver RD, Grant RJ, et al. (2018) Silage review: Interpretation of chemical, microbial, and organoleptic components of silages. *J Dairy Sci* 101: 4020–4033.
62. Milanovic V, Osimani A, Garofalo C, et al. (2020) Selection of cereal-sourced lactic acid bacteria as candidate starters for the baking industry. *PLoS One* 15: e0236190.
63. Vieco-Saiz N, Belguesmia Y, Raspoet R, et al. (2019) Benefits and inputs from lactic acid bacteria and their bacteriocins as alternatives to antibiotic growth promoters during food-animal production. *Front Microbiol* 10: 57.
64. Adebayo-Tayo B, Fashogbon R (2020) *In vitro* antioxidant, antibacterial, in vivo immunomodulatory, antitumor and hematological potential of exopolysaccharide produced by wild type and mutant *Lactobacillus delbureckii* subsp. bulgaricus. *Heliyon* 6: e03268.
65. Hellal A, Amrouche L, Ferhat Z, et al. (2011) Characterization of bacteriocin from *Lactococcus* isolated from traditional Algerian dairy products. *Ann Microbiol* 62: 177–185.
66. Li D, Ni K, Pang H, et al. (2015) Identification and antimicrobial activity detection of lactic acid bacteria isolated from corn stover silage. *Asian-Australas J Anim Sci* 28: 620–631.
67. Byrne CM, Kiely PO, Bolton DJ, et al. (2002) Fate of *Escherichia coli* O157: H7 during Silage Fermentation. *J Food Prot* 65: 1854–1860.
68. Pedroso AF, Adesogan AT, Queiroz OCM, et al. (2010) Control of *Escherichia coli* O157:H7 in corn silage with or without various inoculants: efficacy and mode of action. *J Dairy Sci* 93: 1098–1104.
69. Bjornsdottir K, Breidt Jr F, McFeeters RF (2006) Protective effects of organic acids on survival of *Escherichia coli* O157:H7 in acidic environments. *Appl Environ Microbiol* 72: 660–664.
70. Ogunade IM, Jiang Y, Kim DH, et al. (2017) Fate of *Escherichia coli* O157:H7 and bacterial diversity in corn silage contaminated with the pathogen and treated with chemical or microbial additives. *J Dairy Sci* 100: 1780–1794.

71. Amado IR, Fuciños C, Fajardo P, et al. (2012) Evaluation of two bacteriocin-producing probiotic lactic acid bacteria as inoculants for controlling *Listeria monocytogenes* in grass and maize silages. *Anim Feed Sci Technol* 175: 137–149.
72. Amado IR, Fucinos C, Fajardo P, et al. (2016) Pediocin SA-1: A selective bacteriocin for controlling *Listeria monocytogenes* in maize silages. *J Dairy Sci* 99: 8070–8080.
73. Gavrilova E, Anisimova E, Gabdelkhadieva A, et al. (2019) Newly isolated lactic acid bacteria from silage targeting biofilms of foodborne pathogens during milk fermentation. *BMC Microbiol* 19: 248.
74. Wang X, Wang W, Lv H, et al. (2020) Probiotic potential and wide-spectrum antimicrobial activity of lactic acid bacteria isolated from infant feces. *Probiotics & Antimicro Prot* DOI: 10.1007/s12602-020-09658-3.
75. Ogunade IM, Kim DH, Jiang Y, et al. (2016) Control of *Escherichia coli* O157:H7 in contaminated alfalfa silage: Effects of silage additives. *J Dairy Sci* 99: 4427–4436.
76. Davidson PM, Taylor TM, Schmidt SE. Chemical Preservatives and natural antimicrobial compounds. *Food Microbiology: Fundamentals and Frontiers 4th Ed*, 765–801.
77. Santos AO, Avila CLS, Schwan RF (2013) Selection of tropical lactic acid bacteria for enhancing the quality of maize silage. *J Dairy Sci* 96: 7777–7789.
78. Arques JL, Rodriguez E, Langa S, et al. (2015) Antimicrobial activity of lactic acid bacteria in dairy products and gut: effect on pathogens. *Biomed Res Int* 2015: 584183.
79. Broberg A, Jacobsson K, Strom K, et al. (2007) Metabolite profiles of lactic acid bacteria in grass silage. *Appl Environ Microbiol* 73: 5547–5552.
80. Wilkinson JM, Rinne M (2018) Highlights of progress in silage conservation and future perspectives. *Grass Forage Sci* 73: 40–52.
81. Miron J, Zuckerman E, Adin G, et al. (2007) Comparison of two forage sorghum varieties with corn and the effect of feeding their silages on eating behavior and lactation performance of dairy cows. *Anim Feed Sci Technol* 139: 23–39.
82. Da Silva TC, Da Silva LD, Santos EM, et al. (2017) Importance of the fermentation to produce high-quality silage. *Fermentation Processes* 1–20.
83. Doi K, Nishizaki Y, Kimura H, et al. (2013) Identification of thermo tolerant lactic acid bacteria isolated from silage prepared in the hot and humid climate of Southwestern Japan. *SpringerPlus* 2: 485.
84. Pinho RMA, Santos EM, Pinto de Carvalho GG, et al. (2013) Microbial and fermentation profiles, losses and chemical composition of silages of buffel grass harvested at different cutting heights. *Rev Bras Zootec* 42: 850–856.
85. Khota W, Pholsen S, Higgs D, et al. (2016) Natural lactic acid bacteria population of tropical grasses and their fermentation factor analysis of silage prepared with cellulase and inoculant. *J Dairy Sci* 99: 9768–9781.
86. Ennahar S, Cai Y, Fujita Y (2003) Phylogenetic diversity of lactic acid bacteria associated with paddy rice silage as determined by 16S ribosomal DNA analysis. *Appl Environ Microbiol* 69: 444–451.
87. Tamang JP, Cotter PD, Endo A, et al. (2020) Fermented foods in a global age: East meets West. *Compr Rev Food Sci Food saf* 19: 184–217.
88. Rezac S, Kok CR, Heermann M, Hutkins R (2018) Fermented foods as a dietary source of live organisms. *Front Microbiol* 9: 1785.

89. Andlauer W, Furst P (2003) Absorption and metabolism of phenolic phytochemicals in the isolated rat small intestine with special reference to isoflavones and anthocyanins—A review. *Polish J Food Nutri Sci* 12: 117–127.
90. Babaeinasab Y, Rouzbehan Y, Fazaeli H, et al. (2015) Chemical composition, silage fermentation characteristics, and *in vitro* ruminal fermentation parameters of potato-wheat straw silage treated with molasses and lactic acid bacteria and corn silage. *J Anim Sci* 93: 4377–4386.
91. Zhang LY, Liu S, Zhao XJ, et al. (2019) *Lactobacillus rhamnosus* GG modulates gastrointestinal absorption, excretion patterns, and toxicity in Holstein calves fed a single dose of aflatoxin B1. *J Dairy Sci* 102: 1330–1340.
92. Goel G, Raghav M, Beniwal V, et al. (2015) Anaerobic degradation of tannins in *Acacia nilotica* pods by *Enterococcus faecalis* in co-culture with ruminal microbiota. *J Gen Appl Microbiol* 61: 31–33.
93. Aggoun M, Boussada A, Arhab R, et al. (2017) Effect of natural bioflavonoid on *in vitro* ruminal microbiota activity in sheep rumen liquor. *J BioSci Biotechnol* 6: 31–35.
94. Zhang F, Wang X, Lu W, et al. (2019) Improved quality of corn silage when combining cellulose-decomposing bacteria and *Lactobacillus buchneri* during silage fermentation. *Biomed Res Int* 2019: 4361358.
95. Sun L, Wang Z, Gentu G, et al. (2019) Changes in microbial population and chemical composition of corn stover during field exposure and effects on silage fermentation and *in vitro* digestibility. *Asian-Australas J Anim Sci* 32: 815–825.



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